

Supplementary Information

Engineering of Green Sterilization Technology to Obtain Biocompatible Aerogels: Supercritical CO₂ versus Ethylene Oxide and Gamma Radiation

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SI 1. Materials and methods

SI 1.1. Proton Quantitative Nuclear Magnetic Resonance (^1H q NMR) spectroscopy

All starch samples were prepared by dissolving about 10 mg of aerogel in 750 μL of deuterated dimethylsulfoxide-d₆ (d-DMSO), at 320K; spectra were referenced to the residual solvent peak at 2.50 ppm. Quantitative acquisition parameters for ^1H spectra: 90° pulse as calculated for each sample; relaxation delay 40.0 s. Spectral width 9615 Hz; number of transient 128, 90° pulse 7.93 μs PL1 -11.48 dB. The proton spin-lattice relaxation time (T₁) of 40s was considered longer enough to ensure accurate quantification results, according to literature data. Data processing: exponential line broadening of 0.1 Hz was applied as resolution enhancement function; zero-filling to 32 K prior FT. Acquisition parameters for ^1H spectra of alginate aerogels in D₂O: 90° pulse 8.24 μs PL1 -11.48 dB; relaxation delay 2.0 s. Spectral width 10869 Hz; number of transient 128. For the quantification of H₂O₂ entrapped within the alginate matrix aerogels, a defined amount of aerogel was incubated in d-DMSO at 40°C for 30 minutes, then the supernatant was immediately transferred in an NMR tube for data acquisition. This procedure differed from that used for starch-based aerogels due to the limited solubility of alginate in DMSO.

The H₂O₂ on the scCO₂ sterilized samples were quantified using both external and an internal standard. The external standard consisted of a solution with a known concentration of H₂O₂ in d-DMSO. A 0.153 mol·L⁻¹ stock solution was prepared by dissolving 10 μL of H₂O₂ solution in 0.75 mL of d-DMSO, and subsequent external standard solutions were obtained by serial dilution. The concentration of the external standard solutions is listed in Table S1. Among them, solution six, with a concentration of 6.13 mM/L, was selected as the external standard to determine the concentration of H₂O₂ in the aerogel samples and to verify the concentration of the prepared solutions, thereby validating the protocol. The results demonstrated excellent agreement between the prepared and experimentally measured concentrations except for solution 4, which represents the detection limit.

Table S1. H₂O₂ concentration as prepared and determined by qNMR

Solution	Prepared [mmol/L]	Experimental [mmol/L]*
1	0.078	0.079
2	1.55	1.60
3	76.50	74.21
4	4.6 10 ⁻⁵	2.05 10 ⁻⁵
5	38.25	38.28
6	6.13	
7	14.28	14.77

*Calculated with respect to solution 6, selected as external standard solution

The model was validated using a toluene solution of known molar concentration, (6.27 mmol/L), as internal standard, following the ¹H qNMR protocol on a set of randomly selected samples.

SI 2. Results

SI 2.1. H₂O₂ concentration quantification by ¹H q NMR

The results in table S2 demonstrated good agreement, confirming the reliability of the model.

Table S2. Comparison of H₂O₂ concentration by using an external and an internal standard. Conditions in bold text correspond to the selected optimized conditions.

Sample	External standard	Internal standard
	ppm	ppm
<i>STA_scCO₂Co</i>	183	182
<i>STA_scCO₂Co_V2</i>	509	-
<i>STA_scCO₂Co_V24</i>	132	131.7
<i>STA_scCO₂Co_2.5bar</i>	178	177.5
<i>STA_scCO₂Co_50bar</i>	238	-

The external standard is a solution of [H₂O₂] = 6.13 mmol/L. The internal standard is a toluene solution in d-DMSO

