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Abstract: Non-destructive tools to evaluate the lycopene content in tomatoes is of large interest for the whole fruit chain because of the increasing demand for health beneficial products. With the aim to develop compact low-cost reflectance sensors for lycopene determination, we compared prediction Partial Least Squares (PLS) models by using either directional or total reflectance in the 500-750 nm range. Directional reflectance at 45° with respect to the LED lighting direction was acquired by a compact spectrometer sensor. Total reflectance was acquired through a 50-mm integrating sphere connected to a spectrometer. The analysis was conducted on two hydroponic greenhouse cultivated red tomato varieties, large round 'Dometica' (average diameter of 57 mm) and small cherry 'Juanita' (average diameter 26 mm). For both varieties, the spectral variance of directional reflectance was well correlated to that of total reflectance. Performances of PLS prediction models were also similar, with R2 of cross-validation between 0.73 and 0.81. The prediction error, relative to the mean lycopene content of full ripe tomatoes, was similar and around 16-17% for both varieties and sensors. Our results showed that directional reflectance measured by portable, low-cost and compact LED-based sensors can be used with a fair precision for the non-destructive assessment of lycopene in tomatoes.



Sesto Fiorentino (Florence), 13 October 2015

Editor-in-Chief
J. Harnly
U.S. Department of Agriculture (USDA)
Beltsville, Maryland, USA

Dear Editor,

the manuscript entitled

Directional versus total reflectance spectroscopy for the in situ determination of lycopene in tomato fruits

by

Leonardo Ciaccheri, Lorenza Tuccio, Andrea A. Mencaglia, Anna G. Mignani, Ewelina Hallmann, Kalina Sikorska-Zimny, Stanislaw Kaniszewski, Michèl J. Verheul, Giovanni Agati

we are submitting for publication in JFCA is based on results of a multidisciplinary collaboration within the frame of an ERA-NET SUSFOOD European project.

The aim was that to test the performance of a new developed reflectance optical sensor for the non-destructive estimate of lycopene in tomatoes. We wanted to compare directional versus total reflectance by using destructive analysis of lycopene as reference and PLS regression analysis. We think that this can be a useful step to design new compact sensors to control rapidly the lycopene level of tomatoes in the field and during storage.

As reviewers for the present manuscript, I would suggest:

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Best regards

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Directional versus total reflectance spectroscopy for the in situ determination of lycopene in tomato fruits

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30 ABSTRACT

31 Non-destructive tools to evaluate the lycopene content in tomatoes is of large interest for the whole
32 fruit chain because of the increasing demand for health beneficial products. With the aim to develop
33 compact low-cost reflectance sensors for lycopene determination, we compared prediction Partial
34 Least Squares (PLS) models by using either directional or total reflectance in the 500-750 nm range.
35 Directional reflectance at 45° with respect to the LED lighting direction was acquired by a compact
36 spectrometer sensor. Total reflectance was acquired through a 50-mm integrating sphere connected
37 to a spectrometer. The analysis was conducted on two hydroponic greenhouse cultivated red tomato
38 varieties, large round ‘Dometica’ (average diameter of 57 mm) and small cherry ‘Juanita’ (average
39 diameter 26 mm). For both varieties, the spectral variance of directional reflectance was well
40 correlated to that of total reflectance. Performances of PLS prediction models were also similar,
41 with R^2 of cross-validation between 0.73 and 0.81. The prediction error, relative to the mean
42 lycopene content of full ripe tomatoes, was similar and around 16-17% for both varieties and
43 sensors.
44 Our results showed that directional reflectance measured by portable, low-cost and compact LED-
45 based sensors can be used with a fair precision for the non-destructive assessment of lycopene in
46 tomatoes.

47

48

49 Keywords

50 Lycopene; *Lycopersicon esculentum*; Non-destructive assessment; Optical Sensor; Partial Least
51 Squares regression; Reflectance; Tomato.

