Accepted Manuscript

Title: The expression of microRNAs and exposure to environmental contaminants related to human health

Authors: Maria Rosaria Tumolo, Alessandra Panico, Antonella De Donno, Pierpaolo Mincarone, Carlo Giacomo Leo, Roberto Guarino, Francesco Bagordo, Francesca Serio, Adele Idolo, Tiziana Grassi & Saverio Sabina



Received 24 Jan 2020, Accepted 14 Apr 2020, Published online: 12 May 2020

https://doi.org/10.1080/09603123.2020.1757043

International Journal of Environmental Health Research

Please cite this article as: Maria Rosaria Tumolo, Alessandra Panico, Antonella De Donno, Pierpaolo Mincarone, Carlo Giacomo Leo, Roberto Guarino, Francesco Bagordo, Francesca Serio, Adele Idolo, Tiziana Grassi & Saverio Sabina (2022) The expression of microRNAs and exposure to environmental contaminants related to human health: a review, International Journal of Environmental Health Research, 32:2, 332-354, DOI: 10.1080/09603123.2020.1757043.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



The expression of microRNAs and exposure to environmental contaminants related to human health: a review

Journal:	International Journal Of Environmental Health Research
Manuscript ID	CIJE-2020-0035.R1
Manuscript Type:	Review Article
Keywords:	microRNAs, pathway, environmental pollutants, health

SCHOLARONE[™] Manuscripts

2	
3	1
4	
5	2
6 7	
7 8	
9	3
10	5
11	Λ
12	4
13	E
14	5
15	c
16	6
1/ 10	_
10	/
20	-
21	8
22	
23	9
24	
25	10
26	
27 20	11
20 29	
30	12
31	
32	13
33	
34	14
35	
36	15
3/ 20	_0
20 20	16
40	10
41	17
42	1
43	10
44	10
45	10
46	19
4/	
48 40	20
49 50	
51	21
52	
53	22
54	
55	23
56	
57	24
58 50	
22	h-

The expression of microRNAs and exposure to environmental contaminants related to human health: a review

Abstract

Environmental contaminants exposure may lead to detrimental changes to the microRNAs (miRNAs) expression resulting in several health effects. miRNAs, small non-coding RNAs that regulate gene expression, have multiple transcript targets and thereby regulate several signaling molecules. Even a minor alteration in the abundance of one miRNA can have deep effects on global gene expression. Altered patterns of miRNAs can be responsible for changes linked to various health outcomes, suggesting that specific miRNAs are activated in pathophysiological processes. In this review, we provide an overview of studies investigating the impact of air pollution, organic chemicals, and heavy metals on miRNA expression and the potential biologic effects on humans.

Keywords: microRNAs, pathway, environmental pollutants, health

Abbreviations: AHRR, aryl-hydrocarbon receptor repressor; AHR, aryl-hydrocarbon receptor; As,
arsenic; BCL2, B-cell lymphoma 2; BCL2L11, B-cell lymphoma 2 like 11; BCL6, B-cell lymphoma
6; BPA, bisphenol A; CVD, cardiovascular diseases; CD40, cluster of differentiation 40; CCND1,
Cyclin D1; CDKN1A, cyclin-dependent kinase inhibitor 1A; COL1A2, eollagen type I alpha 2 chain;
Cr, chromium; CSF1, colony stimulating factor 1; CTBP1, C-terminal binding protein 1; CXCL12,
C-X-C motif chemokine ligand 12; CYP3A4, eytochrome P450 family 3 subfamily A member 4;
CYP2E1, cytochrome P450 family 2 subfamily E member ; DAZAP1, deleted in azoospermia
associated protein 1; DEP, diesel exhaust particles; EGFR, epidermal growth factor receptor; eNOS,
endothelial nitric oxide synthase; ERB1, eukaryotic ribosome biogenesis protein; EVs, extracellular
vesicles; FAK, focal adhesion kinase; FAS, fas cell surface death receptor; FOXO4, forkhead box
O4; GSTP1, glutathione-S-transferase pi 1; HbA1c, glycated hemoglobin; Hg, mercury; HLA-A,

human leukocyte antigen A; HMGB1/AGER, high mobility group protein B-box 1/advanced glycosylation end-product-specific receptor; ICAM-1, intercellular adhesion molecule 1; IFNAR2, interferon alpha receptor subunit 2; IL-6, interleukin-6; IRAK1, interleukin 1 receptor associated kinase 1; JAK/STAT, janus kinase/signal transducers and activators of transcription; LRRK2, leucine-rich repeat kinase 2; MAPK, mitogen-activated protein kinase; MEF2C, myocyte enhancer factor 2C; miRNAs, microRNAs; MVs, microvesicles; NCDs, noncommunicable diseases; NFAT, nuclear factor of activated T cells; NFkB, nuclear factor kappa B; NRF2, nuclear factor, erythroidderived 2; NFE2L2, nuclear factor, erythroid 2 like 2 NGF, nerve growth factor; NRG3, neuregulin 3; O₃, ozone; OP, organophosphorus pesticides; PAHs, polycyclic aromatic hydrocarbons; Pb, lead; PCBs, polychlorinated biphenyls; PDCD4, programmed cell death 4; PDGFB, platelet derived growth factor subunit beta; PDGFR, platelet derived growth factor receptor; PI3K/Akt, phosphoinositide-3-kinase/protein kinase B; PKA, protein kinase A; PM, particulate matter; PRKCQ, protein kinase C theta; PTEN, phosphatase and tensin homolog; PTGES3, prostaglandin E synthase 3; SLAM-SAP, signaling lymphocytic activation molecule-associated protein; SORT1, sortilin 1; TGFβ, transforming growth factor-β; TFIIH, transcription factor II human; TLR, toll-like receptor; TNF, tumor necrosis factors; TRAF16, tumor necrosis factors-receptor associated factors 16; TRAP, traffic-related air pollution; TREM1, triggering receptor expressed on myeloid cells 1; TRIAP1, TP53 regulated inhibitor of apoptosis 1; VCAM-1, vascular cell adhesion molecule 1; VEGFA, vascular endothelial growth factor A; XRCC2, X-ray repair cross complementing 2; YBX2, Y-box-binding protein 2; ZEB1, zinc finger E-box-binding homeobox 1; ZEB2, zinc finger E-box-binding homeobox 2; 8-OH-dG, 8-hydroxy-guanine.

48 Introduction

Exposure to environmental contaminants, including air pollution, organic chemicals, and heavy metals, is a global public health problem associated with adverse health effects (Humphrey et al. 2019). Some sub-cellular effects caused by environmental factors have been investigated both in vitro (Bonetta et al. 2019) and in vivo (Domingues et al. 2018; Panico et al. 2020). Recent evidence suggests that the exposure to toxic compounds influences microRNAs (miRNAs) expression, which contributes to disease development later in life (Miguel et al. 2018).

miRNAs are a class of short non-coding RNA with 18-25 nucleotides in length (Popovic et al. 2013) that play an active role in epigenetic regulation of gene expression, and are also involved in posttranscriptional gene silencing. They have been detected not only intracellularly, but also in extracellular human body fluids, such as serum/plasma, saliva, urine, etc. (Gallo et al. 2012). Despite is the presence of high extracellular RNase activity, miRNAs are highly stable in extracellular area since packaged in apoptotic bodies, microvesicles (MVs), or high density lipoprotein particles (Turchinovich et al. 2012). Therefore,

miRNAs regulate many aspects of biology, including developmental timings, cell differentiation, intercellular communication, embryogenesis, metabolism, organogenesis, and apoptosis (Turchinovich et al. 2016). Through multiple transcript targets, miRNAs regulate signaling molecules or pathways, and they can even be transcriptional targets, providing a mechanism for dysregulation of genes by activation of transcription factors (Hoesel & Schmid 2013). Even a minor alteration to the abundance of one miRNA can have deep effects on global gene expression (Humphrey et al. 2019).

Altered patterns of miRNAs can be responsible for changes linked to various health outcomes,
suggesting that specific miRNAs are activated in pathophysiological processes (Ardekani & Naeini
2010).

Recently, many investigations have examined the relationship between environmental factors and
 miRNAs expression, identifying several chemical contaminants that dysregulated this class of

molecules (Vrijens et al. 2015). As such, we need to understand the biological process underlying miRNA alteration in response to environmental contaminants in order to explore their potential as biomarkers in the management of noncommunicable diseases (NCDs) linked to environmental exposure. This narrative review provides an overview of studies in humans investigating the impact of environmental factors, that is air pollution, organic chemicals, and heavy metals, on miRNA expression, also considering the potential biological mechanism that may lead to pathological condition. -and the potential biologic effects on humans.

miRNAs affected by air pollution

Particulate Matter (PM). Particulate matter consists of a mixture of airborne particles originated from natural sources and anthropogenic activities. Data show that acute PM exposure leads to adverse health effects through oxidative stress generation and inflammation induction, with the highest impacts on cancer and cardiovascular diseases (CVD) (Martinelli et al. 2013).

 Table 1 summarizes the studies on the expression of miRNAs in response to air pollution, also
 including sample types, study designs, and methodologies for miRNA detection and for analysis of

 their targets. miRNA nomenclature was reported adopting the most recent official version used in

 miRbase (Griffiths-Jones et al. 2008; Kozomara et al. 2019).

Insert Table 1 about here.

PM concentration has been linked to several clinical manifestations of CVD, which, in turn, lead to altered miRNAs expression (Vrijens et al. 2015). Louwies and colleagues conducted a study that investigated relationship between air pollutants and miRNAs expression in combination with microvascular responses to PM. They measured by real-time quantitative reverse-transcriptase polymerase chain reaction (qRT-PCR) the levels of three candidate miRNAs in the blood of 50 healthy adults. Of the three miRNAs examined, only miR-21-5p and miR-222-3p were negatively associated with PM₁₀₂-A retinal microvascular response to variation in PM could be explained by dysregulated miRNAs, miR-21-5p and miR-222-3p, in blood of healthy adults. Bioinformatic

analysis revealed gene targets of these miRNAs that were associated with These miRNAs are involved in inflammatory and oxidative stress-pathways, that is phosphatase and tensin homolog (PTEN) signaling pathway and high-mobility group protein B (HMGB). They also assessed the width of retinal blood vessels by eye fundus photos. Thus, a retinal microvascular response to variation in PM could be explained by these dysregulated miRNAs, that have a role in these pathways (Louwies et al. 2016). Their effect on the high mobility group box 1 (HMGB1)/advanced glycosylation endproduct-specific receptor (AGER) signaling pathway leads to enhanced production of proinflammatory cytokines, adhesion molecules, and coagulation factors . Additionally, the downregulation of these miRNAs in the phosphatase and tensin homolog (PTEN) signaling pathway upregulates PTEN expression that in turn either inhibits the endothelial nitric oxide synthase (eNOS) pathway or increases intercellular adhesion molecule 1 (ICAM-1) expression resulting in retinal vessels narrowing, or both (Louwies et al., 2016).

The same miRNAs were significantly increased in blood of steel plant workers collected after three workdays exposed to high PM. For these specific miRNAs, miR-222 is associated with mitogenactivated protein kinase (MAPK) signaling and nerve growth factor (NGF) signaling, while miR-21 is associated with PTEN, MAPK, and phosphoinositide 3 kinase (PI3K)/protein kinase B (Akt) signaling (Bollati et al. 2010). In other studies of PM exposure implicated in CVD, microvesicleassociated miRNA expression (miR-128 and miR-302c) was overexpressed after three days of workplace PM exposure in healthy electric steel plant facility workers. In this case study, nuclear factor kappa B (NFkB), commonly associated with a prototypical pro-inflammatory signaling pathway, was found to be a central molecule in the networks of both miRNAs (Bollati et al. 2015). Air pollution could alter intercellular communication by extracellular vesicles (EVs), such as MVs, that can transfer miRNAs between tissues (Pavanello et al. 2016), Rodosthenous et al. investigated relationship between short-, intermediate-, and long-term exposures to PM and levels of EV-miRNAs in a cohort of healthy adults. The profile of 800 miRNAs was screened using Nanostring Technologies' nCounter[®] assay that revealed an association between long-term ambient PM exposure

4 5

6 7

8 9

126 is also associated with increased disrupting background levels of somesixteen EV-extracellular 127 vesicle (EV) miRNAs circulating in serum; in silico analysis showed that their which target genes (for example interleukin 6 - IL-6, C-X-C motif chemokine ligand 12 - CXCL12, vascular cell 128 10129 adhesion molecule 1 - VCAM-1, cluster of differentiation 40 - CD40, platelet derived growth factor 11 ¹² 130 subunit beta - PDGFB, etc.) are linked relevant to CVD-related pathways, such as oxidative stress, 13 14 ₁₅ 1**3**1 inflammatory response, ion and atherosclerosis, toll-like receptor (TLR) etc. (Rodosthenous et al. 16 17 1 32 2016). Among these miRNAs, miR-146a-5p is an important regulator of pro-inflammatory cytokines 18 19 183 such as interleukin-6 (IL 6) via the NFkB pathway; moreover, this miRNA targets tumor necrosis 20 21 --22 134 factors (TNF)-receptor associated factors 6 (TRAF6) and interleukin 1 receptor associated kinase 1 23 24 1 35 (IRAK1), proteins that are part of the cluster of differentiation 40 (CD40) signaling pathway, which 25 ²⁶ 186 serves an important role in cellular communication during inflammatory responses ; miR-23a-3p 27 29 137 28 interacts with C-X-C motif chemokine ligand 12 (CXCL12), which has been reported to regulate 30 31 138 inflammation; miR-126a-3p is associated with vascular cell adhesion molecule 1 (VCAM-1) 32 33 1 3 9 expressed by endothelial cells in response to inflammation and plays a critical role in recruiting 34 ³⁵ 140 leukocytes with implications for vascular inflammation and atherosclerosis ; miR-150-5p interacts 36 37 with platelet derived growth factor subunit beta (PDGFB), a protein expressed by smooth muscle ₃₈ 141 39 40 1 4 2 endothelial and epithelial cells that plays a central role in cell proliferation and has been implicated 41 ⁴² 143 in inflammatory responses and atherosclerosis; let-7g and miR-130-3p target other cytokines, such 43 44 45¹⁴⁴ as colony stimulating factor 1 (CSF1) and collagen type I alpha 2 chain (COL1A2), mediating 46 communication between immune cells in the inflammatory process (Rodosthenous et al. 2016). 47 145 48 ⁴⁹ 146 Prenatal exposure to PM has been associated with fetal growth restriction, low birth weight, preterm 50 51 1<mark>47</mark> birth (Gianicolo et al. 2012; Gianicolo et al. 2014), and cause adverse health outcomes in adulthood. 53 54 148 Maternal exposure to air pollution has been suggested to adversely affect pregnancy by inducing 55 56 149 oxidative stress and inflammation, which may result in impaired placental angiogenesis (van den 57 ⁵⁸ 150 Hooven et al. 2012). In utero, PM exposure during different periods of gestation affects miRNAs 59 60 151 levels. In a recent study of Tsamou and colleagues, miRNA expression was analyzed by qRT-PCR