SI.2.2. Gel permeation chromatography of sterilized aerogel samples

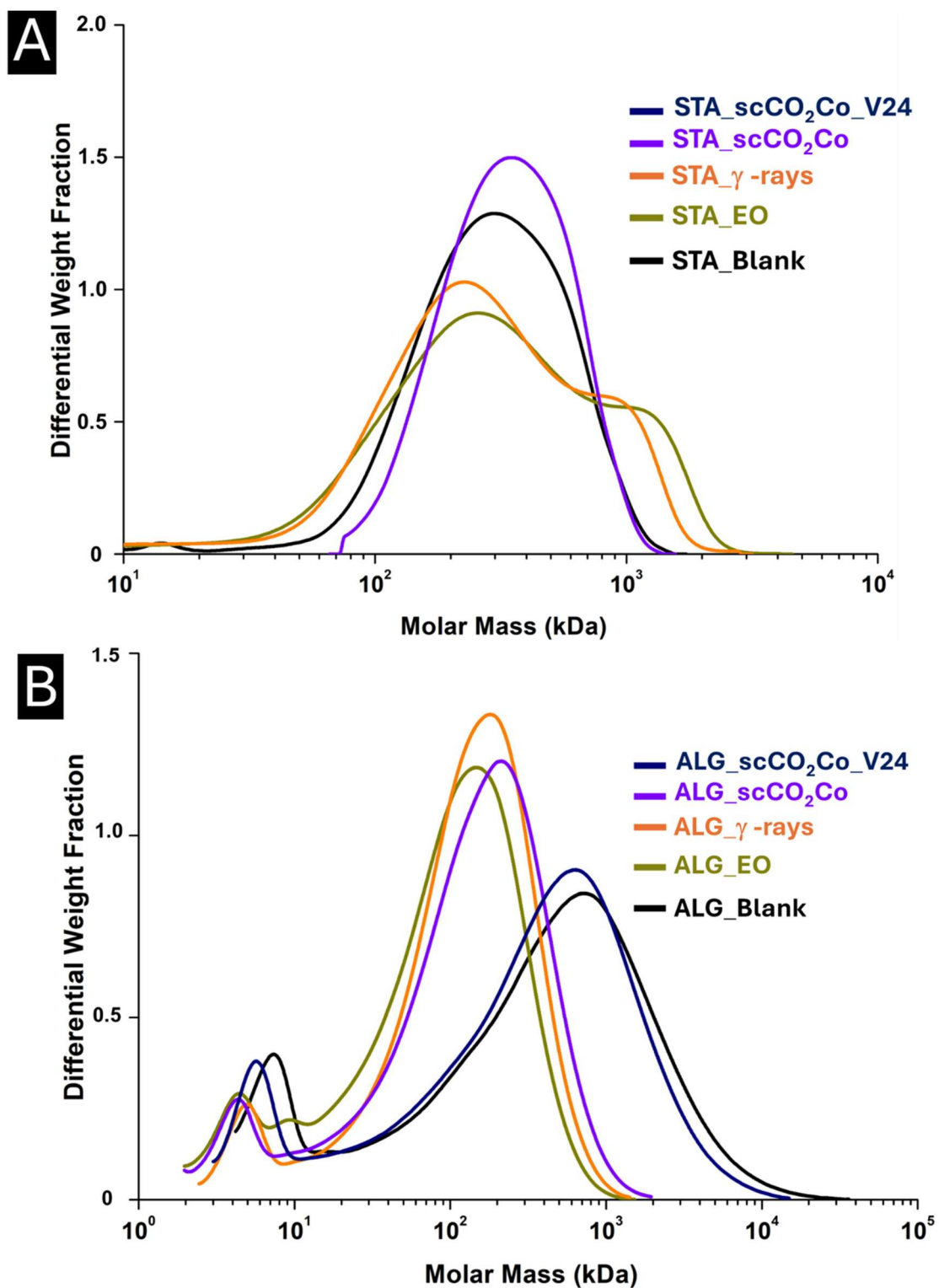


Figure SI.1. Gel permeation chromatography molecular weight distribution of sterilized (A) starch samples and (B) alginate samples