52

53 Abbreviations

54 PLS, Partial Least Squares; RMSEC, Root Mean Square Error of Calibration; RMSECV, Root
55 Mean Square Error of Cross-Validation.

57 **1. Introduction**

58 Lycopene, possessing high oxygen-radical scavenging properties, is the main antioxidant compound
59 present in ripe tomato (*Lycopersicon esculentum* Mill.) fruits (Shi and Maguer, 2000). Because of
60 this, its assumption can significantly reduce the risk of cancer and cardiovascular diseases (Müller
61 et al., 2016; Rao and Agarwal, 2000). Therefore, it should be of large interest in producing tomatoes
62 with the largest content of lycopene and to quantify this compound as quality added-value.

63 Usually, the concentration of lycopene is determined destructively on sample extracts by means of
64 unsafe solvents and time-consuming methods (Davis et al., 2003a; Fish et al., 2002). To improve
65 this procedure, several non-destructive spectroscopic methods have been proposed (Baranska et al.,
66 2006; Choudhary et al., 2009; Clément et al., 2008). Other more sophisticated techniques concerned
67 tomato spectral imaging that, besides the prediction of lycopene, can give information of the spatial
68 heterogeneity of it (Liu et al., 2015; Polder et al., 2004). However, a standardized system and
69 procedure for the fast estimate of lycopene to be transferred to tomato producers is not available
70 yet.

71 Reflectance spectroscopy has been used for a long time to assess in situ fruit quality (Cozzolino,
72 2014; Nicolai et al., 2007). The method is rapid, non-destructive and can be applied both in the
73 laboratory and in the field. The information that can be obtained from a reflectance spectrum
74 depends on the absorbance and scattering properties of the sample and on the spectral range
75 investigated. Highly pigmented fruits absorb visible light within the first few micron of tissue, then
76 reflectance between 400 and 700 nm can provide data only on the superficial part of the sample.
77 NIR, instead, penetrates deeper into the fruit tissues and can provide information on internal
78 compounds and pulp structure.

79 Specificity of reflectance spectroscopy can be limited when, as usual, a high number of overlapping
80 bands occur. For this, chemometrics can be used to extract information from the spectral data
81 concerning sample quality features (Wang et al., 2015).

The geometry of the irradiation and detection system is also important. In a directional-hemispherical system, irradiation occurs within a solid angle oriented with respect to the normal to the sample surface and the reflected radiation is collected over the whole hemispherical space above the sample surface.

Bidirectional reflectance of samples can be fully characterized by using a spectrogoniophotometer that allows measurements as function of the irradiation and detection angles (Combes et al., 2007). Since fruit and vegetable surfaces are far to be Lambertian, the angle of detection with respect to the irradiation direction can significantly affect the reflectance values. Including or excluding the specular reflection is also fundamental especially for glossy samples (Combes et al., 2007).

In principal, collecting all the radiation reflected by the sample by using an integrating sphere, with internal diameter ≥ 150 mm and port fraction below 4%, should be the ideal in order to get the highest accuracy and sensitivity. However, with the aim to develop low-cost, user-friendly sensor as much compact as possible the choice can go towards a simpler configuration (Mignani et al., 2015).

In this study, we compared directional versus diffuse reflectance spectroscopy for the prediction of lycopene in tomatoes. The two non-destructive techniques were correlated to the real content in lycopene extracted from the very same samples measured spectroscopically and quantified by HPLC. We used two tomato varieties with quite different size, in order to investigate if this could have an effect on the method performance.

2. Materials and Methods

2.1. Fruit material

Fully developed round tomatoes (*Lycopersicon esculentum* Mill. ‘Dometica’) and cherry tomatoes (*Lycopersicon esculentum* Mill. ‘Juanita’) were harvested at different ripening stages at the beginning of August 2014 from two commercial greenhouses, Orre Gartneri – Orre and Lauvsnes Gartneri – Finnøy, respectively, in southwestern Norway.