in 210 placental tissues from mother-newborn pairs. The results indicated that miR-21-5p, miR-146a-5p, and miR-222-3p were inversely associated with PM exposures during the second trimester of pregnancy, while placental expression of miR-20a-5p and miR-21-5p was positively associated with first trimester exposure. In silico prediction tools showed that aA common putative target of these miRNAs is the tumor suppressor PTEN, involved in many key cellular processes by negatively regulating PI3K/Akt pathway related to cell survival, cell cycle, angiogenesis, and metabolism (Kitagishi & Matsuda 2013) that was validated measuring its expression by qRT-PCR in a subset of the same cohort. miR-21-5p, miR-20a-5p, and miR-222-3p were inversely correlated with PTEN, confirming the miRNA-PTEN co-expression in placental tissue (Tsamou et al. 2018). Metal rich-PM. PM also contains carcinogenic and toxic heavy metals, which can induce epigenetic modifications. Metals are one of the most important factors causing cardiovascular and respiratory diseases due to systemic activation of pro-inflammatory pathways occurring after exposure (Mercorio et al. 2017). A cohort study conducted by Bollati et al. analyzed blood miRNA profile of steel plant workers; the samples were collected at the beginning and at the end of the working week. The analysis through qRT-PCR showed that miR-21-5p and miR-222-3p were significantly increased in leukocytes after three high metal-rich PM exposure workdays. Moreover, miR-21-5p level was positively correlated with 8-hydroxy-guanine (8-OH-dG), indicating a relationship with oxidative stress. Also bioinformatic analysis confirmed that these specific miRNAs are involved in pathways related to oxidative stress and inflammation, such as mitogen-activated protein kinase (MAPK) signaling, chemochine signaling pathway, transforming growth factor- β (TGF β), TLR, and other signaling pathways related to general function (focal adhesion, apoptosis, etc.) (Bollati et al. 2010). Afterwards, another study done again by Bollati et al. investigated whether PM and metal-rich PM alter MVs signaling. The results showed that the expression of miR-128 and miR-302c was significantly overexpressed after three days of workplace PM exposure compared with the beginning of the 177 working week. The authors suggested that these pollutants could affect MVs-associated miRNAs,

1

representing a novel mechanism of air pollution toxicity. Bioinformatic approaches revealed that nuclear factor kappa B (NFkB) was found to be a central molecule in the networks of both miRNAs, which also regulated gene expression linked with CVD (Bollati et al. 2015). Additionally, the effect of metal-rich PM was evaluated in a subset of the same study population identifying four PMsensitive miRNAs (miR-29a-3p, miR-146a-5p, miR-421, and let-7g-5p)

The effect of metal-rich PM in foundry workers identified four PM-sensitive miRNAs (miR-29a, miR-146a, miR-421, and let-7g) that were differentially expressed in post-exposure compared with baseline samples and seem to be implicated in the inflammatory processes. A quantitative PCR was performed to examine mRNA expression of eighteen predicted target genes of the dysregulated miRNAs. Notably, miR-29a-3p negatively correlates directly interacts with PTEN mRNA, let-7g-5p interacts with NFkB and transforming growth factor β (TGF β), miR-146a-5p targets as many as eight mRNAs, from which a complex network of interactions originates that converge to ultimately influence TGF β mRNA expression and positively with endothelial nitric oxide synthase (eNOS) and platelet derived growth factor receptor (PDGFR). All of these interactions promote a proinflammatory response leading potentially to several diseases, including CVD and respiratory illness (Motta et al. 2013).

The environmental impact on spermatogenesis is also an important issue, influencing male reproductive health. Li et al. assessed expression of miRNAs -In spermatozoa of men living in areas polluted from electronic waste compared with men living in a non-polluted site. Microarray analysis identified 182 dysregulated miRNAs and only eleven of these were further validated by qRT-PCR. This analysis showed that miRNA level expression was differentially altered (miR-10b-5p, miR-33b-5p, miR-106a-5p, miR-155-5p, miR-183-5p, miR-205-5p, miR-208a, miR-222-3p, miR-223-3p were up-regulated while miR-363-3p and let-7d-5p were down-regulated) in the polluted group than the control one-from men living in a non-polluted site. Cluster aAnalyses_-showed-displayed that miR-10b-5p and let-7d-5p were linked to spermatogenesis, leading to possible male reproductive disorder,

and that the most significant signal was for the Notch signaling pathway (Yan Li et al. 2012) which
 plays a major role in the regulation of embryonic development and promotes proliferative signaling
 during neurogenesis (Pierfelice et al. 2011). and that disruption of miR-10b and let-7d effect
 spermatogenesis, leading to possible male reproductive disorder (Yan Li et al. 2012).

208 Diesel exhaust particles (DEP). Long-term exposures to DEP, the main source of genotoxic 209 substances in urban areas, can contribute to chronic outcomes, such as cardiovascular, metabolic, and 210 respiratory diseases (Rider & Carlsten 2019).

Rider et al. analyzed the bronchial brushings miRNA expression levels of fifteen subjects with atopy
carrying out a double-blinded crossover study. The objective of their study was to determine whether
exposure to allergen, or DEP, or coexposures modulated miRNA profile that was examined applying
Nanostring Technologies' nCounter[®] assay. They found a weak relationship between miRNAs and
DEP exposure, but miR-183-5p, miR-324-5p, miR-132-3p, and miR-331-3p were significantly
associated with allergen exposure. Bioinformatic analysis showed a negative correlation both for
miR-132-3p with cyclin-dependent kinase inhibitor 1A (CDKN1A) and miR-183-5p with human
leukocyte antigen A (HLA-A) (Rider et al. 2016).

After DEP exposure, miR-132-3p was up-regulated and miR-183-5p was down-regulated, inducing asthma and other respiratory diseases. miR-183-5p, which was significantly repressed following allergen exposure, targets forkhead box O1 (FOXO1), a transcription factor crucial in responding to oxidative stress. miR-132-3p works as a repressor of cyclin-dependent kinase inhibitor 1A (CDKN1A). It encodes its inhibitor (p21), which regulates cell proliferation by binding to cyclin-CDK complexes. In short, reduced CDKN1A expression could promote dysregulation of cell cycle control, increased epithelial cell apoptosis, and activity of pro-inflammatory pathways (Rider et al. 2016).

Adverse health effects associated with DEP exposure could be mediated in part by oxidative stress.
 Yamamoto and colleagues described an association between miR-144 and oxidative stress. The
 randomized crossover design of the study included thirteen When subjects with mild asthma were

1

exposed to DEP. miRNA profiling using Nanostring nCounter[®] assay showed increased levels of miR-21-5p, miR-30e, miR-215, and miR-144 in their peripheral blood. The validation phase by qRT-PCR confirmed a significant up-regulation of miR-144. To investigate the biological function of miR-144, the authors conducted a PCR analysis that showed a negative association of nuclear factor, erythroid-derived 2 (NRF2) and its downstream antioxidant genes with miR-144. The latter was also positively correlated with 8-OH-dG (Yamamoto et al. 2013)., miR-144 was found upregulated in their peripheral blood_and thus, associated with oxidative stress. This miRNA targets nuclear factor, erythroid2 like 2 (NRF2)FE2L2), is a transcription factor regulating the cellular responses to oxidative stress (Sangokoya et al. 2010).In fact, NFE2L2 that regulates the expression of detoxifying enzymes, determining an adaptive response to oxidant pollutants exposure (Lodovici & Bigagli 2011) (Yamamoto et al. 2013).

Traffic-related air pollution (TRAP). Road transport contributes considerably to air quality problems through vehicle emissions and leads to adverse cardiorespiratory effects including exacerbation of asthma, reduced lung function, myocardial infarction, cardiovascular mortality, and neurodegenerative diseases (Matz et al. 2019).

Krauskopf et al. investigated the relationship between TRAP exposure and miRNAs expression in 24
non-smoking participants using next-generation sequencing technology. In this an randomized
experimental crossover study, twenty-four -comparison of subjects who walked for 2 hours along
Oxford Street in London and then, in a separate session, for other 2 hours through traffic-free Hyde
Park. The plasma miRNA profile of the two different sessions was compared with that of the same
subjects who walked, in a separate session, for 2 hours through traffic free Hyde Park, miRNA
expression pattern showed and decreased levels of miR-27a-5p, miR-133a-3p, miR-145-5p, miR-193b-3p, miR-433-3p, miR-580-3p, miR-6716-3p, and increased levels of miR-1224-5p and miR-3127-5p were observed for the TRAP exposure samples. Further bioinformatic analysis indicated
showed that the potential targets of these miRNAs included were genes involved in CVD, respiratory
diseases, cancer-related pathways (breast cancer, non-small cell lung cancer, etc.), and signaling

pathways such as the PI3K-Akt and p53 (Krauskopf et al. 2018). As an example, miR-145-5p inhibits

growth and migration of breast cancer and it has also been identified to inhibit the proliferation of non-small cell lung cancer cells by targeting the oncogene c-Myc, while miR-1224- 5p, silences leucine-rich repeat kinase 2 (LRRK2), a crucial factor known to be down-regulated during pathogenesis of Parkinson's disease (J.-Q. Li et al. 2014)(Krauskopf et al. 2018). Ozone (03). Ozone is a secondary air pollutant associated with various adverse health effects, predominantly attributable to respiratory diseases (Zhang et al. 2019). Fry et al. analyzed tThe sputum of healthy non- asthmatic subjects collected before and after exposure to O_3 for 2 hours was analyzed using a microarray approach. and <u>T</u> the results showed that O_3 significantly increased the levels of ten seven miRNAs (miR-25-3p, miR-132-3p, miR-143-3p, miR-145-5p, miR-199a-3p, miR-199b-5p, miR-222-3p, miR-223-3p, and miR-434-5p, and miR-582-5p) that, according to computational prediction, are involved in inflammation process and immunerelated diseases. In inflammation response, miR-222 is known to be related to neutrophil hyperactivity and granulocyte development targeting myocyte enhancer factor 2C (MEF2C), a transcription factor that promotes myeloid progenitor proliferation, miR-143 plays a role in neutrophil influx, and miR-145-5p is linked to several physiological features of asthma. In immune response, miR-199b-5p directly regulates the nuclear factor of activated T cells (NFAT) pathway, miR-132-3p effects interferon-stimulated gene expression, and miR-434-5p and miR-25-3p regulate immune cell differentiation Moreover, human monocyte-derived macrophages were used in an in vitro model, in order to validate the dysregulation of miR-145-5p and miR-199b-5p and their mRNA targets. Cyclin D1 (CCND1) and v-myc avian myelocytomatosis viral oncogene homolog (MYC) showed a significant decreased expression which demonstrate the induced effect of O₃ on these two miRNAs and their targets (Fry et al. 2014).

miRNAs affected by organic chemicals

1

Polycyclic Aromatic Hydrocarbons (PAHs). Exposure to PAHs, environmental pollutants formed during the incomplete combustion of organic materials, <u>may_generates_various</u> adverse health outcomes, such as respiratory diseases and some cancers (Kim et al. 2013). Moreover, PAHs could be an additional risk factor for impaired vascular health and atherogenic processes that gradually lead to CVD (Kim et al. 2013; Xu et al. 2013). <u>Results from the human studies concerning miRNA alteration after organic chemical exposure are shown in Table 2.</u>

7 <u>Insert Table 2 about here.</u>

Ruiz-Vera and colleagues in their cross-sectional study assessed pPlasma levels of vascular-related miRNAs in women exposed to PAHs via biomass combustion smoke (using wood as a fuel source in their house). After qRT-PCR analysis, they found had higher levels of miR-126a-3p and miR-155-5p in this group compared to than women not exposed to PAHs. Bioinformatic analysis to predict the target genes and pathways of miR-126a-3p and miR-155-5p showed a possible relationship with cardiovascular events, notably in the progression of atherosclerosis; in fact the predicted target genes/pathways are linked to inflammation, vascular endothelial health, and other similar pathways (Ruiz-Vera et al. 2019). These miRNAs are important regulators in the progression of atherosclerosis. miR-126 is involved in angiogenesis via vascular endothelial growth factor (VEGF) pathway (Nicoli et al. 2010) and is associated with the vascular smooth muscle cell turnover related to atherosclerosite plaque thinning (Zhou et al. 2013). miR-155 is classified as a pro inflammatory agent regulating inflammation-related genes and is related to macrophage foam cell formation in atherosclerotic lesions (Nazari-Jahantigh et al. 2015) (Ruiz-Vera et al. 2019).

Additionally, PAHs are metabolically activated to form stable PAH-DNA adducts and cause DNA oxidation. This event may lead to DNA damage, a common cause of cancer (Xue & Warshawsky 2005). In this sense, <u>in the study of Deng et al.</u> exposure to relatively high concentrations of PAHs <u>lead to altered miRNAs expressions in previously healthy coke oven workers relative to control</u> groups. <u>Specifically, As a result of PAHs exposure, only miR-150-5p was up-regulated, while the</u> <u>down-regulation of four miRNAs (miR-24-3p, miR-27a-3p, miR-142-5p, and miR-28-5p) were</u> down-regulated. These four miRNAs regulated genes that could protect against adverse effects of PAH exposure and the increased level of miR-150-5p is linked to a decreased immune response (Deng et al. 2014). miR-24-3p negatively regulates H2AX, which encodes a protein crucial for double-stranded break repair; thus, reduced expression of this gene might increase cellular sensitivity to DNA-damaging agents and genomic instability (Lal et al. 2009). miR-27a-3p can be downregulated by reactive oxygen species and may enhance transcription factor II human (TFIIH), which is involved in DNA repair processes. In this regard, exposure to PAHs results in an increased DNA repair capacity and decreased chromosome damage . miR-142-5p down-regulation can lead to upregulation of signaling lymphocytic activation molecule (SLAM)-associated protein (SAP) and to increase T-cell function and IgG production; it may protect individuals against the deleterious effects of PAHs. The down-regulation of miR-28 may elevate the expression of NFE2L2 and may protect cells from DNA damage. As a result of PAH exposure, only miR-150-5p was up-regulated; it is a key regulator of *c-Myb*, an important gene for immune cell differentiation and activation. miR-150-5p deficiency can lead to an enhanced immune response; therefore, miR-150-5p up-regulation may decrease immune response to PAH exposure and make subjects more susceptible to their dangerous effects (Xiao et al. 2007)(Deng et al. 2014).

