

Article

Visible Foliar Injury and Ecophysiological Responses to Ozone and Drought in Oak Seedlings

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Abstract: To verify the responses of visible foliar injury (VFI), we exposed seedlings of three oak species for 4.5 months in an open air facility, using differing ozone (O₃) and drought treatments: O₃ (three levels from ambient to ×1.4 ambient), and drought (three levels of irrigation from 40% to 100% field capacity). We related the accumulated phytotoxic O₃ dose (POD₁) and cumulative drought index (CDI) to the O₃ and drought VFI and assessed growth increment (height, diameter, leaf number), biomass (of all organs), and physiological parameters: net photosynthesis per plant (P_n), photosynthetic nitrogen (PNUE) and phosphorus use efficiency (PPUE)). The results indicated that an increase in POD₁ promoted O₃ VFI in *Quercus robur* and *Quercus pubescens*, while *Quercus ilex* was asymptomatic. The POD₁-based critical level at the onset of O₃ VFI was lower for *Q. robur* than for *Q. pubescens* (12.2 vs. 15.6 mmol m⁻² POD₁). Interestingly, drought reduced O₃ VFI in *Q. robur* but increased it in *Q. pubescens*. Both O₃ and drought were detrimental to the plant biomass. However, *Q. robur* and *Q. pubescens* invested more in shoots than in roots, while *Q. ilex* invested more in roots, which might be related to a hormetic mechanism. P_n, PNUE and PPUE decreased in all species under drought, and only in the sensitive *Q. robur* (PPUE) and *Q. pubescens* (PNUE) under O₃. This study confirms that POD₁ is a good indicator to explain the development of O₃ VFI and helps a differential diagnosis of co-occurring drought and O₃ VFI in oak forests.

Keywords: tropospheric ozone; leaf symptoms; PODy; water stress; risk assessment



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1. Introduction

Tropospheric ozone (O₃) is an oxidative pollutant harmful to plants [1]. Ozone enters the leaves through the stomata, reacts in the mesophyll, and triggers the formation of reactive oxidative species (ROS) with a cascade of events eventually promoting cell death and, finally, the appearance of visible foliar injury (VFI), physiological impairment, and growth reduction [2–4]. Furthermore, O₃ inhibits the efficient use of nutrients such as nitrogen (N) and phosphorus (P) and thereby causes a reduction of photosynthetic N and P use efficiency (PNUE and PPUE, respectively) [5,6]. Therefore, critical levels (CL) have been investigated to assess the O₃ negative impacts on several plant species, especially those related to biomass loss [7,8]. CLs are based on cumulative O₃ indexes, e.g., AOT40, defined as the accumulated exposure over 40 ppb hourly concentrations, and PODy, defined as the phytotoxic O₃ dose above an hourly threshold y of stomatal O₃ uptake [9]. PODy is considered the most realistic index with a high correlation with the detrimental effects of O₃ [10,11]. Ozone VFI is a forest-health indicator in forest monitoring programs [12].

The estimation of CL based on O₃ VFI has been proposed as a not destructive and easily repeated observation over long-term monitoring studies [13,14].

Ozone alone can affect plant growth and development, but its effect usually occurs in combination with other factors, such as drought, which is known as the most critical environmental factor limiting plant productivity worldwide [15,16]. The adverse effects of drought are progressive and, thus, are often evaluated by the cumulative drought index (CDI), defined as the accumulated difference of soil moisture relative to field capacity [17]. Drought stress also promotes the formation of specific VFI, which can be distinguished from O₃-induced foliar injury. While O₃ VFI is usually indicated by interveinal, irregular-border, yellow to dark-brown stippling [18,19], drought VFI consists in gradients of leaf margin necrosis increasing in severity from the base to the top of a plant [20], with the injury co-occurring when plants are exposed to a combination of these stress factors.

Both O₃ and drought can limit plant carbon fixation, and the effect of both stress factors has been reported as the cause of biomass loss for *Quercus* species [21,22], which are significant components of temperate forests. Previous papers from the same experiment presented here showed that the interacting factorial impacts of O₃ and drought were species-specific, and the order of O₃ sensitivity was *Q. robur* > *Q. pubescens* > *Q. ilex* from the point of view of total biomass [22] and leaf gas exchange [23,24]. Although physiological acclimations to O₃ and drought are not fully elucidated, diverse adaptation strategies were observed for tolerating stress in different oak species. One of the reasons for the variability of strategies is related to gas exchange regulation depending on their water use strategy (isohydric and anisohydric) [24]. Under elevated O₃ with sufficient water availability, the isohydric *Q. robur* limited O₃ uptake by stomatal closure, while the anisohydric *Q. ilex* and the intermediate *Q. pubescens* activated tolerance mechanisms and did not actively show a closing response of stomata. In particular, Pellegrini et al. [25] found that *Q. ilex* had a well-regulated antioxidative defense system through phenylpropanoid pathways. However, in the combination of O₃ and drought, the anisohydric *Q. ilex* and the intermediate *Q. pubescens* exhibited stomatal closure to prevent severe oxidative damage due to excess generation of ROS.

The present study aimed to characterize the VFI induced by O₃, drought, and their combination and assess their related effects on biomass, biometry, and physiological parameters. The results will help a differential diagnosis of co-occurring drought and O₃ VFI in oak forests. In detail, we addressed the following hypotheses: (1) the development of O₃ VFI may be better explained by PODy than by AOT40, (2) the reduction in soil water availability may reduce or exacerbate the negative impacts of O₃ on VFI, and (3) O₃ VFI may be an indicator to explain biomass reduction or physiological damage in Mediterranean oaks. We postulated that the effects on the development of VFI are modulated by the plant species-specific sensitivity to oxidative stressors.

2. Materials and Methods

2.1. Plant Material and Experimental Setting

The experiment was conducted in an O₃ Free-Air Controlled Exposure (FACE) facility at Sesto Fiorentino, Italy (43°48'59" N, 11°12'01" E, 55 m a.s.l.). Two-year-old plants of *Q. robur* L., *Q. pubescens* Willd., and *Q. ilex* L. were obtained from nurseries and transplanted into 10-L plastic pots. They were exposed to three levels of O₃ (1.0, 1.2, and 1.4 times the ambient air concentration, denoted as AA, ×1.2, and ×1.4, respectively: 24-h averaged concentration, AA = 35.2 ppb, ×1.2 = 42.9 ppb, ×1.4 = 48.9 ppb) and three levels of water irrigation [100, 80, and 40% of field capacity (0.295 m³ m⁻³, Paoletti et al., 2017) on average, denoted as WW-treated (well-watered), MD-treated (moderate drought) and SD-treated (severe drought), respectively]. Three replicated plots were assigned to each treatment, with three plants per combination of species, drought, and O₃. The experiment lasted for 4.5 months, from 1 June to 15 October.

The details of the FACE facility are described in Paoletti et al. [26], and the details of the experimental design are published in Hoshika et al. [27].

2.2. Evaluation of O₃ and Drought Visible Foliar Injury

Two well-trained observers evaluated the presence of O₃ and drought VFI during the experimental period for all plants for a total of 6 evaluation dates (Table 1). We applied photo guides to verify whether O₃ and/or drought VFI was present [28–30]. VFI incidence (INC = number of injured plants/total number of plants × 100) was calculated according to Chappelka et al. [31]. POD₁-based CLs and CDI-based CLs were calculated for the corresponding day when O₃ and drought VFI onset was observed.

Table 1. Phytotoxic O₃ dose (POD₁, mmol m⁻²) and accumulated exposure over 40 ppb hourly concentrations (AOT40, ppm h) calculated at the O₃ visible foliar injury onset, and Cumulative Drought Index (CDI) calculated at the drought visible foliar injury onset for *Q. robur* and *Q. pubescens*.