107 All the different maturity stages: “green”, “breakers”, “turning”, “pink”, “light red” and “red”
108 (USDA, 1975) were represented (Fig. 1). Twenty-seven fruits for each variety were used in the
109 study.
110 Plants were grown as hydroponic greenhouse cultures under optimal standard growth conditions in
111 Norway (Verheul, 2012). Tomatoes were harvested from greenhouse compartments with and
112 without the use of artificial light ($280 \mu\text{mol m}^{-2}\text{s}^{-1}$ PAR for maximum 18h per day). Fruits from
113 trusses 10-12 (Dometica) or 2-3 (Juanita) were harvested. Global radiation outside of the
114 greenhouse the last 14 days before harvesting was on average $19.5 \text{ MJ m}^{-2} \text{ d}^{-1}$. Daily mean
115 temperatures in the greenhouse were 21°C (Dometica) or 21.5°C (Juanita). Plants were irrigated
116 when needed with a standard nutrient solution for tomato, keeping a EC of 4.0 dS m^{-1} (Dometica) or
117 4.5 dS m^{-1} (Juanita) in the rockwool slabs.

118

119 2.2. *Reflectance sensors*

120 Directional reflectance spectra of tomatoes were measured by the LED-based sensor (hereafter
121 LED-sensor) described in Mignani et al. (Mignani et al., 2015).
122 Briefly, it consisted of a group of LEDs for illumination and a compact spectrometer for detection.
123 The light source was made of three sets of four LEDs sequentially placed in a twelve-element
124 circular-array. Each set was made of a white LED and other three LEDs emitting at 740 nm, 420
125 nm, and 680 nm, respectively. Lighting of the samples occurred at 45° with respect to the axis of
126 detection. This opto-geometric arrangement minimizes the influence of any surface texture or
127 roughness effects.

128 The detector was an Ocean Optics STS-VIS spectrometer (Ocean Optics, Dunedin, FL, USA) with
129 optical fibre input, operating in the 350-800 nm spectral range, with a resolution of 1.5 nm. The
130 sensor was USB-connected to a notebook and operated through a custom Labview® software
131 interface. The sampled zone of tomatoes at the exit port of the sensor was 5 mm in diameter.

132 Total reflectance spectra of tomatoes were acquired by the compact spectrometer BRC112E-USB-
133 VIS (B&W Tek, Lübeck, Germany) working in the visible range (380-750 nm), with a 25 µm slit
134 and a resolution at 546 nm of 1 nm, connected through an optical fibre to a 50-mm integrating
135 sphere (AvaSphere-50-LS-HAL, Avantes, The Netherlands), hereafter IS-sensor. An external
136 halogen lamp (LS-1 Tungsten Halogen Light Source, Ocean Optics, Dunedin, FL, USA) connected
137 to the integrating sphere by an optical fibre was used to illuminate directly the tomato samples at an
138 8° angle of incidence. The integrating sphere sensor, hereafter IS-sensor, sampled a zone of
139 tomatoes of 10 mm in diameter.

140 The same day of harvesting, the reflectance spectra of each tomato were measured by the two
141 sensors on four different points, at 90° to each other, along the equatorial zone. The average
142 spectrum of the four measurements was then calculated.

143 A white Spectralon® standard was used as reference.

144 After reflectance measurements, each ‘Dometica’ tomato was finely homogenized in a blender and
145 then immediately frozen in liquid nitrogen. Aliquots 8-10 g were freeze-dried to get about 1 g dried
146 powder. ‘Juanita’ tomatoes were cut in 4 pieces and immediately frozen in liquid nitrogen and then
147 freeze-dried.