Polychlorinated biphenyls (PCBs). Polychlorinated biphenyls <u>PCBsare considered</u> a class of <u>toxic</u> aromatic chemical compounds <u>that bio-accumulate specially in the fatty tissue of humans and animals</u> (Lignell et al. 2016).

<u>Several studies investigated the association between PCBs and miRNAs are linked to increased risk</u>
of multiple types of cancer, including non-Hodgkin lymphoma (Engel et al. 2007; Vrijheid et al.
2016).

<u>Krauskopf et al. reported that PCBs exposure can lead to various type of cancer in humans. In their</u>
 <u>population-based study, PCBs serum levels and other persistent organic pollutants were assessed in</u>
 healthy subjects and <u>microarray analysis identified</u> a total of 93 miRNAs were found significantly
 associated (53 positively and 40 negatively) with <u>PCBs</u> exposure. <u>The miRNA profile integration</u>

with transcriptome profile displayed an interaction with oncogenes such as MYC, CCND1, B-cell lymphoma 2 (BCL2) and vascular endothelial growth factor A (VEGFA). The predicted target genes were related to various types of human cancer and involved in signaling pathways like Wnt, apoptosis, and cell cycle regulation (Krauskopf et al. 2017). Among The most positively correlated miRNAs, namely these, some miRNAs, such as miR-29a, miR-31-5p, miR-34a-5p, miR-152, and miR-193a-3p, were indicated as tumor suppressors miRNAs (Misso et al. 2014; Liang et al. 2015; Yan et al. 2015; Kim et al. 2015; Liu et al. 2016). for example, miR-29a represses a total of eight gene targets, including the lymphoma-related genes, CCND1 and BCL2 (Krauskopf et al. 2017). PCBs exposure may also be attributed to birth defects and embryonic development delays (Vrijheid et al. 2016). Healthy pregnant women living in an area polluted with PCBs who underwent therapeutic abortion due to fetal malformations had PCBs blood concentrations that correlated with miR-191-5p up-regulation compared with women living in a non-polluted area who had a healthy pregnancy. Furthermore, a PCR analysis showed Up-regulation of miR-191 leads to the downregulation of aryl-hydrocarbon receptor repressor (AHRR), C-terminal binding protein 1 (CTBP1) and Fas cell surface death receptor (FAS) in peripheral blood cells (Guida et al. 2013). miR-191-5p has as sequence complementary to the 3'-UTR region of these genes that are involved in pathways related to immune and inflammatory effects which can inducinge oxidative stress, immunotoxicity, and cancer (Pradhan et al. 2011; Tao et al. 2012; Yuan-fang Li et al. 2012). Thus, these changes along with the dangerous effects of PCBs in the CTBP1, FAS and AHRR protein expression can interfere with normal development and health conditions. In the study of Li and colleagues, in order to examine the association between miRNAs and some pollutants, healthy placental tissues were collected from a birth cohort. A positive association between PCBs and miR-1537 expression level was found in placental samples, suggesting that miRNA profiles may signal in utero exposure to environmental chemicals. The target of miR-1537 is N-Myc gene, which regulates cell growth, proliferation, and apoptosis. It encodes MYCN proto-oncogene protein, whose overexpression can lead to tumorigenesis(Li et al. 2015).

Phthalates and phenols. Phthalates and phenols are two classes of potential endocrine disrupting chemicals present in the environment, consumer products, and food, which are mainly associated with adverse female fertility outcomes (Vrijheid et al. 2016). In the study of Martinez et al. Eenvironmental exposure to phthalates and phenols with EV-miRNA profiles was evaluated in follicular fluid of 130 women who provided urine samples during ovarian stimulation. This crosssectional study showed an altered expression of eEight EV-miRNAs (miR-15b-5p, miR-19a-3p, miR-24-3p, miR-125b-5p, let-7c, miR-106b-5p, miR-374a-5, and miR-375) were found associated linked to with phenols and phthalate concentrations. Specifically, miR-15b-5p, miR-19a-3p, miR-24-3p, miR-125b-5p, and let-7c were up-regulated, while miR-106b-5p, miR-374a-5, and miR-375 were down-regulated. In silico analysis revealed that the potential target genes of mMost of these miRNAs might be involved in were associated with follicular development and oocyte maturation and function, highlighting the potential effect of phenols and phthalates exposure on female fertility. Some putative pathways regulated by these miRNAs are TGFβ, phosphoinositide-3-kinase/protein kinase B (PI3K/Akt), forkhead box O (FOXO), MAPK, p53, epidermal growth factor receptor (EGFR), and janus kinase/signal transducers and activators of transcription (JAK/STAT) signaling pathways (Martinez et al. 2019). La Rocca et al. reported that the exposure to phthalates and phenols during pregnancy may influence several biological processes implicated in placental and fetal health. In their study, tThe associations between first-trimester urine concentrations of phenols and phthalates metabolites and expression of candidate miRNAs in placenta showed a down-regulation of miR-185 in response to phthalates exposure and down-regulations of miR-15a-5p and miR-142-3p in response to phenols. These miRNAs, according to in silico analysis, are related with regulate protein serine/threonine kinase Akt activity, a major component of the apoptotic pathway, the insulin like growth factor receptor signaling pathway, metencephalon development, and embryonic epithelial tube formation This indicates that prenatal phthalate and phenol exposure may interfere with several biological processes that have been implicated in placental and fetal healt (LaRocca et al. 2016).

4 5

6 7

8 9

11 12

13 14

16

18

20 21

23

25

27

30

32

34

37

39

41

43 44

46

48

50

51

53

55

57

59 60

Among phthalates and phenols, bisphenol A (BPA) was reported to affect neurological, 384 385 cardiovascular, and metabolic diseases (such as diabetes), cancers, and have harmful consequences 386 for the developing fetus (Ikezuki et al. 2002; Jedeon et al. 2013). A cohort of pregnant women 10 387 exposed to BPA was enrolled in a case-control study carried out by De Felice et al., in order to 388 investigate miRNA changes. In placentas from pregnant women exposed to BPA, Microarray analysis ₁₅ 3<mark>8</mark>9 showed the altered profile of eighteen miRNAs; the verification of their dysregulated expression by 17 390 qRT-PCR revealed a significant overexpression of miR-146a-5p-was overexpressed compared to a ¹⁹ 3<mark>9</mark>1 control group. Functional significance of this miRNA was evaluated applying bioinformatic analysis 22 392 and suggested that This miR-146a-5pNA is probably associated to neural disease genes (interleukin 24 3 9 3 1 receptor associated kinase 1- IRAK1, sortilin 1- SORT1 etc.), endocrine system genes pathway ²⁶ 394 (TP53 regulating kinase), cardiovascular disease genes (ABL2) and cancer-related pathways (EGFR, ²⁸ 29 395 p53, toll-like receptor-TLR) (De Felice et al. 2015).

31 396 Organophosphorus pesticides (OP). Organophosphorus pesticides are heavily used in agriculture, but chronic OP exposure is implicated in many adverse health outcomes, such as neurological and 33 397 ³⁵ 398 respiratory effects (Almeida et al. 2019). In a longitudinal study involving 27 farmworker and non-₃₈ 399 farmworker adults, Weldon et al. examined the effect of pesticide exposure on uUrinary miRNAs 40 4 0 0 expression. The samples were collected during two agricultural seasons (thinning and post-harvest).s ⁴² 401 linked to pesticide exposure from 27 parent/child, farmworker/non-farmworker pairs collected during 45⁴02 two agricultural seasons (thinning and post-harvest) were assessed. They found s Significant differences in the miRNA-profiles of six miRNAs (miR-28-5p, miR-133b, miR-223-3p, miR-517b-47 403 49 404 3p, miR-518d-3p, miR-597) were found in between farmworker and non-farmworker adults, as well 52 405 as between seasons. Six miRNAs (miR-28-5p, miR-133b, miR-223-3p, miR-517b-3p, miR-518d-3p, 54 4**0**6 miR-597), some of which are involved in neurological functions including neurotransmitter activity 56 407 and receptor binding, were positively associated withadults farmworkers status during the post-⁵⁸ 408 harvest season, indicating that they may be novel biomarkers of pesticide exposure and early 409 biological response. Bioinformatic analysis identified that miR-28-5p was has been associated with

acetylcholine binding, acetylcholinesterase, and cholinesterase activity <u>and that miR-517b</u>, <u>miR-518d-5p</u>, and <u>miR-597</u> were associated with target genes involved in neurological functions including
 neurotransmitter activity and receptor binding (Weldon et al. 2016).

miR-223-3p modulates activities of cytochrome P450 family 3 subfamily A member 4 (CYP3A4)
and cytochrome P450 family 2 subfamily E member 1 (CYP2E1) genes, which are responsible for
the breakdown of many toxic environmental chemicals and carcinogens that enter the body, in
addition to basic metabolic reactions such as fatty acid oxidations . miR-133b impairs the expression
of glutathione S- transferase pi 1 (GSTP1), an important factor for fulfilling protective and
detoxifying functions in tumor cells miR-517b, miR-518d-5p, and miR-597 are associated with target
genes involved in neurological functions including neurotransmitter activity and receptor binding
(Weldon et al. 2016).

miRNAs affected by heavy metals

Arsenic (As). Human exposure to high As concentration, a toxic metalloid widely distributed in the environment, is related to many health disorders, mainly CVD and cancer (Vrijheid et al. 2016; Rehman et al. 2018). Many epidemiological studies have shown evidence that exposure to inorganic As could have harmful effects on the cardiovascular system of humans, such as ischemic heart failure, cardiac arrhythmias, and endothelial dysfunction (Stea et al. 2014). Results of the described studies are displayed in Table 3.

29 <u>Insert Table 3 about here.</u>

In a cross-sectional study by Pérez-Vázquez et al. Research has found a significant negative association between urinary As concentration and plasma miR-126-3p levels in Mexican children was found (Pérez-Vázquez et al. 2017). Other studies indicate that miR-126-3p is the most abundant miRNA in endothelial cells that contributes to regulate vascular integrity and developmental angiogenesis besides to be involved in many cardiovascular pathways; this makes miR-126-3p down-

regulation an early biomarker of CVD diseases (Fish et al. 2008; Wei et al. 2013) (Pérez-Vázquez et al. 2017).

Accumulated evidence suggests that the bladder epithelium may be one of the primary targets of Asinduced carcinogenesis. <u>Michailidi et al. analyzed Uurine samples from subjects exposed to different</u> level of As, showing miR-200c-3p and miR-205-5p inversely associated with As exposure <u>compared</u> to unexposed controls. Moreover, the authors validated the expression of these miRNAs in urine samples from patients with urothelial carcinoma in comparison with controls without cancer. The results displayed a low expression of miR-205-5p, suggesting its potential use as biomarker for bladder cancer (Michailidi et al. 2015). Both of these-miR-200c-3p and miR-205-5pNAs have tumor suppressive functions. miR-200c-3p can reverse epithelial mesenchymal transition via regulation of zinc finger E-box-binding homeobox 1 (ZEB1) and ZEB2 (Wellner et al. 2009). <u>Hence, these</u> miRNAsBoth-miR-200 and miR-205 may play a role in the tumor initiation and progression (<u>Michailidi et al. 2015</u>).

A case-control study conducted by Banerjee et al. investigated peripheral blood mononuclear cell (PBMC) miRNA profile of individuals chronically exposed to As through drinking water (experimental group) and in unexposed control. The experimental group was divided into subjects with As induced skin lesions and skin cancer and those without any skin lesions. The Elevels of miR-21-5p, analyzed by qRT-PCR, were up-regulated in individuals with skin lesions exposed to As compared to the control group and were higher in those with skin lesion than the no-skin lesion subgroup. _through drinking water; itTo_our_knowledge, this was one of the few studies that experimentally analyzed, by Western blot analysis, miRNA targets (Banerjee et al. 2017). miR-21-5p has been designated as a miRNA associated with carcinogenic outcomes (Sun et al. 2014) due to its direct genes targets, PTEN and programmed cell death 4 (PDCD4); that were found to be inversely correlated to miR-21-5p expression. Again, as expected, the expression of survival protein increased a neoplastic transformation inhibitor (Banerjee et al. 2017). All these results were also validated through an in vitro experiment that showed similar trends (Banerjee et al. 2017).

In another multi-stage study, Sun et al. assessed the levels of 754 miRNAs in the plasma of subjects exposed to As through coal-burning. The expression of 74 miRNAs was dysregulated by using microarray, and twelve were further analyzed by qRT-PCR. The levels of four miRNAs (miR-21-5p, miR-145, miR-155-5p and miR-191-5p) were higher in Differential expression of plasma miRNAs between people exposed to As through coal burning and compared to a control group,--showed an up-regulation of four miRNAs (miR-21, miR-145, miR-155 and miR-191) which Bioinformatic tools showed that these miRNAs inhibit the target genes of pathways linked to oxidative stress, DNA damage repair, and immune inflammation. miR-21-5p targets were MAPK, JAK/STAT, and chemokine pathways which may be involved in chronic arsenic poisoning. The signaling pathways related to miR-145 target genes were MAPK signaling, associated with oxidative stress, and the FOXO signaling pathway, involved in various types of diseases. miR-155-5p and mir-191-5p are involved in changes in the MAPK, PI3K/Akt, and tumor necrosis factors (TNF) pathways (Sun et al. 2017).