Water Regime	O ₃ Treat.	Onset O ₃ Injury				Onset Drought Injury	
		POD ₁		AOT40		CDI	
		<i>Q. robur</i>	<i>Q. pubescens</i>	<i>Q. robur</i>	<i>Q. pubescens</i>	<i>Q. robur</i>	<i>Q. pubescens</i>
WW-treated	AA	12.07	Asymp.	17.78	Asymp.	Asymp.	Asymp.
	×1.2	12.40	15.69	16.41	23.77	Asymp.	Asymp.
	×1.4	11.62	20.46	15.15	33.38	Asymp.	Asymp.
MD-treated	AA	13.02	Asymp.	21.74	Asymp.	6.10	6.52
	×1.2	12.13	18.41	16.41	30.43	6.10	4.04
	×1.4	13.07	20.04	21.59	33.38	4.96	6.10
SD-treated	AA	Asymp.	Asymp.	Asymp.	Asymp.	10.20	10.20
	×1.2	Asymp.	13.13	Asymp.	30.43	10.20	10.20
	×1.4	10.72	12.81	26.23	26.23	10.20	10.20

2.3. Measure of Growth Parameters

The assessment of total annual biomass production during the experiment was performed based on dry weight per plant (DW) as described in Hoshika et al. [22], additionally discriminating the below-(roots) and above-ground biomass (stem and leaves) to calculate the ratio of root to shoot biomass (Ratio R/S). Furthermore, the total number of leaves, plant height increment (measured with a metric tape) and stem caliber increment (measured just above soil level) were expressed as the absolute values relative to the values at the beginning and end of the experiment.

2.4. Assessment of Photosynthetic Parameters

The net photosynthetic rate (P_n) was previously reported for mid-summer (July: [24]) and early and late summer and autumn (June, August, and October: [23]). Here, these published data of P_n were re-analyzed to address the cumulative effects of O₃ and drought on the photosynthetic activity. The target leaves were fully sun-exposed leaves (4–6th from the shoot tip) of the plant main shoot (one representative leaf per plant, 1 to 3 plants per replicated plot per each O₃ and W treatment). Measurements were made under light-saturated conditions (1500 $\mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD [photosynthetic photon flux density]) with constant CO₂ concentration (400 $\mu\text{mol mol}^{-1}$), relative humidity (40 to 50%), and leaf temperature (25 °C) using a commercial gas exchange system (CIRAS-2 PP Systems, Herts, UK). Measurements were carried out in two campaigns (8–10 June and 27 September–6 October) for all O₃ treatments and an additional campaign (6–9 August) for two O₃ levels (1.2, and ×1.4) on days with clear sky between 9:00 and 12:00 a.m. CET. The other detailed specifications for the photosynthetic measurements were described in our previous studies [23,24].

After the measurement of P_n in August and October, leaves were collected to examine the nitrogen (N) content. Nitrogen content per unit mass (N_{mass}) was determined by the dry combustion method using a LECO TruSpec C/N analyzer (Leco Corporation, St. Joseph, MI, USA). In October, the foliar phosphorus (P) content was also determined. Phosphorus content per unit mass (P_{mass}) was examined by an inductively coupled plasma-optical emission spectroscopy (ICP-OES) (iCAP7000, Thermo Fisher Scientific, Waltham, MA,

USA). We calculated photosynthetic N use efficiency (PNUE) as the product of N_{mass} and mass-based net photosynthetic rate and photosynthetic P use efficiency (PPUE) as the product of P_{mass} and mass-based net photosynthetic rate.

2.5. Calculation of Accumulated Drought and Ozone Indexes

The accumulated drought index (CDI) was calculated from the beginning of the experimental period to the date of observation as follows:

$$\text{CDI} = \sum |Sm - Fc|$$

where, Sm is soil moisture, and Fc is field capacity ($0.295 \text{ m}^3 \text{ m}^{-3}$) [26]; drought stress is considered severe when Sm values are lower than Fc .

AOT40 and POD_1 for each O_3 and drought treatment were calculated following the parameters applied by Hoshika et al. [22] according to the methodology designed by CLRTAP (Convention on Long-range Transboundary Air Pollution) [9].

2.6. Statistical Analysis

Multiple Linear Regression (MLR) analysis was used to estimate the relationship between the O_3 indexes (AOT40 and POD_1) and CDI versus growth (height, diameter, and N. leaves), biomass (Leaf, Shoot, Root, Total, and R/S), VFI (O_3 and Drought) and physiological parameters (P_n , PNUE, and PPUE). Two models were compared, i.e., Model 1 (POD_1 and CDI as predictor variables) and Model 2 (AOT40 and CDI as predictor variables). The statistical analyses were performed using the R software (R version 4.1.2 [32]), considering a significance of $p < 0.05$. Principal component analysis (PCA) was conducted by using OriginPro 2021b software. The PCA was applied considering VFI (O_3 and drought), growth (Height, Diameter, and N. of leaves), biomass (Leaf, Shoot, Root; Total and R/S), and physiological (P_n , PNUE, and PPUE) parameters in order to distinguish the groups of parameters better related to each symptomatic species; in this analysis, the asymptomatic *Q. ilex* species was not included.

3. Results

3.1. Visible Foliar Injury

The O_3 VFI in *Q. robur* was characterized by small homogeneously distributed dots between the primary leaf veins (Figure 1A). *Q. robur* plants from all water regimes but SD (AA and $\times 1.2$) presented O_3 VFI (Tables 1 and 2). In fact, 11% of the SD-treated plants developed O_3 VFI at the end of the experiment, relative to 56% of the WW-treated plants (Table 2).

There were individual-specific differences on the day of VFI onset. The POD_1 values calculated for the O_3 VFI onset in *Q. robur* were similar across O_3 treatments (approximately 10.7 to 13.0 $\text{mmol m}^{-2} \text{POD}_1$, average = 12.1 $\text{mmol m}^{-2} \text{POD}_1$), while the AOT40 values corresponding to the O_3 VFI onset increased from 15–16 ppm h to 26.2 ppm h; for SD-treated plants, the O_3 VFI onset occurred only in $\times 1.4$ (10.7 ppm h, Table 1). In addition, the MLR revealed a positive regression of O_3 VFI with POD_1 or AOT40 and a negative regression with CDI when tested with AOT40 (Model 2), but the effect was not significant when tested with POD_1 (Model 1) (Figure 1B; Table 3).