148

149 2.3. *Lycopene extraction and analysis*

150 Lycopene extraction and analysis of ‘Juanita’ tomatoes were performed according to Hallmann
151 (2012) with some adjustment. Fifty mg of powdered freeze-dried samples were weighted to plastic
152 tubes, to which magnesium carbonate (MgCO₂) and cold n-hexane (5 ml) were added. Samples
153 were shaken on vortex and extracted in ultrasonic cold bath (15 min in the dark) and then
154 centrifuged at 6,000 rpm, 0°C, for 10 min. One ml of supernatant was transferred to Eppendorf
155 tubes and centrifuged again (12,000 rpm, 0°C, 5 min). Nine hundred µl of supernatant were injected
156 into the HPLC column (Max-RP 80i, 250 x 4.60 mm). The HPLC set-up consisted of two LC-20AD
157 pumps, a CMB-20A system controller, SIL-20AC autosampler, ultraviolet–visible SPD-20AV

158 detector, and CTD-20AC oven. Acetonitrile and deionized water with ethyl acetate (90:10:100) at a
159 flow rate of 1 mL min⁻¹ was used as a gradient solvent. The time of analysis was 27 min., with
160 detection at 471 nm. Carotenoids were identified on the basis of external standards (Sigma-Aldrich)
161 with purity of 99.9%.

162 Lycopene extraction and analysis of 'Dometica' tomatoes were performed according to the AOAC
163 method (Latimer, 2012). Five hundred mg of freeze-dried tomato samples were re-suspended with
164 12 ml of water containing butylated hydroxytoluene (BHT, 0.01g). Samples were shaken in
165 ultrasonic bath (30 min), then added with ethanol (16 ml) and dichloromethane (DCM) to a final
166 volume of 40 ml and stored in the dark for 2 h. Twenty µl of supernatant were inject into the HPLC
167 column (Suplex pKb-100 250x4.6 mm, 5 µm) kept at 30°C. Methanol with the addition of BHT
168 (0.005% w:v), 2-propanol (2% v:v), N-ethyl-diisopropylamine (0.02% v:v), ammonium acetate
169 (2.5% v:v) and acetonitrile (45.5% v:v) at a flow rate of 0.8 mL min⁻¹ was used as an isocratic
170 solvent. The time of analysis was 20 min with detection at 488 nm for lycopene, identified by an
171 external standard (Sigma-Aldrich).

172

173 2.4. Statistical analysis

174 Before statistical analysis, reflectance spectra were smoothed by mobile average. The smoothing
175 window was 15 and 11 points for IS- and LED-sensor spectra, respectively, with a width of about 5
176 nm, in both cases. Spectra were then converted to apparent absorbance units by taking the natural
177 logarithm of the reciprocal of reflectance and corrected for the offset.

178 To better compare reflectance data from the two systems, spectra were re-sampled at 2 nm step and
179 analysed by using the same wavelength range (500-750 nm). Wavelengths below 500 nm were
180 discarded because they showed a saturated absorption. The new sampling step was more than
181 adequate in comparison with the average width of spectral features, and provided a further damping
182 of high frequency noise.

Partial Least Squares (PLS) regression analysis was used for lycopene content prediction. PLS is a widely used regression method when many co-linear variables are present. It provides a strong reduction in the number of predictors, thus reducing the probability of overfitting, while maximizing the covariance between the predictor set and the target variable (Esbensen et al., 2002). The accuracy of the calibration and validation models was described by the coefficient of determination (R^2), the Root Mean Square Error of Calibration (RMSEC) and the Root Mean Square Error of Cross-Validation (RMSECV).

The RMSEC was calculated as follows:

$$RMSEC = \sqrt{\frac{\sum_{n=1}^N (y_n - z_n)^2}{N - K}} \quad (1)$$

where y_n are the reference values, z_n are the predicted values, N is the number of samples and K is the number of statistical parameters estimated in calibrating the model.

The expression of RMSECV was similar considering at the denominator simply the number of samples, N .

The R^2 was a measure of the size of RMSEC relative to the standard deviation of y variable given by:

$$R^2 = 1 - \left(\frac{RMSEC}{s_y} \right)^2 \quad (2)$$

where s_y is the standard deviation of y . A similar coefficient can be estimated for RMSECV.

The data processing was carried out by means of CAMO-Unscrambler software. Because of the limited number of samples available, it was not possible to divide the entire set into two sets for calibration and validation, respectively, and the validation was carried out using the Leave-One-Out (LOO) method (Esbensen et al., 2002).