In utero exposures to As can harm the developing fetus, increase risk of spontaneous abortions/disrupt host defenses, and lead to deleterious health outcomes (Farzan et al. 2013), Rager et al. conducted a study in which 40 cord blood samples were selected from mother-newbors pairs from a pregnancy cohort exposed to As. Cord blood samples from prenatal exposure to As-Microarray analysis revealedsulted in an increased expression of twelve miRNAs (miR-16-5p, miR-17-5p, miR-20a-5p, miR-20b-5p, miR-26b-5p, miR-96-5p, miR-98-5p, miR-107, miR-126-3p, miR-195-5p, miR-454-3p, let-7a-5p) associated with As exposure. Then, qRT-PCR was performed, considering only those miRNAs highly involved in disease-associated signaling network, namely miR-107 and miR-26b-5p. The analysis conducted on a subcohort of ten subjects confirmed the microarray results. The twelve analyzed miRNAs, in line with bioinformatic analysis, were linked toinvolved in immune response signaling pathways, such as triggering receptor expressed on myeloid cells 1 (TREM1), TLR, interferon signaling, protein kinase C theta (PRKCQ) signaling in T lymphocytes and B cell receptor signaling (Rager et al. 2014). Some of these miRNAs also have As-related health outcomes including

2 3 487 cancer (let-7a-5p, miR-16-5p, and miR-20b-5p) (Lui et al. 2007; Cascio et al. 2010) and diabetes 4 5 488 mellitus (miR-107, miR-126-3p) (Guay et al. 2011). (Rager et al. 2014). 6 7 489 Lead (Pb). Lead Pb-is a cumulative toxic metal that affects multiple body systems and is associated 8 9 10 4 9 0 with numerous toxic events, such as CVD and chronic kidney disease (Rehman et al. 2018). 11 ¹²491 A cross-sectional study assessed tThe relation association between heavy metals, microalbuminuria 13 14 ₁₅ 492 (a marker of vascular and renal damage), and miRNAs was analyzed in adolescents, showing that 16 17 4 93 urinary Pb and As levels were correlated with miR-21. No relationship between heavy metals and 18 ¹⁹494 microalbuminuria was found, but urinary Pb and As levels were correlated with miR-21-5p that was 20 21 22 495 also associated with microalbuminuria. It was postulated that this miRNA was protective against the 23 24 4 96 development of albuminuria (a marker of vascular and renal damage) as it inhibited apoptosis of 25 26 4 97 podocytes. But, The results showed that miR-21-5p might be involved in the pathogenetic 27 29 29 498 28 mechanisms linking heavy metal exposure and albuminuria (Kong et al. 2012). 30 31 499 Exposure to Pb may also contribute to increased risk of spontaneous abortion and preterm delivery, 32 33 500 disrupting processes involved in normal development (Silbergeld & Patrick 2005). 34 ³⁵ 501 Sanders et al. examined the association between metal levels and the expression of 74 miRNAs using 36 37 Nanostring nCounter®assay. In their cohort study, pregnant women were enrolled. Maternal Pb and 38 502 39 40 503 mercury (Hg) exposure in the cervix during the second trimester of pregnancy have determined shown 41 42 43 504 a decreased expression of miR-575 and miR-4286 in the cervix cells in relation to Pb levels of tibial 44 45 505 bone, while seventeen miRNAs were found to be negatively associated with toenail Hg levels. 46 47 506 Functional analysis revealed that Tthese miRNAs associated to Hg were implicated in reproductive 48 ⁴⁹ 507 system development and morphology, and Pb-associated gene targets were enriched for 50 51 ₅₂ 508 preeclampsia.organismal, cellular, and cardiovascular system development, as well as cell cycle, 53 54 509 cancer, and gene expression. Notably, miR-125b, miR-205, and let-7 were involved in various human 55 ⁵⁶ 510 cancers and directly regulate some oncogenes, including PTEN, p53, and RAS. Other impacted 57 ⁵⁸ 59</sub>511 Furthermore, these miRNAs have known impacts on a number of cell cycle and proliferation 60 512 pathways that could affect parturition and the reproductive system in general. In particular, miR-4286

is predicted to target three genes involved in aryl-hydrocarbon receptor (AHR) signaling pathway
(Sanders et al. 2015), which plays a critical role in the has a role in the processes of female
reproductive system (Hernández-Ochoa et al. 2009)₁, including AHR repressor, prostaglandin E
synthase 3 (PTGES3), and tumor protein p73 (Sanders et al. 2015). Again, In the study of Li et al.,
already described above, some miRNAs were altered following Pb exposure in placenta samples.
Down-regulation of MiR-10a-5p, miR-146a-5p, miR-190b, miR-431-5p and, let-7f-5p were found
to be down-regulated, while miR-651 was up-regulated, suggesting the potential role of miRNAs as
markers of prenatal environmental exposures (Li et al. 2015). Literature analysis indicated that miR146a-5p which-targets genes involving in TLR pathway (tumor necrosis factors-receptor associated
factors 1- TRAF1 and IRAK1) genes that are involved in TLR pathway, that play a key role in the
innate immune response indicates a host cell mediated immune activation in response to Pb (Saba et
al. 2014). Down-regulation of let-7f plays a critical role in early development, primarily by driving
cell differentiation, Dysregulation of miR-190b has been speculated to lead to several mental disorders
due to its regulation of Neuregulin 3 (NRG3)-mediated inhibitory control processes of the amygdala
(Pietrzykowski & Spijker 2014).

Mercury (Hg). Environmental and occupational Hg exposure can occur due to mining and pollution
and may cause serious health risks in the cardiovascular, nervous, and immune systems (Vrijheid et
al. 2016; Rehman et al. 2018).

The expression of miRNAs in plasma samples was evaluated in a case-control study carried out by Ding and colleagues involving workers occupationally exposed to Hg, divided into chronic Hg poisoning group, Hg absorbing group, and control group in a Hg thermometer plant. The authors used miRNA microarray to investigate the expression of 418 miRNAs in these groups and the results showed that four miRNAs (miR-16-5p, miR-30c-3p, miR-181a-5p and let-7e-5p) were downregulated and four miRNAs (miR-92a-3p, miR-122-5p, miR-451a and miR-486-5p) were upregulated in the Hg poisoning group compared to the other two. High levels of miR-92a-3p and miR-486-5p in the mercury poisoned group were confirmed by qRT-PCR (Ding et al. 2016).Since Hg exposure in female workers affected miR-92a-3p and miR-486-5p expression, these miRNAs could be used as biomarker for Hg exposure. Literature analysis showed that Up-regulation of miR-92a is linked to endothelial dysfunction which causes the formation of atherosclerotic lesions targeting eukaryotic ribosome biogenesis protein (ERB1); up-regulation of miR-486 <u>may be</u> is linked to inflammatory diseases through the enhancement of NFkB activation associated with Hg poisoning (Song et al. 2013). (Ding et al. 2016).

Several studies have found that high prenatal exposure to Hg for reproductive females has been related with increased preterm birth risk and other adverse birth outcomes (Silbergeld & Patrick 2005). In the aforementioned study of Li et al., Sseventeen miRNAs were down-regulated in response to high levels of Hg in human placentas, including some let-7 family members which may indicate a state of disrupted placental development in response to chemical exposures (Li et al. 2015).

Chromium (Cr). Chromium^{*F*} is naturally found in rocks and soil, it can be liquid, solid or gas and exists in various oxidation states. Epidemiological studies have suggested that Cr exposure may be linked with metabolic diseases and CVD (Rehman et al. 2018).

Dioni et al. analyzed the leukocytes miRNA expression levels of ninety obese subjects. In the screening phase 43 miRNAs were negatively associated with Cr levels, and only ten miRNAs were chosen for the validation phase. Among these, Nnine miRNAs (miR-451a, miR-301, miR-15b, miR-21-5p, miR-26a-5p, miR-362-3p, miR-182, miR-183-5p and miR-486-3p) confirmed the same trend of the first phase. Functional analysis identified the top canonical pathways for these miRNAs, namely molecular mechanisms of cancer, axonal guidance signaling, protein kinase A (PKA) signaling, role of nuclear factor of activated T cells (NFAT) in cardiac hypertrophy PTEN signaling. were down-regulated in association with Cr exposure including miR-451 and miR-486-3p In particular, miR-486-3p was positively associated with blood pressure and may be a factor risk in CVD due to its interaction with PTEN pathway, as also identified by bioinformatic analysis; on the contrary, miR-451a which-wasere negatively linked to glycated hemoglobin (HbA1c) (Dioni et al. 2017); therefore it and blood pressure, respectively. miR-451-could have a role in the diabetes

pathogenesis because <u>the</u> increases in HbA1c indicate <u>a</u> decreased control of blood glucose levels
 (Soliman et al. 2014).; it also regulates p38 MAP kinase signaling. miR-486-3p may be a factor in
 CVD risk due to its interaction with PTEN pathway (Dioni et al. 2017).

A-<u>The study conducted by Li et al.</u>, on 117 workers in a chromate production plant in China, showed a significant <u>inverse</u> association of <u>high</u> Cr exposure with plasma miR-3940-5p level, which was also linked to micronuclei frequency (Li et al. 2014), a biomarker of DNA damage (Bonassi et al. 2005). In addition to bioinformatic analyses, an enzyme-linked immunosorbent assay was performed, in order to quantify the protein expression levels of miR-3049-5p-mediated genes. The results showed that miRNA-3940-5p regulates the X-ray repair cross complementing 2 (XRCC2) gene (Li et al. 2014) which <u>is has been implicated in DNA repair mechanisms (Serra et al. 2013)</u>, and may increase this activity when reaching a certain exposure of Cr.<u>Hence</u>, <u>t</u>This result demonstrates that miR-3940-5p can play a modulatory role in Cr-induced genetic damage (Li et al. 2014).

Other metals. The association between several metals and PAHs with miRNAs expression was analyzed in <u>a case-control study involving</u> 360 healthy male coke oven workers. The expression of many plasma miRNAs was found to be negatively associated with aluminum, antimony, Pb, and titanium, and positively associated with molybdenum and tin. This study demonstrated a relationship between some miRNAs and biomarkers for genetic damage and oxidative stress, such as micronuclei and <u>8-OH-dG 8- hydroxydeoxyguanosine(Deng et al. 2019)</u>. Among the analyzed miRNAs, some have important functions. In particular, ILet-7b-5p can regulate the expression levels of genes deputated to DNA-repair genes (Spolverini et al. 2017) and it is involved in p53-regulated pro-apoptotic pathway and nucleotide excision repair pathway (Saleh et al. 2011; Encarnación et al. 2016). miR-126-3p plays a role is implicated in cancer-related processes, such as inflammatory responses (Zampetaki & Mayr 2012)_- and cellular protection against reactive oxygen species imbalance. Improper expression of miR-16-5p can negatively affect DNA repair mechanism influencing by modulating the expression of DNA damage-related proteins (Patel et al. 2017). Finally, miR-320b is known to be down-regulated in human cancers, its target is TP53 regulated inhibitor of

1

apoptosis 1 (TRIAP1) through which may control apoptosis process and may exert its activity on apoptosis targeting TP53 regulated inhibitor of apoptosis 1 (TRIAP1) (Li et al. 2016). These findings may suggest a potential <u>linkage mechanistic connection</u> between the <u>complex</u>-metal-PAH <u>complex</u> interactions and <u>the</u> early harmful effects on human health (Deng et al. 2019).

5 **Discussion Conclusions** and future directions

This review reports and aggregates the literature evidence on the effects of environmental chemical exposure to-causinge miRNA dysregulation in humans and consequently the alteration of several biological pathways. This may provides possible explanations for links between exposure and disease pathogenesis.

We observed a great heterogeneity in body samples, methodologies for analysis of miRNAs and their
 targets, and study designs, so that it is difficult to directly compare such different studies.

With regard to the sample type adopted for the miRNA analysis, we found that blood (and its components like serum, plasma, and PBMC) was the most used, maybe because of its numerous advantages. miRNAs are highly stable into the bloodstream where are released in a quantity that allows their collection for testing. They derive from target tissues (i.e., brain, kidney, lung) and may reflect their status; in fact, the miRNA expression pattern in tissue and blood is similar (Powrózek et al. 2020). In the blood, the different expression of miRNA in pathological condition may be used as disease biomarker, since it helps to discriminate between patients and healthy subjects. This allows obtaining information on the health condition and the organ physiology, suggesting the high utility of blood, and especially of plasma, for diagnostic, predictive and prognostic purposes (Panagopoulos & Lambrou 2018; Iacob et al. 2020). However, among the different body fluids, the expression pattern of miRNAs may change. Moreover, the extraction methodology (for example the extraction of miRNAs incorporated in MVs may yeld different results) and the platform employed can enhance these differences, making difficult miRNAs comparison and functional analysis.

2 2	~	• ~
3 4	6	16
5	6	17
6	0.	L/
7 0	6	18
o 9	0.	10
10	6	19
11	-	-
12	62	20
13 14		
15	62	21
16		
17	6	22
18 19		
20	62	23
21	~	
22	6.	24
23 24	<u>د</u> ،	
25	0.	25
26	6	26
27	0.	-0
28	62	27
30		
31	6'	20
32	0.	20
33 24	6	29
35	0.	
36	63	30
37		
38	63	31
40		
41	6	32
42	~	
43 11	6.	33
45	6	л
46	0.	54
47	6	35
48 40	0.	
49 50	63	36
51		
52	6	27
53	0.	, ,
55	6	38
56	5.	
57	63	39
58		
14		

60

16	The different analytical methods used for identifying miRNAs may cause differences in their
17	detection levels, indicating the need for a standardized approach to miRNA analytical processing.
18	qRT-PCR was the most adopted instrument of analysis, as it is considered the "gold standard" because
19	of its higher precision and sensitivity, rapidity and ease of use (Hunt et al. 2015); it is often used to
20	validate results from the microarray platform. The latter is cheaper than qRT-PCR but has a low
21	dynamic range in detection and a low specificity (Heller 2002). Another rarely adopted methodology
22	is Nanostring Technologies' nCounter [®] platform, a hybridization-based method that provides a simple
23	solution for multiplexed detection of up to 800 miRNAs in a single reaction, but it is expensive
24	(Mathew et al. 2020), as is next-generation sequencing that is capable of concurrently detecting new
25	miRNAs (Hunt et al. 2015). As observed in the analyzed studies, since qRT-PCR can detect only
26	annotated miRNAs, the sequencing approach should be more considered because it could allow the
27	discovery of novel miRNAs to be used as biomarkers.