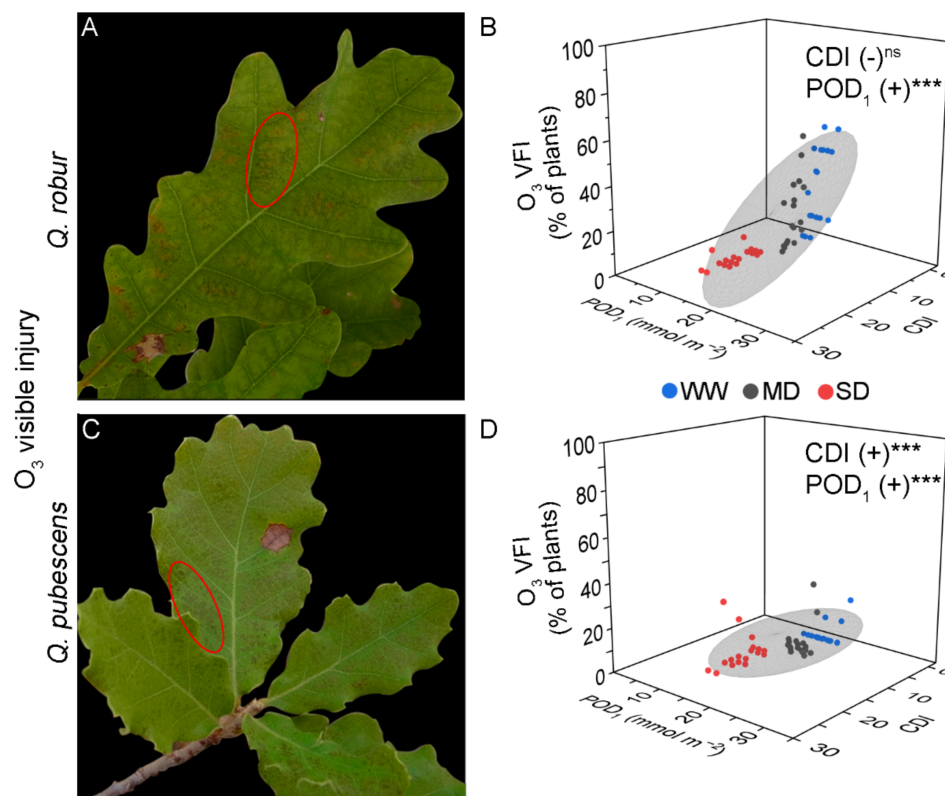


Figure 1. Illustrative examples of O₃ visible foliar injury in *Quercus robur* (A) and *Q. pubescens* (C) characterized by small homogeneously distributed dots between the primary leaf veins (ellipse). (B,D) Results from the linear multiple regression of O₃ visible foliar injury with Cumulative Drought Index (CDI) and phototoxic O₃ dose (POD₁) as predictor factors in *Quercus robur* (B) and *Q. pubescens* (D). Colored dots represent well-watered (WW-blue), moderate drought (MD-grey), and severe drought (SD-red). The grey ellipsoid represents a confidence level of 75%. (+) positive regression, *** = $p < 0.001$, ns = not significant.

Table 2. Evaluation of O₃ and drought incidence of visible foliar injury (VFI) along the experimental period for *Q. robur* and *Q. pubescens* exposed to different levels of O₃ and drought.

	Water Regime	O ₃ Treat./DOY	<i>Q. robur</i>						<i>Q. pubescens</i>					
			209	218	232	240	247	266	209	218	232	240	247	266
O ₃ VFI incidence	WW-treat.	AA	-	-	11	11	11	50	-	-	-	-	-	-
		×1.2	-	11	44	44	44	44	-	-	-	-	11	11
		×1.4	22	33	33	44	56	56	-	-	-	-	-	22
	MD-treat.	AA	-	-	-	11	22	44	-	-	-	-	-	-
		×1.2	-	22	22	22	44	56	-	-	-	-	-	33
		×1.4	-	-	22	33	33	33	-	-	-	-	-	22
SD-treat.	AA	-	-	-	-	-	-	-	-	-	-	-	-	
	×1.4	-	-	-	-	11	11	-	-	-	-	11	33	
Drought VFI incidence	WW-treat.	AA	-	-	-	-	-	-	-	-	-	-	-	-
		×1.2	-	-	-	-	-	-	-	-	-	-	-	-
		×1.4	-	-	-	-	-	-	-	-	-	-	-	-
	MD-treat.	AA	-	-	33	33	50	67	-	-	-	11	11	11
		×1.2	-	-	22	58	67	67	11	11	11	22	33	33
		×1.4	-	44	33	44	50	63	-	-	22	33	33	33
	SD-treat.	AA	22	22	67	78	83	89	22	22	39	67	67	67
		×1.2	11	56	61	78	78	89	11	11	22	33	44	44
		×1.4	33	56	78	78	100	100	44	44	67	78	78	78

Table 3. Regression coefficients of the multiple linear regression for the species *Q. robur*, *Q. pubescens*, and *Q. ilex*, considering Cumulative Drought Index (CDI) and phototoxic O₃ dose (POD₁) for Model 1, and CDI and accumulated exposure over 40 ppb hourly concentrations (AOT40) for Model 2 as predictor factors, and Growth: Plant height increment (cm), Stem diameter increment (cm), and leaf number increment (N. leaves—n); Biomass: Leaf (g), Shoot (g), Root (g), Total biomass (g), and Ratio root/shoot (Ratio R/S); Visible foliar injury (O₃ and drought—% of plants); Physiological parameters: Photosynthesis (P_n—μmol m⁻²s⁻¹), Photosynthetic nitrogen use efficiency (PNUE—μmol m⁻²s⁻¹) and Photosynthetic phosphorus use efficiency (PPUE—μmol m⁻²s⁻¹) as dependent parameters. Levels of significance (*p*), intercepts and determination coefficients (R²) are shown.

Parameters	Model 1 (POD ₁ , CDI)						Model 2 (AOT40, CDI)										
	POD ₁	<i>p</i>	CDI	<i>p</i>	Intercept	<i>p</i>	R ²	AOT40	<i>p</i>	CDI	<i>p</i>	Intercept	<i>p</i>	R ²			
<i>Q. robur</i>	Injury	O ₃	6.736	***	-0.043	n.s.	-64.967	***	0.717	0.002	***	-2.027	***	-20.600	***	0.738	
		Drought	4.802	***	5.882	***	-68.059	***	0.887	0.001	***	4.524	***	-31.350	***	0.861	
	Growth	Height	0.316	**	0.035	n.s.	-1.197	n.s.	0.308	0.159	***	-0.041	n.s.	-0.213	n.s.	0.501	
		Diameter	0.032	n.s.	-0.051	**	5.182	***	0.323	0.019	n.s.	-0.059	***	5.214	***	0.338	
		N. Leaves	0.068	n.s.	-2.338	***	190.955	***	0.521	0.003	n.s.	-2.351	***	191.913	***	0.521	
	Biomass	Leaf	0.240	***	0.013	n.s.	1.741	n.s.	0.404	0.068	*	-0.040	n.s.	3.743	***	0.191	
		Shoot	-0.043	n.s.	-0.176	***	12.517	***	0.404	-0.017	n.s.	-0.166	***	12.268	***	0.404	
		Root	-0.784	**	-0.568	***	40.098	***	0.508	-0.287	*	-0.391	***	35.097	***	0.476	
	Physiology	Total	-0.588	n.s.	-0.732	***	54.357	***	0.428	-0.236	n.s.	-0.597	***	51.108	***	0.427	
		Ratio R/S	-0.063	***	-0.019	***	2.600	***	0.692	-0.022	***	-0.005	n.s.	2.163	***	0.491	
		P _n	-0.629	***	-0.283	***	14.579	***	0.824	0.000	***	-0.152	**	11.930	***	0.738	
		PNUE	-0.569	n.s.	-0.307	*	14.724	*	0.558	-0.152	n.s.	-0.175	*	10.032	**	0.558	
		PPUE	-3.751	*	-1.560	**	82.525	**	0.916	-0.668	n.s.	-0.671	*	41.067	*	0.767	
	<i>Q. pubescens</i>	Injury	O ₃	1.827	***	0.645	***	-26.982	***	0.329	0.876	***	0.090	n.s.	-16.377	***	0.387
			Drought	1.279	*	3.658	***	-23.673	*	0.787	0.660	*	3.259	***	-17.183	**	0.792
Growth		Height	0.241	***	-0.001	n.s.	-0.472	n.s.	0.373	0.115	***	-0.060	*	1.039	n.s.	0.371	
		Diameter	-0.151	***	-0.052	***	8.020	***	0.485	-0.052	**	-0.017	n.s.	6.595	***	0.244	
		N. Leaves	-4.901	***	-1.801	**	175.047	***	0.390	-2.011	**	-0.644	n.s.	136.376	***	0.283	
Biomass		Leaf	-0.019	n.s.	-0.012	n.s.	4.183	***	-0.070	-0.001	n.s.	-0.008	n.s.	3.865	***	-0.075	
		Shoot	-0.205	**	-0.130	***	13.137	***	0.511	-0.091	**	-0.081	**	11.687	***	0.474	
		Root	-0.054	n.s.	-0.223	***	22.085	***	0.366	-0.039	n.s.	-0.208	***	22.072	***	0.372	
Physiology		Total	-0.278	n.s.	-0.364	***	39.404	***	0.400	-0.132	n.s.	-0.297	***	37.625	***	0.398	
		Ratio R/S	-0.022	*	-0.017	***	2.132	***	0.316	-0.009	n.s.	-0.012	**	1.955	***	0.277	
		P _n	-0.370	***	-0.255	***	13.500	***	0.795	0.000	***	-0.179	***	12.290	***	0.771	
		PNUE	-0.485	*	-0.286	**	14.671	**	0.741	-0.173	*	-0.172	**	10.622	***	0.757	
		PPUE	-2.385	*	-1.389	**	70.714	**	0.954	-0.575	n.s.	-0.788	**	41.724	**	0.901	
<i>Q. ilex</i>		Injury	O ₃	0.127	n.s.	0.004	n.s.	15.423	***	-0.050	0.051	n.s.	-0.026	n.s.	15.623	***	-0.357
			Drought	0.153	**	0.031	*	0.799	n.s.	0.305	0.042	**	-0.003	n.s.	1.483	***	0.229
	Growth	Diameter	3.238	**	1.194	**	17.246	n.s.	0.329	1.218	***	0.445	n.s.	23.955	**	0.468	
		N. Leaves	-0.151	n.s.	-0.035	n.s.	5.825	***	0.022	-0.029	n.s.	-0.002	n.s.	4.846	***	-0.037	
		Leaf	0.011	n.s.	-0.053	n.s.	7.619	***	0.158	-0.000	n.s.	-0.055	*	7.750	***	0.157	
	Biomass	Shoot	0.453	**	0.064	n.s.	7.027	***	0.230	0.072	n.s.	-0.031	n.s.	10.315	***	0.013	
		Root	0.313	n.s.	0.024	n.s.	20.471	***	0.038	0.043	n.s.	-0.089	n.s.	22.911	***	-0.005	
		Total	0.095	*	0.012	n.s.	0.158	n.s.	0.294	0.022	n.s.	-0.008	n.s.	0.7669	n.s.	0.068	
	Physiology	Ratio R/S	0.094	n.s.	-0.173	***	9.090	***	0.630	0.000	n.s.	-0.196	***	9.380	***	0.624	
		P _n	-0.014	n.s.	-0.122	**	5.768	**	0.840	-0.014	n.s.	-0.118	***	5.939	***	0.846	
		PNUE	-0.014	n.s.	-0.122	**	5.768	**	0.840	-0.014	n.s.	-0.118	***	5.939	***	0.846	
		PPUE	-3.036	n.s.	-1.870	*	96.073	*	0.814	-0.478	n.s.	-1.124	*	71.158	*	0.772	