For a biological interpretation of miRNA function, most studies performed bioinformatics analysis, and in some cases also a literature research. This analysis allowed selection of miRNAs with a putative biological function, paving the way for further experimental and clinical investigations that might confirm the consistency and, possibly, the reasons of the observed associations. In fact, only a few investigations (Guida et al. 2013; Motta et al. 2013; Yamamoto et al. 2013; Fry et al. 2014; Y. Li et al. 2014; Banerjee et al. 2017; Tsamou et al. 2018) included in vivo or in vitro experiments in order to better understand the molecular mechanisms triggered by miRNAs after environmental exposures. In such limited cases, it was possible to identify those genes and pathways regulated by specific miRNAs, which, in turn, may determine a pathological condition.

With respect to the different study designs, we have found that some studies have a cross-sectional
 nature that does not allow identification of the causal association between environmental exposure
 and the alteration of the miRNA profile.

2		
3	6	10
4	0.	+0
5	~	
5	64	41
7		
8	64	42
9		
10	64	13
11	0	13
12	~	
13	64	44
14		
15	64	45
16		
17	6	16
18	0	10
19	~	
20	64	47
21		
21 22	64	18
22		
2J 7/	6	19
24	0-	75
25	_	
20	6	50
27 20		
28	6	51
29		
30	6	52
31	0.	52
32		
33	6	53
34		
35	6!	54
36		
37	~	
38	6	22
39		
40	6	56
41		
42	_	
43	6	57
44		
45	6!	58
46	-	_
47	c	-0
48	0.	פנ
49		
50	6	50
51		
52	6	51
53	5.	
54	c.	50
55	0	72
56		
57	6	53
58		
59	6	54

1

We explored the coherence of miRNA expression (up-regulation or down-regulation) in response to a certain pollutant or within the same category of pollutants (air pollution, organic chemicals, heavy metals). As for PM, the altered level of some miRNAs (miR-21-5p, miR-222-3p, let-7 family members) was observed in several studies but with a differential direction of expression (Bollati et al. 2010; Yan Li et al. 2012; Motta et al. 2013; Yamamoto et al. 2013; Louwies et al. 2016; Rodosthenous et al. 2016; Tsamou et al. 2018). On the contrary, if we look at the whole category (air pollution), two miRNAs, miR-223-3p and miR-132, were found up-regulated in response to most airborne pollutants (PM, O₃, DEP) (Yan Li et al. 2012; Fry et al. 2014; Rodosthenous et al. 2016). In relation to the organic chemicals, the dysregulated expression of common miRNAs within the same pollutant was not found. On the contrary, referring to the whole category, we noticed an altered expression of miR-24 and miR-28 in response to most organic chemicals (PAHs, OP, phthalates and phenols), but with a different direction of expression (Deng et al. 2014; Weldon et al. 2016; Martinez et al. 2019). Concerning the heavy metals, miR-21-5p was observed to be up-regulated after As exposure in some studies (Banerjee et al. 2017; Sun et al. 2017), while miR-126 was common in two studies but with different regulations (Rager et al. 2014; Pérez-Vázquez et al. 2017). Considering the whole category, miR-21-5p was found up-regulated not only in response to As but also after exposure to Pb (Kong et al. 2012).

We have observed that miR-21-5p and let-7 family are those most involved in the response of environmental contaminations (PM, Cr, As, DEP, Pb, Hg, phthalates and phenols) and modulate oxidative stress, immune inflammation, tumor suppression, DNA damage repair, and apoptosis. Cardiovascular pathology, metabolic diseases, respiratory diseases, and child development are the most critical areas, due to the effects on the inflammation pathway; when inflammation becomes chronic, inflammatory factors can lead to cancer. We have also observed that studies investigating the same environmental factor identified distinct miRNAs. For metal rich-PM and As, miR-21-5p was up-regulated in some studies and down- regulated in others (Table 1).

Table 1

<u>In addition to the reasons listed above, tTheis conflicting results heterogeneity</u> may be partly explained by the different tissues and cell types used in the analysis, the different exposure durations between various studies, and the effects of age, gender, and healthy status of the participants.

69 Insert Table 4 about here.

When compiling a summary to identify the most relevant miRNAs, our review showed that the most reported miRNAs, whose expression is associated to environmental contaminants, are miR-21-5p, miR-222-3p, miR-223-3p, and let-7 family members. Table 4 shows the biological effects and the health implications of these miRNAs. The predicted target genes are involved in pathways related to inflammation, oxidative stress, cell cycle regulation, apoptosis, and tumorigenesis. In particular, PTEN, MAPK, and NFkB are the pathways most involved. As for health implications, cardiovascular and respiratory diseases, and child development are the most critical areas, due to the effects on the inflammation pathway; when inflammation becomes chronic, inflammatory factors can lead to cancer.

<u>Hence, i</u>It will_could be <u>useful necessary</u> to <u>deeply</u> investigate the biological processes involving miR-21-5p, miR-222-3p, miR-223-3p, and let-7 family <u>biological processes that may which</u> lead to the <u>development onset</u> of several diseases, and this could help further analysis to establish in order to identify new_potential therapeutic approaches in NCDs management. <u>Nevertheless, these miRNAs</u> showed to be sensitive to multiple pollutants and this condition somewhat reduces their specificity as exposure biomarker for a given toxic substance. For this reason, it would be needed to search for miRNAs that more specifically respond to a certain contaminant.

Conclusions

Our review identified numerous miRNAs dysregulated after environmental exposure, although there are few studies conducted in humans. In some cases, important advances have been made in relation to the associations between specific miRNAs and biological responses to environmental risk factors.

]	However, large-scale supplementary investigations are mandatory to identify the actual causative
1	roles. In addition, in vitro or in vivo experiments could be integrated into the study design, in order
1	to have more consistent results. Indeed, an integrated analysis, combining miRNA expression and
1	mRNA expression of critical related genes, may be important to understanding the mechanisms under
	genetic damage.
F	Further efforts should be made to systematize the scientific evidence, both analyzing miRNAs that
5	pecifically intervene in a given disease in relation to a particular pollutant and identifying miRNAs
t1	hat which may be used as possible exposure biomarkers.

1	
3 701 4	Diclosure of interest
5 6 702	The authors report no conflict of interest.
7 8 703	Financial diclosure
10 70 4 11	The authors have no funding to disclose.
¹² 705 13	
14 15 706	References
17 707 18	Almeida DL, Pavanello A, Saavedra LP, Pereira TS, De Castro-Prado MAA, De Freitas Mathias
¹⁹ 708 20	PC. 2019. Environmental monitoring and the developmental origins of health and disease. Journal
²¹ 22 709	of Developmental Origins of Health and Disease. 10(6):608–615.
²³ ²⁴ 25	Ardekani AM, Naeini MM. 2010. The role of microRNAs in human diseases. Avicenna Journal of
26 27 711 28	Medical Biotechnology. 2(4):161–179.
²⁹ 30 ⁷¹²	Banerjee N, Bandyopadhyay AK, Dutta S, Das JK, Roy Chowdhury T, Bandyopadhyay A, Giri
31 32 713	AK. 2017. Increased microRNA 21 expression contributes to arsenic induced skin lesions, skin
34 714 35	cancers and respiratory distress in chronically exposed individuals. Toxicology. 378:10–16.
36 37 715 38	Bollati V, Angelici L, Rizzo G, Pergoli L, Rota F, Hoxha M, Nordio F, Bonzini M, Tarantini L,
³⁹ 716 40	Cantone L, et al. 2015. Microvesicle-associated microRNA expression is altered upon particulate
41 42 43	matter exposure in healthy workers and in A549 cells. Journal of Applied Toxicology. 35(1):59–67.
⁴⁴ 718 45	Bollati V, Marinelli B, Apostoli P, Bonzini M, Nordio F, Hoxha M, Pegoraro V, Motta V, Tarantini
46 47 719	L, Cantone L, et al. 2010. Exposure to metal-rich particulate matter modifies the expression of
40 49 72 0 50	candidate MicroRNAs in peripheral blood leukocytes. Environmental Health Perspectives.
⁵¹ 721 52 53	118(6):763–768.
54 722 55	Bonassi S, Ugolini D, Kirsch-Volders M, Strömberg U, Vermeulen R, Tucker JD. 2005. Human
⁵⁶ 723 57	population studies with cytogenetic biomarkers: Review of the literature and future prospectives.
⁵⁸ 59 724 60	Environmental and Molecular Mutagenesis. 45(2-3):258-270.
725	Bonetta Sara, Bonetta Silvia, Schilirò T, Ceretti E, Feretti D, Covolo L, Vannini S, Villarini M,

2	
3	726
4	
5	777
6	121
7	
, Q	728
0	0
9	
10	729
11	125
12	
13	730
14	
15	721
16	/31
17	
10	722
10	/32
19	
20	733
21	
22	724
23	734
24	
25	
25	735
20	
27	736
28	/30
29	
30	737
31	
32	
33	738
34	
25	720
22	/39
20	
37	740
38	
39	
40	741
41	
42	
43	742
44	
15	743
45	/ 45
40	
47	7//
48	/ 44
49	
50	745
51	
52	746
53	,+0
54	
55	7/7
22	/4/
20	
57	748
58	
59	7/0
60	749

Moretti M, Verani M, et al. 2019. Mutagenic and genotoxic effects induced by PM0.5 of different
Italian towns in human cells and bacteria: The MAPEC_LIFE study. Environmental Pollution.
245:1124–1135.

Cascio S, D'Andrea A, Ferla R, Surmacz E, Gulotta E, Amodeo V, Bazan V, Gebbia N, Russo A.
 2010. miR-20b modulates VEGF expression by targeting HIF-1 alpha and STAT3 in MCF-7 breast
 cancer cells. Journal of cellular physiology. 224(1):242–9.

Deng Q, Dai X, Feng W, Huang S, Yuan Y, Xiao Y, Zhang Z, Deng N, Deng H, Zhang Xiao, et al.
2019. Co-exposure to metals and polycyclic aromatic hydrocarbons, microRNA expression, and
early health damage in coke oven workers. Environment International. 122:369–380.

Deng Q, Huang S, Zhang Xiao, Zhang W, Feng J, Wang T, Hu D, Guan L, Li J, Dai X, et al. 2014.
Plasma microRNA expression and micronuclei frequency in workers exposed to polycyclic
aromatic hydrocarbons. Environmental Health Perspectives. 122(7):719–725.

⁷ Jing E, Zhao Q, Bai Y, Xu M, Pan L, Liu Q, Wang B, Song X, Wang J, Chen L, Zhu B. 2016.

⁵739 Plasma microRNAs expression profile in female workers occupationally exposed to mercury.

Journal of Thoracic Disease. 8(5):833–841.

⁰741 Dioni L, Sucato S, Motta V, Iodice S, Angelici L, Favero C, Cavalleri T, Vigna L, Albetti B,

Fustinoni S, et al. 2017. Urinary chromium is associated with changes in leukocyte miRNA expression in obese subjects. European Journal of Clinical Nutrition. 71(1):142–148.

Domingues ÉP, Silva GG, Oliveira AB, Mota LM, Santos VSV, de Campos EO, Pereira BB. 2018.

Genotoxic effects following exposure to air pollution in street vendors from a high-traffic urban

 2 746 area. Environmental monitoring and assessment. 190(4):215.

Encarnación J, Ortiz C, Vergne R, Vargas W, Coppola D, Matta JL. 2016. High DRC levels are

⁷748 associated with let-7b overexpression in women with breast cancer. International Journal of

Molecular Sciences. 17(6).

1	
2 3 750 4	Engel LS, Laden F, Andersen A, Strickland PT, Blair A, Needham LL, Barr DB, Wolff MS,
5 6 751	Helzlsouer K, Hunter DJ, et al. 2007. Polychlorinated biphenyl levels in peripheral blood and non-
/ 8 752 9	Hodgkin's lymphoma: A report from three cohorts. Cancer Research. 67(11):5545–5552.
¹⁰ 11 753	Farzan SF, Karagas MR, Chen Y. 2013. In utero and early life arsenic exposure in relation to long-
13 7 54 14	term health and disease. Toxicology and applied pharmacology. 272(2):384-90.
15 16 755 17	De Felice B, Manfellotto F, Palumbo A, Troisi J, Zullo F, Di Carlo C, Di Spiezio Sardo A, De
18 7 56 19	Stefano N, Ferbo U, Guida Marco, Guida Maurizio. 2015. Genome-wide microRNA expression
20 757 21 22	profiling in placentas from pregnant women exposed to BPA. BMC medical genomics. 8(1):56.
23 758 24	Fish JE, Santoro MM, Morton SU, Yu S, Yeh RF, Wythe JD, Ivey KN, Bruneau BG, Stainier DYR,
²⁵ 759 26	Srivastava D. 2008. miR-126 Regulates Angiogenic Signaling and Vascular Integrity.
27 28 760 29	Developmental Cell. 15(2):272–284.
³⁰ 31 761	Fry RC, Rager JE, Bauer R, Sebastian E, Peden DB, Jaspers I, Alexis NE. 2014. Air toxics and
32 33 762 34	epigenetic effects: Ozone altered microRNAs in the sputum of human subjects. American Journal
35 763 36 37	of Physiology - Lung Cellular and Molecular Physiology. 306(12).
38 764 39	Gallo A, Tandon M, Alevizos I, Illei GG. 2012. The majority of microRNAs detectable in serum
⁴⁰ 765 41	and saliva is concentrated in exosomes. PLoS ONE. 7(3):e30679.
42 43 766 44	Gianicolo EAL, Bruni A, Rosati E, Sabina S, Guarino R, Padolecchia G, Leo C, Vigotti MA,
⁴⁵ 767 46	Andreassi MG, Latini G. 2012. Congenital anomalies among live births in a polluted area. A ten-
47 48 768 49	year retrospective study. BMC pregnancy and childbirth. 12:165.
⁵⁰ 769	Gianicolo EAL, Mangia C, Cervino M, Bruni A, Andreassi MG, Latini G. 2014. Congenital
52 53 770 54	anomalies among live births in a high environmental risk areaa case-control study in Brindisi
55 771 56 57	(southern Italy). Environmental research. 128:9–14.
57 58 772 59	Griffiths-Jones S, Saini HK, Van Dongen S, Enright AJ. 2008. miRBase: Tools for microRNA
60 773	genomics. Nucleic Acids Research. 36(SUPPL. 1).