* = *p* < 0.05, ** = *p* < 0.01, *** = *p* < 0.001, n.s. = not significant.

The O₃ VFI in *Q. pubescens* was similar to that developed by *Q. robur* (Figure 1C). Independently of the water regime, plants from AA did not show O₃ VFI, while plants from ×1.2 and ×1.4 treatments presented O₃ VFI (Tables 1 and 2). The percentage of plants presenting VFI was lower than for *Q. robur*, with a maximum of 33% presenting VFI (Table 2). VFI occurred for the first time at DOY 247 or 266, i.e., around the end of the experiment. The POD₁ and AOT40 values for the O₃ VFI onset were 12.8–20.46 mmol m⁻² POD₁ (average = 16.8 mmol m⁻² POD₁) and 24–33 ppm h AOT40, respectively (Table 1). The MLR revealed a positive regression with the O₃ indexes (POD₁ and AOT40; Figure 1D, Table 3). Interestingly, CDI positively affected the O₃ VFI when tested with POD₁ (Model 1), but the effect was not significant when tested with AOT40 (Model 2) (Table 3).

The drought VFI of *Q. robur* was evident exclusively on the leaf edge that became dry and brownish (Figure 2A). The VFI progressively increased in MD- and SD-treated plants until the end of the experimental period, while WW-treated plants did not show any injury (Table 2). At the end of the experiment, 89–100% of the SD-treated plants showed drought VFI, relative to 63–67% of the MD-treated plants (Table 2). The CDI calculated at the drought VFI onset was the same for all SD-treated plants (CDI = 10.20 for all AA, ×1.2, ×1.4 treatments, Table 1) and similar for MD-treated plants (CDI= 4.96 to 6.10, Table 1). The MLR revealed a strong positive regression between the *Q. robur* drought VFI and CDI, although POD₁ or AOT40 also increased the extent of drought VFI (Figure 2B, Table 3).

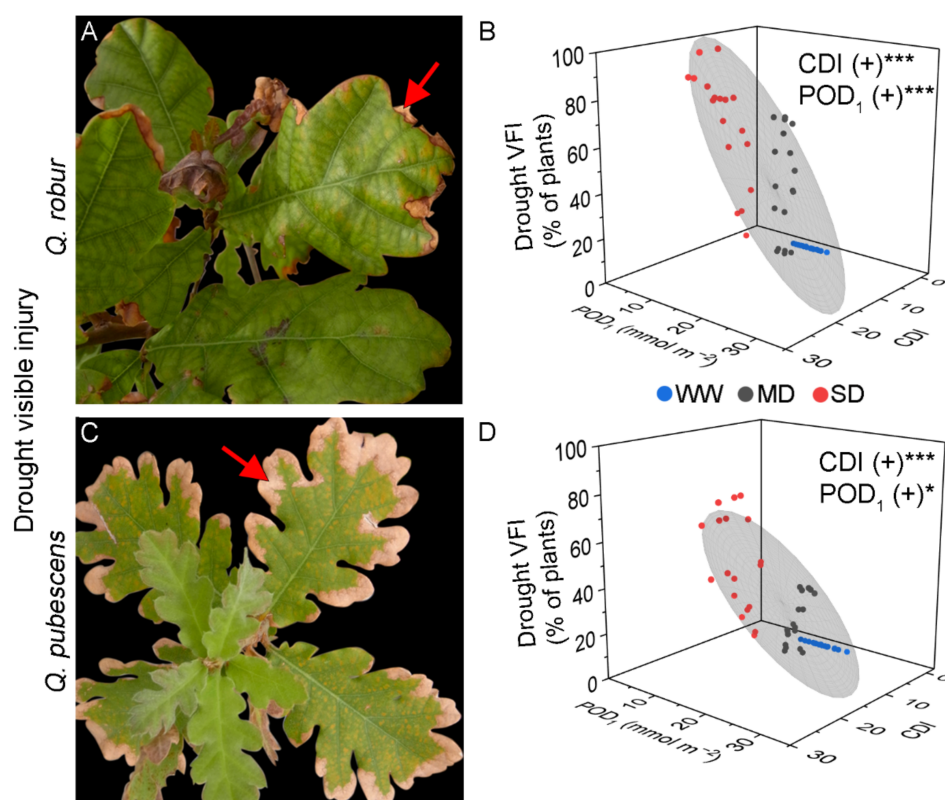


Figure 2. Illustrative examples of drought visible foliar injury in *Q. robur* (A) and *Q. pubescens* (C) characterized by dry and brownish leaf edges (arrow). (B,D) Results from the linear multiple regression of drought visible foliar injury with Cumulative Drought Index (CDI) and phototoxic O₃ dose (POD₁) as predictor factors in *Q. robur* (B) and *Q. pubescens* (D). Colored dots represent well-watered (WW—blue), moderate drought (MD—grey), and severe drought (SD—red). The grey ellipsoid represents a confidence level of 75%. (+) positive regression, * = $p < 0.05$, *** = $p < 0.001$, ns = not significant.

The drought VFI of *Q. pubescens* was similar to that developed by *Q. robur* (Figure 2C). At the end of the experimental period, *Q. pubescens* presented 44–78% of the SD-treated plants with VFI, 11–33% of the MD-treated plants, and no VFI for the WW-treated plants. As found in *Q. robur*, the CDI calculated at the drought VFI onset was the same for all SD-treated plants (CDI = 10.20 for all AA, $\times 1.2$, $\times 1.4$ treatments) and similar for MD-treated plants (CDI = 4.04 to 6.52, Table 1). Interestingly, the CDI values at drought VFI onset were similar in *Q. robur* and *Q. pubescens* within the same O₃ and drought treatments (Table 1). In addition, the MLR revealed a positive regression with CDI, POD₁, and AOT40 (Table 3, Figure 2D). The evergreen *Q. ilex* did not present O₃ or drought VFI.

3.2. Physiological Responses

In both *Q. robur* and *Q. pubescens*, P_n was negatively affected by POD₁ and CDI (Table 3, Figure S1B and D), but it unexpectedly increased with increasing AOT40 (Table 3). Furthermore, PNUE and PPUE were negatively related to CDI and POD₁ except for PNUE in *Q. robur* (Table 3).

For *Q. ilex*, the MLR revealed that CDI negatively affected P_n (Figure S1F), PNUE, and PPUE, with no significant relationship with the O₃ indexes (POD₁ and AOT40, Table 3).

3.3. Growth and Biomass

The MLR indicated that height increment was positively affected by POD₁ or AOT40 in *Q. robur*, while increments of diameter and number of leaves were negatively affected by CDI (Table 3). As confirmed by negative regression coefficients with CDI, most biomass

parameters of *Q. robur* were reduced by drought. On the other hand, POD_1 or $AOT40$ positively affected leaf biomass and negatively affected root biomass, indicating a reduction of the R/S ratio under elevated O_3 exposure (Table 3, Figure S1A).

In *Q. pubescens*, the O_3 indexes (POD_1 and $AOT40$) were positively related to plant height increment, while CDI was negatively related to height only when tested with $AOT40$ (Table 3). Increments in shoot diameter and number of leaves in this species were negatively related to POD_1 and $AOT40$, while they were negatively related to CDI when tested with POD_1 (Model 1, Table 3). Regarding the biomass parameters, leaf biomass was not affected by any factor, while shoot biomass was negatively affected by both O_3 indexes (POD_1 and $AOT40$) and CDI (Table 3). Root and total biomass were negatively related to CDI, and the R/S ratio was negatively influenced by CDI and POD_1 (Table 3, Figure S1C).

In *Q. ilex*, plant height increment was not affected by any factors, while a positive relationship between diameter increments and O_3 indexes was found (Table 3). The increment in the number of leaves was positively affected by POD_1 and $AOT40$ and positively affected by CDI when tested together with POD_1 (Table 3), although leaf and total biomass were not significantly affected by those factors. Shoot biomass was negatively affected by CDI only when tested with $AOT40$, and root biomass was positively affected only by POD_1 (Table 1). The R/S ratio was positively related only to POD_1 (Table 3, Figure S1E).

The raw data off all growth parameters for the species *Q. robur*, *Q. pubescens*, and *Q. ilex* are available in Table S1.

3.4. Principal Component Analysis

The PCA detected separate multivariate spaces between the two symptomatic species as groups related to different growth, biomass, and physiological parameters related to O_3 or drought VFI (Figure 3).

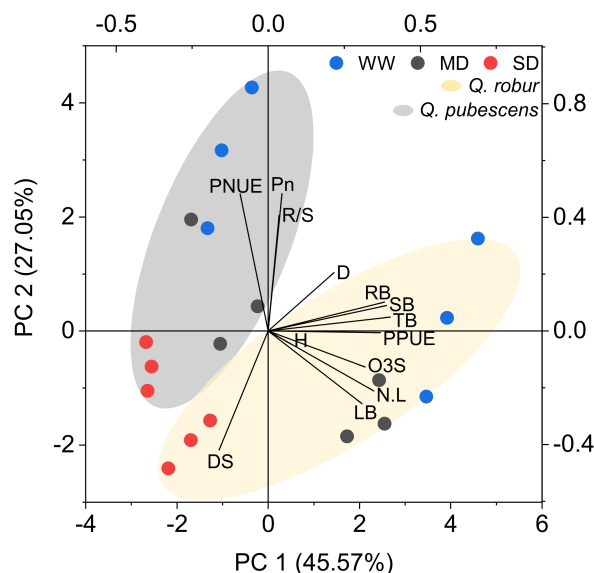


Figure 3. Bi-plot of the principal component analysis on Growth: Plant height increment (H), Stem diameter increment (S), and leaf number increment (N.L); Biomass: Leaf (L), Shoot (S), root (R), Total biomass (TB), and shoot/root ratio (R/S); Visible foliar O_3 (O_3S) and drought injury (DS); Physiological parameters: Photosynthesis (P_n), Photosynthetic nitrogen use efficiency (PNUE) and Photosynthetic phosphorus use efficiency (PPUE).

Since *Q. ilex* did not show VFI, this species was not included in the analysis. The first two components of the PCA explained 45.57 and 27.05% of the variances. The SD-treated plants of both species were grouped near the drought VFI (DS) with no other parameter following the same vector direction. The individuals of *Q. robur* (especially MD-treated

plants) were grouped near the vectors of the growth parameters number of leaves and height, leaf biomass, and O₃ VFI, which presented the same direction, thus indicating that when O₃ VFI increased, these parameters also increased. The individuals of *Q. pubescens* (specially WW-treated plants) were grouped near P_n , PNUE, and R/S, with the vectors in the opposite direction of O₃ and drought VFI, thus indicating that when O₃ and drought VFI increased, these parameters decreased.

4. Discussion

4.1. Development of Visible Injury Due to Ozone and Drought Stress

The POD₁ values corresponding to the onset of O₃ VFI for the two symptomatic deciduous oaks (on average, 14.4 mmol m⁻²) were similar to those estimated for broadleaf species under field conditions in Italy and France (10 mmol m⁻² s⁻¹) [10]. However, the CL for the VFI onset was lower for *Q. robur* than for *Q. pubescens*, indicating its higher sensitivity to O₃, possibly related to its lower antioxidative capacity and inability to protect the cell structure [25]. Furthermore, O₃ VFI increased with increasing POD₁ in the two deciduous oaks. This suggests that POD₁ is a key indicator to describe the development of O₃ VFI [33] once it is well known that O₃ damage is closely related to stomatal O₃ uptake [1]. In fact, the absence of O₃ VFI in *Q. ilex* might be related to its low g_{max} (165 mmol O₃ m⁻² s⁻¹, compared to 225 mmol O₃ m⁻² s⁻¹ and 200 mmol O₃ m⁻² s⁻¹ of *Q. pubescens* and *Q. robur*, respectively, [22] suggesting that the development of VFI might be discussed in terms of the specific-species patterns of stomatal conductance.

For both injured species (*Q. robur* and *Q. pubescens*) at the end of the experimental period, the severe drought treatment reduced POD₁ by 30 to 40% [22], which would be expected to decrease the O₃-induced VFI in plants as reported before for ecophysiological responses in poplars [34]. In *Q. robur*, the presence of O₃ VFI was decreased under drought. On the contrary, drought stress aggravated the O₃ VFI in *Q. pubescens*. Drought has been reported to have the potential to aggravate the harmful effects of O₃ [35]. Furthermore, Hoshika et al. [24] found that the combination of O₃ and drought altered the activity of the antioxidant system so that *Q. pubescens* was not protected from the severe oxidative stress resulting from the combined stress of O₃ and drought.

For the symptomatic species (*Q. robur* and *Q. pubescens*), the progression of drought VFI could be attributed to the obstruction of conducting tissue [20], conferring to both species a high sensitivity. In the asymptomatic *Q. ilex*, this phenomenon might not happen due to its capacity to increase the cell wall thickness by reinforcing the strength and rigidity of the secondary cell walls with hemicellulose and lignin deposition (data not published). Changes in lignin might function as physical desiccation tolerance and maintenance of protein integrity in drought-tolerant species [36], thus helping the photosynthetic recovery activity after re-watering from severe drought episodes [37]. The CDI threshold for the appearance of drought VFI in the two symptomatic species was higher in SD-treated than MD-treated plants, possibly due to the interaction with leaf aging, which is an important physiological and biochemical defense factor against drought stress [38]. In fact, most plants showed drought VFI in mid- or late-summer in both SD-treated and MD-treated plants when leaves were relatively old.