1 2	
³ 774 4	Guay C, Roggli E, Nesca V, Jacovetti C, Regazzi R. 2011. Diabetes mellitus, a microRNA-related
5 6 775	disease? Translational Research. 157(4):253-264.
8 9 776	Guida Maurizio, Marra ML, Marra M, Zullo F, Guida Marco, Trifuoggi M, Biffali E, Borra M, De
10 11 777	Mieri G, D'Alessandro R, De Felice B. 2013. Association between exposure to dioxin-like
12 13 778 14	polychlorinated biphenyls and miR-191 expression in human peripheral blood mononuclear cells.
15 779 16 17	Mutation research. 753(1):36–41.
18 780 19	Heller MJ. 2002. DNA Microarray Technology: Devices, Systems, and Applications. Annual
²⁰ 781 21 22	Review of Biomedical Engineering. 4(1):129–153.
23 782 24	Hernández-Ochoa I, Karman BN, Flaws JA. 2009. The role of the aryl hydrocarbon receptor in the
²⁵ 783 26 27	female reproductive system. Biochemical Pharmacology. 77(4):547-559.
²⁸ 784 29	Hoesel B, Schmid JA. 2013. The complexity of NF-kB signaling in inflammation and cancer.
³⁰ 31 32	Molecular Cancer. 12(1).
³³ 786 34	van den Hooven EH, Pierik FH, de Kluizenaar Y, Hofman A, van Ratingen SW, Zandveld PYJ,
³⁵ 36787	Russcher H, Lindemans J, Miedema HME, Steegers EAP, Jaddoe VW V. 2012. Air pollution
37 38 788 39	exposure and markers of placental growth and function: the generation R study. Environmental
⁴⁰ 789 41	health perspectives. 120(12):1753–9.
42 43 790 44	Humphrey KM, Pandey S, Martin J, Hagoel T, Grand'Maison A, Ohm JE. 2019. Establishing a role
⁴⁵ 791 ₄₆	for environmental toxicant exposure induced epigenetic remodeling in malignant transformation.
⁴⁷ 48 49	Seminars in cancer biology. 57:86–94.
⁵⁰ 793 51	Hunt EA, Broyles D, Head T, Deo SK. 2015. MicroRNA Detection: Current Technology and
52 53 794 54	Research Strategies. Annual Review of Analytical Chemistry. 8(1):217–237.
⁵⁵ 56 795	Iacob DG, Rosca A, Ruta SM. 2020. Circulating microRNAs as non-invasive biomarkers for
57 58 796 59	hepatitis B virus liver fibrosis. World Journal of Gastroenterology. 26(11):1113-1127.
⁶⁰ 797	Ikezuki Y, Tsutsumi O, Takai Y, Kamei Y, Taketani Y. 2002. Determination of bisphenol A

Jedeon K, De la Dure-Molla M, Brookes SJ, Loiodice S, Marciano C, Kirkham J, Canivenc-Lavier

Kim HS, Lee KS, Bae HJ, Eun JW, Shen Q, Park SJ, Shin WC, Yang HD, Park M, Park WS, et al.

2015. MicroRNA-31 functions as a tumor suppressor by regulating cell cycle and epithelial-

mesenchymal transition regulatory proteins in liver cancer. Oncotarget. 6(10):8089–102.

Kim KH, Jahan SA, Kabir E, Brown RJC. 2013. A review of airborne polycyclic aromatic

hydrocarbons (PAHs) and their human health effects. Environment International. 60:71-80.

Kong APS, Xiao K, Choi KC, Wang G, Chan MHM, Ho CS, Chan I, Wong CK, Chan JCN, Szeto

CC. 2012. Associations between microRNA (miR-21, 126, 155 and 221), albuminuria and heavy

metals in Hong Kong Chinese adolescents. Clinica chimica acta; international journal of clinical

Kozomara A, Birgaoanu M, Griffiths-Jones S. 2019. miRBase: from microRNA sequences to

Krauskopf J, Caiment F, van Veldhoven K, Chadeau-Hyam M, Sinharay R, Chung KF, Cullinan P,

Collins P, Barratt B, Kelly FJ, et al. 2018. The human circulating miRNome reflects multiple organ

disease risks in association with short-term exposure to traffic-related air pollution. Environment

Rantakokko P, Kyrtopoulos SA, Kleinjans JC. 2017. MicroRNA profile for health risk assessment:

Krauskopf J, De Kok TM, Hebels DG, Bergdahl IA, Johansson A, Spaeth F, Kiviranta H,

Environmental exposure to persistent organic pollutants strongly affects the human blood

M-C, Boudalia S, Bergès R, Harada H, et al. 2013. Enamel defects reflect perinatal exposure to

concentrations in human biological fluids reveals significant early prenatal exposure. Human

reproduction (Oxford, England). 17(11):2839-41.

chemistry. 413(13–14):1053–7.

International. 113:26–34.

function. Nucleic acids research. 47(D1):D155–D162.

microRNA machinery. Scientific Reports. 7(1).

bisphenol A. The American journal of pathology. 183(1):108–18.

1
2
³ /98
5 - 00
⁻ 799
7
⁸ 800
9
10
11 001
12 802
13 802
15
16 803
17
18 804
19
²⁰ 805
21
22
²³ 806 24
25 007
26 807
27
²⁸ 808
29
³⁰ 809
31
32 33 810
34
35 811
36
37
38 812
39
⁴⁰ 813
41
<u></u> 43 81 <i>1</i>
44
45 815
46
47 016
48 810
49
50817
ו כ 52
53 818
54
55 819
56
⁵⁷ 820
58
59 60 821
ou

URL: http://mc.manuscriptcentral.com/cije Email: ijehr-els@salford.ac.uk

1	
2 3 822 4	LaRocca J, Binder AM, McElrath TF, Michels KB. 2016. First-Trimester Urine Concentrations of
⁵ 823	Phthalate Metabolites and Phenols and Placenta miRNA Expression in a Cohort of U.S. Women.
7 8 824 9	Environmental health perspectives. 124(3):380–7.
10 11 825	Li Q, Kappil MA, Li A, Dassanayake PS, Darrah TH, Friedman AE, Friedman M, Lambertini L,
13 826 14	Landrigan P, Stodgell CJ, et al. 2015. Exploring the associations between microRNA expression
¹⁵ 827 16	profiles and environmental pollutants in human placenta from the National Children's Study (NCS).
¹⁷ 18 828 19	Epigenetics. 10(9):793-802.
²⁰ 829 21	Li Yan, Li M, Liu Y, Song G, Liu N. 2012. A Microarray for MicroRNA Profiling in Spermatozoa
22 23 830	from Adult Men Living in an Environmentally Polluted Site. Bulletin of Environmental
25 831 26	Contamination and Toxicology. 89(6):1111–1114.
27 28 832 29	Li Y, Li P, Yu S, Zhang J, Wang T, Jia G. 2014. miR-3940-5p associated with genetic damage in
30 833 31 22	workers exposed to hexavalent chromium. Toxicology letters. 229(1):319–26.
33 834 34	Li Y, Tang X, He Q, Yang X, Ren X, Wen X, Zhang J, Wang Y, Liu N, Ma J. 2016.
35 835 36	Overexpression of Mitochondria Mediator Gene TRIAP1 by miR-320b Loss Is Associated with
³⁷ 836 38	Progression in Nasopharyngeal Carcinoma. PLoS genetics. 12(7):e1006183.
⁴⁰ 837 41	Li Yuan-fang, Wang D, Zhao B, Wang W, Yuan S, Huang C, Chen Y, Zheng Y, Keshari RP, Xia J,
⁴² 43838	Zhou Z. 2012. Poor prognosis of gastric adenocarcinoma with decreased expression of AHRR. PloS
44 45 839 46	one. 7(8):e43555.
47 48 840 49	Liang H, Liu M, Yan X, Zhou Y, Wang W, Wang X, Fu Z, Wang N, Zhang S, Wang Y, et al. 2015.
50 841 51	miR-193a-3p functions as a tumor suppressor in lung cancer by down-regulating ERBB4. The
52 842 53	Journal of biological chemistry. 290(2):926–40.
55 843 56	Lignell S, Winkvist A, Bertz F, Rasmussen KM, Glynn A, Aune M, Brekke HK. 2016.
⁵⁷ 844 58	Environmental organic pollutants in human milk before and after weight loss. Chemosphere.
⁵⁹ 60 845	159:96–102.

1	
2 3 846 4	Liu X, Li J, Qin F, Dai S. 2016. miR-152 as a tumor suppressor microRNA: Target recognition and
5 6 7	regulation in cancer. Oncology letters. 11(6):3911–3916.
8 8 9	Lodovici M, Bigagli E. 2011. Oxidative stress and air pollution exposure. Journal of toxicology.
10 ₁₁ 849 12	2011:487074.
¹³ 850	Louwies T, Vuegen C, Panis LI, Cox B, Vrijens K, Nawrot TS, De Boever P. 2016. miRNA
15 16 851 17	expression profiles and retinal blood vessel calibers are associated with short-term particulate
18 852 19	matter air pollution exposure. Environmental Research. 147:24–31.
20 21 853 22	Lui WO, Pourmand N, Patterson BK, Fire A. 2007. Patterns of known and novel small RNAs in
23 854 24 25	human cervical cancer. Cancer Research. 67(13):6031–6043.
26 855 27	Martinelli N, Olivieri O, Girelli D. 2013. Air particulate matter and cardiovascular disease: A
²⁸ 856 29 30	narrative review. European Journal of Internal Medicine. 24(4):295–302.
31 857 32	Martinez RM, Hauser R, Liang L, Mansur A, Adir M, Dioni L, Racowsky C, Bollati V, Baccarelli
³³ 858 34 35	AA, Machtinger R. 2019. Urinary concentrations of phenols and phthalate metabolites reflect
36 859 37	extracellular vesicle microRNA expression in follicular fluid. Environment International. 123:20–
38 860 39	28.
40 41 861 42	Mathew R, Mattei V, Al Hashmi M, Tomei S. 2020. Updates on the Current Technologies for
43 862 44	microRNA Profiling. MicroRNA (Shariqah, United Arab Emirates). 9(1):17–24.
45 46 863 47	Matz CJ, Egyed M, Hocking R, Seenundun S, Charman N, Edmonds N. 2019. Human health effects
48 864 49	of traffic-related air pollution (TRAP): a scoping review protocol. Systematic reviews. 8(1):223.
50 51 865 52	Mercorio R, Bonzini M, Angelici L, Iodice S, Delbue S, Mariani J, Apostoli P, Pesatori AC, Bollati
⁵³ 866 54	V. 2017. Effects of metal-rich particulate matter exposure on exogenous and endogenous viral
56 867 57	sequence methylation in healthy steel-workers. Environmental Research. 159:452-457.
⁵⁸ 868 59	Michailidi C, Hayashi M, Datta S, Sen T, Zenner K, Oladeru O, Brait M, Izumchenko E, Baras A,
869	VandenBussche C, et al. 2015. Involvement of epigenetics and EMT-related miRNA in arsenic-

2	
3	870
4	
5	871
6	071
7	
8	872
9	
10	873
11	
12	87/
13	074
14	
16	875
17	
18	876
19	070
20	077
21	8//
22	
23	878
24	070
25	070
26	0/9
27	
28	880
29	
30	881
31	
32	002
33	882
34	
35	883
36	
3/	001
38	884
39 40	
40 1	885
41 42	
42 43	886
44	
45	887
46	
47	
48	888
49	
50	889
51	
52	890
53	
54	801
55	160
56	
57	892
58	

induced neoplastic transformation and their potential clinical use. Cancer Prevention Research.
8(3):208–221.

872 Miguel V, Cui JY, Daimiel L, Espinosa-Díez C, Fernández-Hernando C, Kavanagh TJ, Lamas S.

2018. The Role of MicroRNAs in Environmental Risk Factors, Noise-Induced Hearing Loss, and

874 Mental Stress. Antioxidants and Redox Signaling. 28(9):773–796.

Misso G, Di Martino MT, De Rosa G, Farooqi AA, Lombardi A, Campani V, Zarone MR, Gullà A, 7 8 876 Tagliaferri P, Tassone P, Caraglia M. 2014. Mir-34: a new weapon against cancer? Molecular 9 0 877 therapy Nucleic acids. 3:e194.

Motta V, Angelici L, Nordio F, Bollati V, Fossati S, Frascati F, Tinaglia V, Bertazzi PA, Battaglia C, Baccarelli AA. 2013. Integrative Analysis of miRNA and inflammatory gene expression after acute particulate matter exposure. Toxicological sciences : an official journal of the Society of Toxicology. 132(2):307–16.

Panagopoulos P, Lambrou G. 2018. The Involvement of MicroRNAs in Osteoarthritis and Recent
 Developments: A Narrative Review. Mediterranean Journal of Rheumatology. 29(2):67–79.

Panico A, Grassi T, Bagordo F, Idolo A, Serio F, Tumolo MR, De Giorgi M, Guido M, Tutino M,
 De Donno A. 2020. Micronucleus Frequency in Exfoliated Buccal Cells of Children Living in an
 Industrialized Area of Apulia (Italy). International journal of environmental research and public
 health. 17(4).

Patel N, Garikapati KR, Pandita RK, Singh DK, Pandita TK, Bhadra U, Bhadra MP. 2017. miR-

15a/miR-16 down-regulates BMI1, impacting Ub-H2A mediated DNA repair and breast cancer cell
 sensitivity to doxorubicin.[Erratum appears in Sci Rep. 2017 Oct 10;7(1):12932; PMID: 29018209].
 Scientific Reports. 7(1):4263.

⁵⁷ 892 Pavanello S, Bonzini M, Angelici L, Motta V, Pergoli L, Hoxha M, Cantone L, Pesatori AC,

⁵⁹₆₀893 Apostoli P, Tripodi A, et al. 2016. Extracellular vesicle-driven information mediates the long-term

1 ว	
2 3 894 4	effects of particulate matter exposure on coagulation and inflammation pathways. Toxicology
5 6 7	letters. 259:143–150.
8 8 9	Pérez-Vázquez MS, Ochoa-Martínez ÁC, RuÍz-Vera T, Araiza-Gamboa Y, Pérez-Maldonado IN.
¹⁰ 11 897	2017. Evaluation of epigenetic alterations (mir-126 and mir-155 expression levels) in Mexican
12 13 898	children exposed to inorganic arsenic via drinking water. Environmental science and pollution
14 15 899 16	research international. 24(36):28036–28045.
17 18 900 19	Pierfelice T, Alberi L, Gaiano N. 2011. Notch in the vertebrate nervous system: an old dog with
²⁰ 901 21	new tricks. Neuron. 69(5):840–55.
22 23 902 24	Pietrzykowski AZ, Spijker S. 2014. Impulsivity and comorbid traits: a multi-step approach for
²⁵ 903 26	finding putative responsible microRNAs in the amygdala. Frontiers in neuroscience. 8:389.
²⁸ 904 29	Popovic R, Shah MY, Licht JD. 2013. Epigenetic therapy of hematological malignancies: where are
³⁰ 31905 32	we now? Therapeutic advances in hematology. 4(2):81–91.
³³ 906 34	Powrózek T, Porgador A, Małecka-Massalska T. 2020. Detection, prediction, and prognosis: blood
³⁵ 36907	circulating microRNA as novel molecular markers of head and neck cancer patients. Expert review
37 38 908 39	of molecular diagnostics. 20(1):31–39.
40 41 909 42	Pradhan S, Nagashri MN, Gopinath KS, Kumar A. 2011. Expression profiling of CYP1B1 in oral
43 910 44	squamous cell carcinoma: counterintuitive downregulation in tumors. PloS one. 6(11):e27914.
45 46 911 47	Rager JE, Bailey KA, Smeester L, Miller SK, Parker JS, Laine JE, Drobná Z, Currier J, Douillet C,
48 912 49	Olshan AF, et al. 2014. Prenatal arsenic exposure and the epigenome: altered microRNAs
⁵⁰ 913 51	associated with innate and adaptive immune signaling in newborn cord blood. Environmental and
52 53 914 54	molecular mutagenesis. 55(3):196–208.
⁵⁵ 915 56	Rehman K, Fatima F, Waheed I, Akash MSH. 2018. Prevalence of exposure of heavy metals and
57 58 916 59	their impact on health consequences. Journal of Cellular Biochemistry. 119(1):157-184.
⁶⁰ 917	Rider CF, Carlsten C. 2019. Air pollution and DNA methylation: Effects of exposure in humans.

2	
3 918 4	Clinical Epigenetics. 11(1).
6 919 7	Rider CF, Yamamoto M, Günther OP, Hirota JA, Singh A, Tebbutt SJ, Carlsten C. 2016.
8 920 9	Controlled diesel exhaust and allergen coexposure modulates microRNA and gene expression in
10 11 12	humans: Effects on inflammatory lung markers. Journal of Allergy and Clinical Immunology.
13 922 14	138(6):1690–1700.
15 16 923	Rodosthenous RS, Coull BA, Lu Q, Vokonas PS, Schwartz JD, Baccarelli AA. 2016. Ambient
18 924 19	particulate matter and microRNAs in extracellular vesicles: A pilot study of older individuals.
²⁰ 925 21	Particle and Fibre Toxicology. 13(1).
22 23 9 2 6 24	Ruiz-Vera T, Ochoa-Martínez ÁC, Pruneda-Álvarez LG, Domínguez-Cortinas G, Pérez-Maldonado
²⁵ 26927	IN. 2019. Expression levels of circulating microRNAs-126, -155, and -145 in Mexican women
27 28 928	exposed to polycyclic aromatic hydrocarbons through biomass fuel use. Environmental and
30 9 2 9 31	Molecular Mutagenesis. 60(6):546–558.
32 33 930	Saba R, Sorensen DL, Booth SA. 2014. MicroRNA-146a: A Dominant, Negative Regulator of the
35 931 36	Innate Immune Response. Frontiers in immunology. 5:578.
37 38 932 39	Saleh AD, Savage JE, Cao L, Soule BP, Ly D, DeGraff W, Harris CC, Mitchell JB, Simone NL.
⁴⁰ 933 41	2011. Cellular stress induced alterations in microrna let-7a and let-7b expression are dependent on
42 43 934 44	p53. PLoS ONE. 6(10).
⁴⁵ 935 46	Sanders AP, Burris HH, Just AC, Motta V, Amarasiriwardena C, Svensson K, Oken E, Solano-
47 48936	Gonzalez M, Mercado-Garcia A, Pantic I, et al. 2015. Altered miRNA expression in the cervix
50 937 51	during pregnancy associated with lead and mercury exposure. Epigenomics. 7(6):885-896.
52 53 938	Serra H, Da Ines O, Degroote F, Gallego ME, White CI. 2013. Roles of XRCC2, RAD51B and
54 55 939 56	RAD51D in RAD51-independent SSA recombination. PLoS genetics. 9(11):e1003971.
57 58 940	Silbergeld EK, Patrick TE. 2005. Environmental exposures, toxicologic mechanisms, and adverse
59 60 941	pregnancy outcomes. In: American Journal of Obstetrics and Gynecology. Vol. 192. [place

2	
³ 942	unknown]: Mosby Inc.; p. S11-21.
5	
6 943	Soliman A, DeSanctis V, Yassin M, Elalaily R, Eldarsy NE. 2014. Continuous glucose monitoring
7	
8 944	system and new era of early diagnosis of diabetes in high risk groups. Indian journal of
10	
₁₁ 945	endocrinology and metabolism. 18(3):274–82.
12	
¹³ 946	Song L, Lin C, Gong H, Wang C, Liu L, Wu J, Tao S, Hu B, Cheng S-Y, Li M, Li J. 2013. miR-
15	
₁₆ 947	486 sustains NF-κB activity by disrupting multiple NF-κB-negative feedback loops. Cell research.
17	
18 948	23(2):274–89.
19 20	
₂₁ 949	Spolverini A, Fuchs G, Bublik DR, Oren M. 2017. let-7b and let-7c microRNAs promote histone
22	
23 950	H2B ubiquitylation and inhibit cell migration by targeting multiple components of the H2B
24 25 a = 4	
-* 951 26	deubiquitylation machinery. Oncogene. 36(42):5819–5828.
27	
²⁸ 952	Stea F, Bianchi F, Cori L, Sicari R. 2014. Cardiovascular effects of arsenic: Clinical and
29 30	
₃₁ 953	epidemiological findings. Environmental Science and Pollution Research. 21(1):244–251.
32	
³³ 954	Sun B, Xue J, Li J, Luo F, Chen X, Liu Y, Wang Q, Qi C, Zou Z, Zhang A, Liu Q. 2017.
34 35	
₃₆ 955	Circulating miRNAs and their target genes associated with arsenism caused by coal-burning.
37	4
38 956	Toxicology research. 6(2):162–172.
39 40	
41 957	Sun Z, Li S, Kaufmann AM, Albers AE. 2014. MiR-21 increases the programmed cell death 4
42	
43 958	gene-regulated cell proliferation in head and neck squamous carcinoma cell lines. Oncology
44 45 a = a	
46	Reports. 32(5):2283–2289.
47	
⁴⁸ 960	Tao J, Qiu B, Zhang D, Wang Y. 2012. Expression levels of Fas/Fas-L mRNA in human brain
49 50	
50 961 51	glioma stem cells. Molecular medicine reports. 5(5):1202–6.
52	
⁵³ 962	Tsamou M, Vrijens K, Madhloum N, Lefebvre W, Vanpoucke C, Nawrot TS. 2018. Air pollution-
54 55	
55 56 963	induced placental epigenetic alterations in early life: a candidate miRNA approach. Epigenetics.
57	12(2) 125 14(
58 964	13(2):133-146.
59 60	
^{°°} 965	Turchinovich A, Tonevitsky AG, Burwinkel B. 2016. Extracellular miRNA: A Collision of Two

1	
2 3 966 4	Paradigms. Trends in Biochemical Sciences. 41(10):883-892.
5 6 967 7	Turchinovich A, Weiz L, Burwinkel B. 2012. Extracellular miRNAs: The mystery of their origin
8 9 9	and function. Trends in Biochemical Sciences. 37(11):460-465.
10 11 969 12	Vrijens K, Bollati V, Nawrot TS. 2015. MicroRNAs as potential signatures of environmental
¹³ 14 970	exposure or effect: A systematic review. Environmental Health Perspectives. 123(5):399-411.
15 ¹⁶ 971 17	Vrijheid M, Casas M, Gascon M, Valvi D, Nieuwenhuijsen M. 2016. Environmental pollutants and
¹⁸ 19972	child health-A review of recent concerns. International Journal of Hygiene and Environmental
20 21 973 22	Health. 219(4–5):331–342.
²³ 24 974	Wei Y, Nazari-Jahantigh M, Neth P, Weber C, Schober A. 2013. MicroRNA-126, -145, and -155: a
25 26 975 27	therapeutic triad in atherosclerosis? Arteriosclerosis, thrombosis, and vascular biology. 33(3):449-
²⁸ 976 29	54.
31 977 32	Weldon BA, Shubin SP, Smith MN, Workman T, Artemenko A, Griffith WC, Thompson B,
³³ 978 34	Faustman EM. 2016. Urinary microRNAs as potential biomarkers of pesticide exposure.
35 36979 37	Toxicology and Applied Pharmacology. 312:19–25.
³⁸ 39980	Wellner U, Schubert J, Burk UC, Schmalhofer O, Zhu F, Sonntag A, Waldvogel B, Vannier C,
40 41 981 42	Darling D, Hausen A Zur, et al. 2009. The EMT-activator ZEB1 promotes tumorigenicity by
43 982 44	repressing stemness-inhibiting microRNAs. Nature Cell Biology. 11(12):1487–1495.
45 46 983 47	Xu X, Hu H, Kearney GD, Kan H, Sheps DS. 2013. Studying the effects of polycyclic aromatic
48 984 49	hydrocarbons on peripheral arterial disease in the United States. The Science of the total
⁵⁰ 985 51 52	environment. 461–462:341–7.
53 986 54	Xue W, Warshawsky D. 2005. Metabolic activation of polycyclic and heterocyclic aromatic
⁵⁵ 987 56 987 57	hydrocarbons and DNA damage: A review. Toxicology and Applied Pharmacology. 206(1):73–93.
⁵⁸ 988 59	Yamamoto M, Singh A, Sava F, Pui M, Tebbutt SJ, Carlsten C. 2013. MicroRNA expression in
⁶⁰ 989	response to controlled exposure to diesel exhaust: Attenuation by the antioxidant N-acetylcysteine

1	
2 3 990 4	in a randomized crossover study. Environmental Health Perspectives. 121(6):670-675.
5 6 991 7	Yan B, Guo Q, Fu FJ, Wang Z, Yin Z, Wei YB, Yang JR. 2015. The role of miR-29b in cancer:
8 9 9	Regulation, function, and signaling. OncoTargets and Therapy. 8:539-548.
10 11 993 12	Zampetaki A, Mayr M. 2012. MicroRNAs in vascular and metabolic disease. Circulation Research.
¹³ 14 994	110(3):508–522.
15 16 995 17	Zhang JJ, Wei Y, Fang Z. 2019. Ozone Pollution: A Major Health Hazard Worldwide. Frontiers in
¹⁸ 19996	immunology. 10:2518.
20 21 997 22	
23 24 998	
25 26 27	
27 28 29	
30 31	
32 33	
34 35	
36 37	
38 39	
40 41	
42 43	
44 45	
46 47	
48	
50	
52	
53 54	
55 56	
57 58	
59 60	

Table 1. Human studies on miRNAs dysregulation after air pollution exposure and their targets

miRNA	Methodology	<u>miRNA</u> <u>expression</u>	Sample type*	<u>Study</u> design	<u>miRNA target</u> <u>analysis</u>	<u>Signaling</u> pathways or target genes	<u>Contaminant</u>	Reference
<u>21-5p, 223-3p</u>	<u>qRT-PCR</u>	Ť	Blood	<u>Cohort</u>	<u>Bioinformatics</u> analysis	<u>HMGB, PTEN</u>	<u>PM</u>	Louwies et al. 2016
<u>15a-5p, 19b-3p,</u> <u>23a-3p, 93-5p,</u> <u>126-3p, 130-3p,</u> <u>142-3p, 146a-5p,</u> <u>150-5p, 191-5p,</u> <u>223-3p, let-7a-5p,</u> <u>let-7g-5p</u>	<u>Nanostring</u> nCounter [®] assay		<u>Serum</u>	<u>Cohort</u>	Bioinformatics analysis	Genes linked to CVD-related pathways	<u>PM</u>	Rodosthenous et al. 2016
<u>20a-5p, 21-5p (1st</u> trimester)	<u>qRT-PCR</u>	1	<u>Placenta</u>	<u>Cohort</u>	<u>Bioinformatics</u> analysis	<u>PTEN</u>	<u>PM</u>	Tsamou et al. 2018
<u>21-5p, 146a-5p,</u> <u>222-3p (2nd</u> trimester)		Ť			<u>qRT-PCR</u>	<u>PTEN (miR-21,</u> <u>miR-20a, and miR-</u> 222)		
<u>21-5p, 222-3p</u>	<u>qRT-PCR</u>	1	Blood leukocytes	<u>Cohort</u>	Bioinformatics analysis	<u>MAPK, TGFβ,</u> FAK, TLR	Metal rich-PM	Bollati et al. 2010
<u>128, 302c</u>	<u>qRT-PCR</u>	1	<u>Plasma</u>	Cohort	Bioinformatics analysis	NFKB	Metal rich-PM	Bollati et al. 2015
182 miRNAs	Microarray	Dysregulated	Blood	<u>Cohort</u>			Metal rich-PM	Motta et al. 2013
<u>29a</u> <u>146a</u> 421	<u>qRT-PCR</u>	1			<u>qRT-PCR</u>	<u>PTEN</u> <u>TGFβ</u> <u>IFNAR2, eNOS,</u> <u>PDGFR</u>		
let-7g 182 miRNAs	Microarray	Dysregulated	Spermatozoa	Case-control		<u>NFKB, TGFβ</u>	Metal rich-PM	Li et al. 2012
<u>10b-5p</u>	<u>qRT-PCR</u>	1			<u>Bioinformatics</u> analysis	BCL2L11, DAZAP1, BCL6, Notch		
<u>33b-5p, 106a-5p,</u> <u>155-5p, 183-5p,</u> <u>205-5p, 208a,</u> <u>222-3p, 223-3p</u>						<u>Notch</u>		
<u>let-7d-5p</u> <u>363-3p</u>		Ŧ				YBX2 Notch		
<u>132-3p</u>	Nanostring nCounter®assay	Ĺ	Bronchial epithelial cells	Randomized	Bioinformatics analysis	CDKN1A	DEP	<u>Rider et al. 2016</u>
<u>183-5p</u>	neounier assay	Ţ		<u>crossover</u>	<u>unu1y515</u>	HLA-A		