4.2. Effects of Ozone and Drought Stress on Growth and Biomass Parameters

For both deciduous species (*Q. robur* and *Q. pubescens*), height increment was higher when exposed to O₃ treatment. This phenomenon was verified in other species, such as *Populus* sp. [39], and it is possibly related to promoting a new leaf development as a compensative response against O₃ damage. However, the decrease in the number of leaves was eventually found to be due to O₃ exposure, which might be related to the potential O₃ phytotoxicity that triggers programmed cell death, promoting an increase in leaf senescence [40]. When combined with drought stress, the effect can be more substantial once the lack of water and nutrients promotes a decrease in new leaf development.

Both O₃ and drought stresses were detrimental to the plant biomass increment in all the oak species. In fact, the reduction of biomass due to drought stress is reported for many species and is related to the reduction of water content, diminished leaf water potential and turgor loss, promotion of stomatal closure, and decreased cell enlargement growth [41,42]. As previously revealed by Alonso et al. [21], drought stress does not protect holm oak from O₃ effects when considering the whole plant response. However, differences between the species responses must be considered when comparing the species sensitivity. For example, we observed that *Q. robur* and *Q. pubescens* invested more in shoots than in roots when exposed to both stresses, while *Q. ilex* performed the opposite, which might be another strategy of *Q. ilex* indicating a hormetic mechanism of tolerance for increasing conducting tissue and maintaining the water flow. These tolerance mechanisms may be associated with morphological/anatomical adjustments, such as a versatile root system, conservative growth and carbon allocation patterns, and diverse adaptations in the leaf morphology [20,43]. This might increase the apoplastic water fraction [44] and promote the species tolerance to O₃ and drought stress.

4.3. Effects of Ozone and Drought Stress on Physiological Parameters

The O₃ and drought stress negatively affected the physiological parameters. Drought stress induced a decrease of P_n regardless of the different species, as confirmed by a negative relationship with CDI. A decrease in P_n with increasing POD₁ was verified for both sensitive species (*Q. robur* and *Q. pubescens*), while no such reduction was found in *Q. ilex*. The present discussion is based on the species responses to POD₁ once the flux-based index is more realistic [9]. In fact, AOT40 was positively related to P_n in the two deciduous oaks, which does not agree with a consensus about O₃ negatively affecting photosynthetic capacity [45]. In fact, the regression coefficient was very low (=0.000), although the regression slope was numerically significant. Even though data was generated from an underlying distribution, the significance is a rather unlikely biological sense. The data suggest that a biological-sound index such as POD₁ is superior to AOT40 for the studies of the O₃ effects on vegetations because it can consider the principal physiological cause of O₃ damage, i.e., stomatal O₃ uptake.

Drought stress decreased PNUE and PPUE for all three species, while O₃ stress negatively affected PNUE for *Q. pubescens* and PPUE for the sensitive species *Q. robur* and *Q. pubescens*. Drought stress is directly related to changes in the allocation of N and P to leaves, no matter the species sensitivity to O₃ stress. However, a reduced allocation of N and P to the photosynthetic apparatus [5,6,46] is more pronounced in O₃ sensitive species. The N-uptake efficiency and leaf N efficiency are important traits to improve growth under drought [47]; thus, the decline in root biomass might explain the decrease in PNUE and PPUE for those species, once reduced quantity of absorptive roots reduces water and nutrient uptake as verified for the same oak species in a previous study [48].

4.4. Is the Ozone Visible Injury an Indicator to Explain Biomass Reduction or Physiological Damage in Mediterranean Oaks?

The PCA biplot contains the strength of VFI, physiology, and growth relationships, along with the species-specific sensitivity to drought and O₃ stress. Relationships between O₃ VFI and biomass growth were discussed by other authors [49,50]. In the present study, we observed that the vector of O₃ VFI injury (O₃S) and total biomass (TB) were crossing at the right angle of each other, suggesting a weak association between these two parameters in Mediterranean oaks. However, the O₃S vector shows the same direction as those of leaf parameters (number of leaves [N.L] and leaf biomass [LB]) in plants presenting more O₃ VFI, which may indicate the promotion of carbon allocation to leaves as a compensation response against O₃ injury. In addition, opposite directions of the vectors were found for O₃ VFI (O₃S) and net photosynthesis (P_n), PNUE, and the R/S ratio, highlighting a negative correlation between O₃ VFI and these parameters. The results indicate that O₃ VFI was not a direct indicator of biomass reduction under elevated O₃ in these oaks but provides

important insights regarding the impairment of photosynthetic capacity and biomass partitioning to roots. Mediterranean oak species generally develop taproots that grow deep into the soil, enhancing resistance to abiotic stress such as drought [51]. However, small amounts of roots due to O₃ exposure imply a loss of water and nutrient uptake, suggesting that O₃ VFI should be considered a bioindicator in forests exposed to the combination of O₃ pollution and drought.

5. Conclusions

We examined O₃- and drought-induced VFI and their effects on growth, biomass, and physiological parameters by using cumulative indexes and oak species known for showing differential sensitivity to these stressors. The increase in POD₁ promoted the development of specific O₃ VFI in the isohydric *Q. robur* and the intermediate *Q. pubescens*, while the anisohydric *Q. ilex* was asymptomatic. In *Q. robur*, the presence of O₃ VFI was decreased under drought probably because drought-induced stomatal closure reduced O₃ uptake and thus limited O₃ damage. However, drought stress aggravated O₃ VFI in *Q. pubescens*. This result indicates the importance of the protective role of antioxidant activity under the combination of O₃ and drought, which may be weakened by the combined stress factors and become a dominant factor in species that are not strictly isohydric. On the other hand, the drought VFI was clearly distinguished from the O₃-induced VFI, and it developed with increasing CDI in *Q. robur* and *Q. pubescens* but not in *Q. ilex*, suggesting a high tolerance of *Q. ilex* to drought stress. Therefore, we suggest using the specific O₃ or drought VFI as a bioindicator, especially for establishing the onset injury CL.

We also confirmed that P_n was decreased progressively with POD₁ and CDI in the two deciduous oaks, in tandem with PNUE decline, suggesting a cumulative effect of O₃ and drought on photosynthetic capacity. As a result, both stress factors showed a deleterious effect on the development of VFI and biomass growth. Interestingly, the two deciduous oaks increased the allocation to shoot growth rather than to root growth when exposed to both stresses, while an opposite result was found in *Q. ilex*. The imbalance in carbon allocation to roots may reduce the stability against strong winds and impair water uptake under the warming climate expected in future climate change [52,53].

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/plants11141836/s1>, Figure S1: Results of the multiple linear regression for shoot/root (Ratio R/S) and Photosynthesis (P_n) parameters; Table S1: Raw data of growth parameters.

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References

1. Mills, G.; Pleijel, H.; Malley, C.S.; Sinha, B.; Cooper, O.R.; Schultz, M.G.; Neufeld, H.S.; Simpson, D.; Sharps, K.; Feng, Z.; et al. Tropospheric Ozone Assessment Report: Present-Day Tropospheric Ozone Distribution and Trends Relevant to Vegetation. *Elementa* **2018**, *6*, 47. [[CrossRef](#)]
2. Paoletti, E. Ozone Impacts on Forests. *CAB Rev. Perspect. Agric. Vet. Sci. Nutr. Nat. Resour.* **2007**, *2*, 13. [[CrossRef](#)]
3. Dusart, N.; Gandin, A.; Vaultier, M.N.; Joffe, R.; Cabané, M.; Dizengremel, P.; Jolivet, Y. Importance of Detoxification Processes in Ozone Risk Assessment: Need to Integrate the Cellular Compartmentation of Antioxidants? *Front. For. Glob. Chang.* **2019**, *2*, 45. [[CrossRef](#)]
4. Grulke, N.E.; Heath, R.L. Ozone Effects on Plants in Natural Ecosystems. *Plant Biol.* **2020**, *22*, 12–37. [[CrossRef](#)] [[PubMed](#)]
5. Watanabe, M.; Hoshika, Y.; Inada, N.; Wang, X.; Mao, Q.; Koike, T. Photosynthetic Traits of Siebold's Beech and Oak Saplings Grown under Free Air Ozone Exposure in Northern Japan. *Environ. Pollut.* **2013**, *174*, 50–56. [[CrossRef](#)] [[PubMed](#)]
6. Hoshika, Y.; Brilli, F.; Baraldi, R.; Fares, S.; Carrari, E.; Zhang, L.; Badea, O.; Paoletti, E. Ozone Impairs the Response of Isoprene Emission to Foliar Nitrogen and Phosphorus in Poplar. *Environ. Pollut.* **2020**, *267*, 115679. [[CrossRef](#)]
7. Büker, P.; Feng, Z.; Uddling, J.; Briolat, A.; Alonso, R.; Braun, S.; Elvira, S.; Gerosa, G.; Karlsson, P.E.; Le Thiec, D.; et al. New Flux Based Dose Response Relationships for Ozone for European Forest Tree Species. *Environ. Pollut.* **2015**, *206*, 163–174. [[CrossRef](#)]
8. Hoshika, Y.; Paoletti, E.; Agathokleous, E.; Sugai, T.; Koike, T. Developing Ozone Risk Assessment for Larch Species. *Front. For. Glob. Chang.* **2020**, *3*, 45. [[CrossRef](#)]
9. CLRTAP. Mapping Critical Levels for Vegetation, Chapter III. Manual on Methodologies and Criteria for Modelling and Mapping Critical Loads and Levels and Air Pollution Effects, Risks and Trends. In *UNECE Convention on Long-Range Transboundary Air Pollution*; UNECE: Geneva, Switzerland, 2017.
10. Sicard, P.; De Marco, A.; Carrari, E.; Dalstein-Richier, L.; Hoshika, Y.; Badea, O.; Pitar, D.; Fares, S.; Conte, A.; Popa, I.; et al. Epidemiological Derivation of Flux-Based Critical Levels for Visible Ozone Injury in European Forests. *J. For. Res.* **2020**, *31*, 1509–1519. [[CrossRef](#)]
11. Bagard, M.; Jolivet, Y.; Hasenfratz-Sauder, M.-P.; Gérard, J.; Dizengremel, P.; Le Thiec, D. Ozone Exposure and Flux-Based Response Functions for Photosynthetic Traits in Wheat, Maize and Poplar. *Environ. Pollut.* **2015**, *206*, 411–420. [[CrossRef](#)]
12. Schaub, M.; Calatayud, V.; Ferretti, M.; Brunialti, G.; Lövblad, G.; Krause, G.; Sanz, M.J. Part VIII: Monitoring of Ozone Injury. In *Manual on Methods and Criteria for Harmonized Sampling, Assessment, Monitoring and Analysis of the Effects of Air Pollution on Forests*; UNECE ICP Forests Programme Coordinating Centre, Ed.; Thünen Institute of Forest Ecosystems: Eberswalde, Germany, 2016; p. 14.
13. Sicard, P.; De Marco, A.; Dalstein-Richier, L.; Tagliaferro, F.; Renou, C.; Paoletti, E. An Epidemiological Assessment of Stomatal Ozone Flux-Based Critical Levels for Visible Ozone Injury in Southern European Forests. *Sci. Total Environ.* **2016**, *541*, 729–741. [[CrossRef](#)]
14. Sicard, P.; Hoshika, Y.; Carrari, E.; De Marco, A.; Paoletti, E. Testing Visible Ozone Injury within a Light Exposed Sampling Site as a Proxy for Ozone Risk Assessment for European Forests. *J. For. Res.* **2021**, *32*, 1351–1359. [[CrossRef](#)]
15. Hanjra, M.A.; Qureshi, M.E. Global Water Crisis and Future Food Security in an Era of Climate Change. *Food Policy* **2010**, *35*, 365–377. [[CrossRef](#)]
16. Xu, C.; McDowell, N.G.; Fisher, R.A.; Wei, L.; Sevanto, S.; Christoffersen, B.O.; Weng, E.; Middleton, R.S. Increasing Impacts of Extreme Droughts on Vegetation Productivity under Climate Change. *Nat. Clim. Chang.* **2019**, *9*, 948–953. [[CrossRef](#)]
17. Peters, M.P.; Iverson, L.R.; Matthews, S.N. *Spatio-Temporal Drought Trends by Forest Type in the Conterminous United States, 1960–2013*; US Department of Agriculture, Forest Service, Northern Research Station: Madison, WI, USA, 2014.
18. Vollenweider, P.; Ottiger, M.; Günthardt-Goerg, M.S. Validation of Leaf Ozone Symptoms in Natural Vegetation Using Microscopical Methods. *Environ. Pollut.* **2003**, *124*, 101–118. [[CrossRef](#)]
19. Moura, B.B.; Alves, E.S.; Marabesi, M.A.; de Souza, S.R.; Schaub, M.; Vollenweider, P. Ozone Affects Leaf Physiology and Causes Injury to Foliage of Native Tree Species from the Tropical Atlantic Forest of Southern Brazil. *Sci. Total Environ.* **2018**, *610–611*, 912–925. [[CrossRef](#)]
20. Vollenweider, P.; Menard, T.; Arend, M.; Kuster, T.M.; Günthardt-Goerg, M.S. Structural Changes Associated with Drought Stress Symptoms in Foliage of Central European Oaks. *Trees—Struct. Funct.* **2016**, *30*, 883–900. [[CrossRef](#)]
21. Alonso, R.; Elvira, S.; González-Fernández, I.; Calvete, H.; García-Gómez, H.; Bermejo, V. Drought Stress Does Not Protect *Quercus Ilex* L. from Ozone Effects: Results from a Comparative Study of Two Subspecies Differing in Ozone Sensitivity. *Plant Biol.* **2014**, *16*, 375–384. [[CrossRef](#)]
22. Hoshika, Y.; Moura, B.; Paoletti, E. Ozone Risk Assessment in Three Oak Species as Affected by Soil Water Availability. *Environ. Sci. Pollut. Res.* **2018**, *25*, 8125–8136. [[CrossRef](#)]
23. Cocozza, C.; Paoletti, E.; Mrak, T.; Zavadlav, S.; Levanič, T.; Kraigher, H.; Giovannelli, A.; Hoshika, Y. Isotopic and Water Relation Responses to Ozone and Water Stress in Seedlings of Three Oak Species with Different Adaptation Strategies. *Forests* **2020**, *11*, 864. [[CrossRef](#)]
24. Hoshika, Y.; Fares, S.; Pellegrini, E.; Conte, A.; Paoletti, E. Water Use Strategy Affects Avoidance of Ozone Stress by Stomatal Closure in Mediterranean Trees—A Modelling Analysis. *Plant Cell Environ.* **2020**, *43*, 611–623. [[CrossRef](#)]