URL: http://mc.manuscriptcentral.com/cije Email: ijehr-els@salford.ac.uk

Page 43 of 49

<u>21-5p, 30e, 215,</u> <u>144</u> 144	Nanostring nCounter®assay	1 ↑	Peripheral Blood	Randomized double-blinded crossover	PCR	NRF2	DEP	Yamamoto et al. 2013
<u>1224-5p, 3127-5p</u> <u>27a-5p, 133a-3p,</u> <u>145-5p, 193b-3p,</u> <u>433-3p, 580-3p,</u> <u>6716-3p</u>	Next-generation sequencing	1 1	<u>Plasma</u>	Randomized crossover	<u>Bioinformatics</u> analysis	<u>PI3K-Akt, p53</u>	TRAP	<u>Krauskopf et al.</u> 2018
<u>25-3p, 132-3p,</u> 199a-3p, 222-3p, <u>434-5p, 582-5p</u> <u>143-3p, 223-3p</u> <u>145-5p, 199b-5p</u>	<u>Microarray</u>		Induced sputum	Experimental	Bioinformatics analysis N/P in vitro model	Inflammatory pathways CCND1, MYC	<u>O</u> ₂	<u>Fry et al. 2014</u>
*Sample type is ref \downarrow / \uparrow , difference in 1 N/P, not performed	erred to miRNAs d miRNAs expression	etection 1, respectively up- and o	down-expressed;					
		URL: I	http://mc.manuscriptce	ntral.com/cije I	Email: ijehr-els@sa	alford.ac.uk		

Table 2. Human studies on miRNAs dysregulation after organic chemicals exposure and their targets

miRNA	Methodology	miRNA	Sample type*	Study design	miRNA target	Signaling	Contaminant	Reference
		<u>expression</u>			<u>analysis</u>	pathways or		
<u>126a-3p, 155-5p</u>	<u>qRT-PCR</u>	1	<u>Plasma</u>	Cross-sectional	<u>Bioinformatics</u> analysis	Genes liked to cancer, inflammation, apoptosis, vascular endothelial health	PAHs	<u>Ruiz-Vera et al.</u> 2019
<u>150-5p</u>	<u>qRT-PCR</u>	1	<u>Plasma</u>	Cross-sectional	<u>Bioinformatics</u> analysis	Genes linked to immune response	<u>PAHs</u>	Deng et al. 2014
<u>142-5p</u> <u>24-3p</u> <u>27a-3p</u> 28-5p						<u>Genes linked to</u> DNA damage		
93 miRNAs	Microarray	$53\uparrow 40\downarrow$	Serum	Cohort	Bioinformatics analysis	MYC, CCND1, BCL2, VEGFA	<u>PCBs</u>	Krauskopf et al. 2017
<u>191-5p</u>	<u>qRT-PCR</u>	1	Peripheral blood	Case-control	PCR	<u>AHRR, CTBP1,</u> FAS	<u>PCBs</u>	Guida et al. 2013
1537	<u>Nanostring</u> nCounter®assay	1	<u>Placenta</u>	Cohort	<u>N/P</u>		<u>PCBs</u>	Li et al. 2015
<u>15b-5p, 19a-3p, 24-</u> <u>3p, 125b-5p, let-7c</u>	Microarray	Ĺ	Follicular fluid	Cross-sectional	Bioinformatics analysis	<u>TGFβ, PI3K/Akt,</u> <u>FOXO, MAPK, p53,</u> <u>EGFR, JAK/STAT</u>	Phthalates and phenols	Martinez et al. 2019
<u>106b-5p, 374a-5p, 375</u>		¥						
<u>185 (phthalates)</u> <u>15a-5p, 142-3p</u> (phenols)	<u>qRT-PCR</u>	Ŧ	<u>Placenta</u>	<u>Cohort</u>	Bioinformatics analysis	Akt, insulin like growth factor receptor signaling, embryonic epithelial tube formation	Phthalates and phenols	La Rocca et al. 2016
<u>18 miRNAs</u>	<u>Microarray</u>	Dysregulated	<u>Placenta</u>	Case-control			Phthalates and phenols	De Felice et al. 2015
<u>146a-5p</u>	<u>qRT-PCR</u>	Ĺ			<u>Bioinformatics</u> analysis	<u>IRAK1, SORT1,</u> <u>TP53, ABL2, EGFR,</u> <u>p53, TLR</u>	<u></u>	
<u>28-5p,</u>	<u>qRT-PCR</u>	ĺ	Urine	Nested case-control	Bioinformatics analysis	acetylcholinesterase and cholinesterase activity	<u>OP</u>	Weldon et al. 2016
<u>517b-3p, 518d-3p,</u> <u>597</u>						genes involved in neurological functions		
<u>133b, 223-3p,</u>	d to miDNAs detection:					<u>N/S</u>		

*Sample type is referred to miRNAs detection; ↓/↑, difference in miRNAs expression, respectively up- and down-expressed;

 <u>N/P, not performed;</u> N/S, not specified

For peer Review Only

Table 3. Human studies on miRNAs dysregulation after heavy metals exposure and their target

<u>NA</u>	<u>Methodology</u>	<u>miRNA</u> expression	Sample type*	<u>Study design</u>	<u>miRNA target</u> analysis	<u>Signaling</u> pathways or target genes	<u>Contaminant</u>	<u>Reference</u>
	<u>q-RT-PCR</u>	Ť	<u>Plasma</u>	Cross-sectional	Literature analysis	<u>N/S</u>	As	Pérez-Vázquez et al. 2017
p <u>, 205-5p</u>	<u>q-RT-PCR</u>	*	Urine	Case-control	Literature analysis	<u>N/S</u>	As	Michailidi et al. 2015
	<u>q-RT-PCR</u>	1	<u>PBMC</u>	Case-control	Western blotting	PTEN, PDC4	As	Banerjee et al. 2017
<u>NAs</u>	Microarray	$\frac{56\uparrow}{18\downarrow}$	<u>Plasma</u>	Case-control			As	<u>Sun et al. 2017</u>
	<u>qRT-PCR</u>	1			<u>Bioinformatics</u> analysis	MAPK, JAK/STAT, chemokine pathway MAPK, FOXO MAPK MAPK, PI3K-Akt, TNF		
<u>17-5p, 20a-</u>)-5p, 26b-5p, 98-5p, 107, , 195-5p, 454- 7a-5p	<u>Microarray</u>	1 I	Cord blood	Cohort	Bioinformatics analysis	TREM1, TLR, PRKCQ	As	<u>Rager et al. 2014</u>
<u>, 107</u>	<u>qRT-PCR</u>	Ţ						
	<u>qRT-PCR</u>	1	Urine	Cross-sectional,	Literature analysis	<u>N/S</u>	<u>Pb</u>	Kong et al. 2012
<u>286</u>	Nanostring nCounter® assay	Ţ	<u>Cervix cells</u>	<u>Cohort</u>	Bioinformatics analysis	Pathways linked to cell cycle and proliferation, like AHR	<u>Pb</u>	Sanders et al. 2015
<u>p</u> , <u>431-5p, let-</u>	<u>Nanostring</u> nCounter® assay	⊥ 1	<u>Placenta</u>	<u>Cohort</u>	Literature analysis	<u>N/S</u>	<u>Pb</u>	<u>Li et al. 2015</u>
						N/C	II	_
INAS	Microorrow	<u>+</u>	Diagma	Casa control	Literatura analyi-	<u>IN/S</u>	Hg	Ding at al. 2016
<u>p</u> 5 <u>p</u>	<u>wheroarray</u>	⊥ 1	<u>Piasma</u>	<u>Case-control</u>	Literature analysis	<u>N/S</u>	<u>118</u>	<u>טווק et al. 2016</u>
		ĺ						

URL: http://mc.manuscriptcentral.com/cije Email: ijehr-els@salford.ac.uk

<u>+00 5p</u>	<u>qRT-PCR</u>	Ţ						
<u>92a-3p</u> 486-5p								
<u>15b, 21-5p, 26a-5p,</u> <u>362-3p, 182, 183-5p,</u> <u>451a,</u> 486-3p	<u>qRT-PCR</u>	Ť	Blood-leukocytes	<u>Cohort</u>	<u>Bioinformatics</u> analysis	PTEN, axonal guidance and PKA signaling	<u>Cr</u>	<u>Dioni et a</u>
18 miRNAs	<u>Microarray</u>	$13\downarrow$ 3 \uparrow	<u>Plasma</u>	Case-control			<u>Cr</u>	<u>Li et al. 2</u>
<u>590-5p</u> <u>3940-5p</u>	<u>qRT-PCR</u>	Ţ			<u>N/P</u> Enzyme- linked immunosorbent	XRCC2		
71 miRNAs	Solexa sequencing	Dysregulated	<u>Plasma</u>	Case-control	<u>4550 y</u>		(Other metals):	Deng et a
<u>16-5p, 24-3p, 27a- 3p, 28-5p, 126-3p, 142-5p, 150-5p, 320b, 451a, let-7b- 5p</u>	<u>qRT-PCR</u>	Ţ			Literature analysis	<u>N/S</u>	<u>Antimony</u>	
<u>16-5p, 320b</u>		¥					Aluminum	
<u>27a-3p</u>		Ť					<u>Pb</u>	
<u>126-3p</u>		1					Molybdenum	
<u>16-5p, 24-3p, 27a-</u> <u>3p, 28-5p, 126-3p,</u> <u>142-5p, 320b, let-7b-</u> <u>5p</u>		ĺ					<u>Tin</u>	
<u>24-3p, 27a-3p, 28-</u> <u>5p, 126-3p, 320b,</u> <u>let-7b-5p</u>		Ť					<u>Titanium</u>	
*Sample type is referred ↓ / ↑, difference in miRN N/S, not specified; N/P, not performed	to miRNAs detection; NAs expression, respect	ively up- and down-ex	pressed;					

URL: http://mc.manuscriptcentral.com/cije Email: ijehr-els@salford.ac.uk

Page 47 of 49

Table 4. List of promising miRNAs involved in environmental exposure, biological function and health implication

miRNA	Environmental	Signaling pathway or	Biological function	Health implication	<u>Reference</u>
	exposure	target genes			
<u>21-5p</u>	<u>PM, Metal rich-PM,</u> As, Cr	PTEN (inhibitor of PI3K-Akt)	Tumor suppressor, inflammation	CVD, birth defects, cancer	Bollati et al. 2010; Louwies et al. 2016; Baneriee et al. 2017; Dioni et
		MAPK	Inflammation, oxidative stress, cell proliferation, differentiation, cell survival and apoptosis		al, 2017; -Sun et al. 2017; Tsamou et al. 2018
		JAK/STAT	Inflammation, oxidative stress, cell survival and proliferation		
		<u>p53</u>	Cell proliferation and apoptosis		
		TGFβ	Embryonal development, cellular differentiation, inflammation, immune response		
<u>222-3p</u>	PM, Metal rich-PM,	PTEN (inhibitor of PI3K-Akt)	Tumor suppressor, inflammation	CVD, birth defects, reproductive	Bollati et al. 2010; Li et al. 2012; Fry
	<u>O</u> ₃	<u>MAPK</u>	Inflammation, oxidative stress, cell proliferation, differentiation, cell survival and apoptosis	disorders, respiratory diseases, cancer	<u>et al. 2014; Tsamou et al. 2018</u>
		<u>NFkB</u>	Inflammation, immune response, cell proliferation, tumorigenesis		
<u>223-3p</u>	<u>PM, Metal rich-PM,</u> O ₃	PTEN (inhibitor of PI3K-Akt)	Tumor suppressor, inflammation	<u>CVD</u> , male reproductive disorders, respiratory diseases, cancer	Li et al. 2012; Louwies et al. 2016; Rodosthenous et al. 2016
	<u> </u>	<u>NFkB,</u>	Inflammation, immune response, cell proliferation, tumorigenesis		
Let-7 family	<u>PM, Metal rich-PM,</u> Phthalates and	PTEN (inhibitor of PI3K-Akt)	Tumor suppressor, inflammation	<u>CVD</u> , male reproductive disorders, adverse female fertility outcomes.	Motta et al. 2013; Li et al. 2012; Ding et al. 2016: Rodosthenous et al. 2016:
	phenols, Hg, Other metals	<u>MAPK</u>	Inflammation, oxidative stress, cell proliferation, differentiation, cell survival and apoptosis	respiratory diseases, cancer Deng et al. 2019; Martinez et al. 2019	
		JAK/STAT	Inflammation, oxidative stress, cell survival and proliferation		
		<u>NFkB</u>	Inflammation, immune response, cell proliferation, tumorigenesis		
		<u>p53</u>	Cell proliferation and apoptosis		

URL: http://mc.manuscriptcentral.com/cije Email: ijehr-els@salford.ac.uk

1	
2	
3	<u>TGFβ</u> <u>Embryonal development, cellular</u>
4	<u>differentiation, inflammation,</u>
5	<u>Immune function</u>
6	
7	
8	
9	
10	
11	
17	
12	
13	
14	
נו 16	
10	
17	
1ð 10	
19	
20	
21	
22	
25	
24	
25	
20	
27	
20	
30	
31	
32	
32	
34	
35	
36	
37	
38	
39	
40	
41	
42	
43	
44	UKL: http://mc.manuscriptcentral.com/cije_Email: ijenr-eis@salford.ac.uk
45	
46	