25. Pellegrini, E.; Hoshika, Y.; Dusart, N.; Cotrozzi, L.; Gérard, J.; Nali, C.; Vaultier, M.N.; Jolivet, Y.; Lorenzini, G.; Paoletti, E. Antioxidative Responses of Three Oak Species under Ozone and Water Stress Conditions. *Sci. Total Environ.* **2019**, *647*, 390–399. [[CrossRef](#)] [[PubMed](#)]
26. Paoletti, E.; Carriero, G. A New-Generation 3D Ozone FACE (Free Air Controlled Exposure). *Sci. Total Environ.* **2017**, *575*, 1407–1414. [[CrossRef](#)] [[PubMed](#)]
27. Moura, B.B.; Hoshika, Y.; Ribeiro, R.V.; Paoletti, E. Exposure- and Flux-Based Assessment of Ozone Risk to Sugarcane Plants. *Atmos. Environ.* **2018**, *176*, 252–260. [[CrossRef](#)]
28. Innes, J.L.; Skelly, J.M.; Schaub, M. *Ozone and Broadleaved Species: A Guide to the Identification of Ozone-Induced Foliar Injury/ Ozon, Laubholz-Und Krautpflanzen: Ein Fuhrer Zum Bestimmen von Ozonsymptomen*; Haupt: Bern, Switzerland, 2001.
29. Vollenweider, P.; Gunthardtgoerg, M. Erratum to “Diagnosis of Abiotic and Biotic Stress Factors Using the Visible Symptoms in Foliage”. *Environ. Pollut.* **2006**, *140*, 562–571. [[CrossRef](#)]
30. Günthardt-Goerg, M.S.; Kuster, T.M.; Arend, M.; Vollenweider, P. Foliage Response of Young Central European Oaks to Air Warming, Drought and Soil Type. *Plant Biol.* **2013**, *15*, 185–197. [[CrossRef](#)]
31. Chappelka, A.; Renfro, J.; Somers, G.; Nash, B. Evaluation of Ozone Injury on Foliage of Black Cherry (*Prunus Serotina*) and Tall Milkweed (*Asclepias Exaltata*) in Great Smoky Mountains National Park. *Environ. Pollut.* **1997**, *95*, 13–18. [[CrossRef](#)]
32. Team R Development Core. A Language and Environment for Statistical Computing. 2018. Available online: <https://www.R-project.org/> (accessed on 21 May 2022).
33. Fernandes, F.F.; Moura, B.B. Foliage Visible Injury in the Tropical Tree Species, *Astronium Graveolens* Is Strictly Related to Phytotoxic Ozone Dose (POD_y). *Environ. Sci. Pollut. Res.* **2021**, *28*, 41726–41735. [[CrossRef](#)]
34. Gao, F.; Catalayud, V.; Paoletti, E.; Hoshika, Y.; Feng, Z. Water Stress Mitigates the Negative Effects of Ozone on Photosynthesis and Biomass in Poplar Plants. *Environ. Pollut.* **2017**, *230*, 268–279. [[CrossRef](#)]
35. Grulke, N.E. The Physiological Basis of Ozone Injury Assessment Attributes in Sierran Conifers. *Dev. Environ. Sci.* **2003**, *2*, 55–81. [[CrossRef](#)]
36. Yoshimura, K. Programmed Proteome Response for Drought Avoidance / Tolerance in the Root of a C₃ Xerophyte (Wild Watermelon) Under Water Deficits. *Plant Cell Physiol.* **2008**, *49*, 226–241. [[CrossRef](#)]
37. Le Gall, H.; Philippe, F.; Domon, J.M.; Gillet, F.; Pelloux, J.; Rayon, C. Cell Wall Metabolism in Response to Abiotic Stress. *Plants* **2015**, *4*, 112. [[CrossRef](#)]
38. Pinheiro, C.; Chaves, M.M. Photosynthesis and Drought: Can We Make Metabolic Connections from Available Data? *J. Exp. Bot.* **2011**, *62*, 869–882. [[CrossRef](#)]
39. Pell, E.J.; Sinn, J.P.; Johansen, C.V. Nitrogen Supply as a Limiting Factor Determining the Sensitivity of *Populus Tremuloides* Michx. to Ozone Stress. *New Phytol.* **1995**, *130*, 437–446. [[CrossRef](#)]
40. Matyssek, R.; Sandermann, H. Impact of Ozone on Trees: An Ecophysiological Perspective. In *Progress in Botany*; Esser, K., Lüttge, U., Beyschlag, W., Hellwig, F., Eds.; Springer: Berlin/Heidelberg, Germany, 2003; ISBN 978-3-642-55819-1.
41. Jaleel, C.A.; Manivannan, P.; Wahid, A.; Farooq, M.; Al-Juburi, H.J.; Somasundaram, R.; Panneerselvam, R. Drought Stress in Plants: A Review on Morphological Characteristics and Pigments Composition. *Int. J. Agric. Biol.* **2009**, *11*, 100–105.
42. Chaturvedi, R.K.; Tripathi, A.; Raghubanshi, A.S.; Singh, J.S. Functional Traits Indicate a Continuum of Tree Drought Strategies across a Soil Water Availability Gradient in a Tropical Dry Forest. *For. Ecol. Manag.* **2021**, *482*, 118740. [[CrossRef](#)]
43. Moura, B.B.; Alves, E.S. Climatic Factors Influence Leaf Structure and Thereby Affect the Ozone Sensitivity of *Ipomoea Nil* “Scarlet O’Hara”. *Environ. Pollut.* **2014**, *194*, 11–16. [[CrossRef](#)]
44. Serrano, L.; Peñuelas, J. Contribution of Physiological and Morphological Adjustments to Drought Resistance in Two Mediterranean Tree Species. *Biol. Plant.* **2005**, *49*, 551–559. [[CrossRef](#)]
45. Watanabe, M.; Agathokleous, E.; Anav, A.; Araminiene, V.; Carrari, E.; De Marco, A.; Hoshika, Y.; Proietti, C.; Sicard, P.; Paoletti, E. Impacts of Ozone on the Ecophysiology of Forest Tree Species. In *Tropospheric Ozone—A Hazard for Vegetation and Human Health*; Agrawal, S.B., Agrawal, M., Singh, A., Eds.; Cambridge Scholars: Newcastle, UK, 2021; pp. 277–306.
46. Shang, B.; Xu, Y.; Dai, L.; Yuan, X.; Feng, Z. Elevated Ozone Reduced Leaf Nitrogen Allocation to Photosynthesis in Poplar. *Sci. Total Environ.* **2019**, *657*, 169–178. [[CrossRef](#)]
47. Weih, M.; Bonosi, L.; Ghelardini, L.; Rönnberg-Wästljung, A.C. Optimizing Nitrogen Economy under Drought: Increased Leaf Nitrogen Is an Acclimation to Water Stress in Willow (*Salix* Spp.). *Ann. Bot.* **2011**, *108*, 1347–1353. [[CrossRef](#)]
48. Mrak, T.; Štraus, I.; Grebenc, T.; Gričar, J.; Hoshika, Y.; Carriero, G.; Paoletti, E.; Kraigher, H. Different Belowground Responses to Elevated Ozone and Soil Water Deficit in Three European Oak Species (*Quercus Ilex*, *Q. Pubescens* and *Q. Robur*). *Sci. Total Environ.* **2019**, *651*, 1310–1320. [[CrossRef](#)]
49. Somers, G.L.; Chappelka, A.H.; Rosseau, P.; Renfro, J.R. Empirical Evidence of Growth Decline Related to Visible Ozone Injury. *For. Ecol. Manag.* **1998**, *104*, 129–137. [[CrossRef](#)]
50. Marzuoli, R.; Gerosa, G.; Bussotti, F.; Pollastrini, M. Assessing the Impact of Ozone on Forest Trees in an Integrative Perspective: Are Foliar Visible Symptoms Suitable Predictors for Growth Reduction? A Critical Review. *Forests* **2019**, *10*, 1144. [[CrossRef](#)]
51. Chirino, E.; Vilagrosa, A.; Hernández, E.I.; Matos, A.; Vallejo, V.R. Effects of a Deep Container on Morpho-Functional Characteristics and Root Colonization in *Quercus Suber* L. Seedlings for Reforestation in Mediterranean Climate. *For. Ecol. Manag.* **2008**, *256*, 779–785. [[CrossRef](#)]

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52. Giovannelli, A.; Traversi, M.L.; Anichini, M.; Hoshika, Y.; Fares, S.; Paoletti, E. Effect of Long-Term vs. Short-Term Ambient Ozone Exposure on Radial Stem Growth, Sap Flux and Xylem Morphology of O₃-Sensitive Poplar Trees. *Forests* **2019**, *10*, 396. [[CrossRef](#)]
 53. Agathokleous, E.; Saitanis, C.J.; Wang, X.; Watanabe, M.; Koike, T. A Review Study on Past 40 Years of Research on Effects of Tropospheric O₃ on Belowground Structure, Functioning, and Processes of Trees: A Linkage with Potential Ecological Implications. *Water, Air, Soil Pollut.* **2016**, *227*, 33. [[CrossRef](#)]