208 3. Results and Discussion

209

210 The two varieties of tomato fruits differed significantly in weight and size. The average weight was
211 93 ± 11 g for ‘Dometica’ and 10 ± 2 g for ‘Juanita’. The average diameter of ‘Dometica’ fruits was
212 almost double that of ‘Juanita’ (56.7 ± 2.7 versus 25.9 ± 1.5 mm).

213 The concentration of lycopene in tomato fruits resulting from the destructive chemical extraction
214 and HPLC analysis ranged from 0 to 79 mg/100 g DW and from 1 to 49 mg/100 g DW for the
215 ‘Dometica’ and ‘Juanita’ varieties, respectively. These data were used as references for the
216 calibration and cross validation of the PLS models.

217 At the full ripe “red” stage the lycopene concentration was 68.1 ± 7.7 and 41.1 ± 5.1 mg/100g DW
218 for ‘Dometica’ and ‘Juanita’, respectively. Considering a dry matter for ‘Dometica’ of 6%, ripe
219 tomatoes had a concentration of about 4 mg/100 g fresh weight that is in line with that found early
220 for this variety cultivated in Norwegian greenhouses (Verheul et al., 2015). Lycopene content for
221 ‘Juanita’ fruits was almost half than that found by Verheul in the same variety in 2013 (data not
222 shown). Nassar et al. (Nassar et al., 2015) found also higher lycopene concentrations in ‘Juanita’
223 hydroponically grown (76.7 mg/100 g DW). This difference can be partially explained by a
224 different ripening stage at harvest, by the fact that fruits were harvested from young plants and,
225 likely, by a difference in the methods of analysis.

226 Figure 2 shows representative directional and total reflectance spectra measured on ‘Dometica’ and
227 ‘Juanita’ tomatoes at different ripening stages by both the LED- and IS- sensors. As expected,
228 reflectance values recorded by the IS-sensor were higher than those recorded by the LED-sensor.
229 This was because: i) specular reflectance was measured by the IS-sensor but not the LED-sensor; ii)
230 tomato fruits are non Lambertian surfaces so that directional reflectance is depending on the view
231 angle of detector and cannot equal hemispherical reflectance. The IS-sensor detected a larger
232 portion of light scattered inside the tomato samples than the LED-sensor.

233 For both varieties, the absorbance band of chlorophyll was clearly evident as a reflectance
234 minimum at around 675 nm for samples at breakers-turning stages. With ripening, reflectance in the
235 blue-yellow decreased due to the accumulation of lycopene, while reflectance in the red increased
236 because of chlorophyll degradation.

237 The absorbance bands of chlorophyll and lycopene for the same fruit appeared slightly more evident
238 in the IS-sensor spectra than in the LED-sensor. This is likely due to a possible multi-pass effect
239 present in the integrating sphere.

240 Partial Least Square regression analysis of reflectance spectra showed that for both tomato varieties
241 and reflectance sensors the first PLS factor was able to explain more than 95% of X-variance (i.e.
242 the summed variance of all column of the spectra matrix). Therefore, the different shapes observed
243 in the directional and total reflectance spectra did not change significantly the analysis of variance.

244 The good correlation found between the scores of first PLS factors, reported in Fig. 3A and B for
245 ‘Dometica’ and ‘Juanita’ tomatoes respectively, demonstrated that such factors were related to a
246 common cause. It was also evident that such cause was the increase of lycopene (and simultaneous
247 decrease of chlorophyll) during ripening.

248 The slope of the regression line between the first factor scores of LED-sensor and IS-sensor data in
249 ‘Dometica’ was very close to 1 (0.95), while for ‘Juanita’ the same slope was about 0.6. This is
250 explainable by a similar X-variance for both sensors in the former case, while in the latter, the X-
251 variance of IS-sensor was significantly lower than LED-sensor. The effect was likely due to the
252 different sizes of interacting area and angular detection range possessed by the two detectors in
253 relation with the small size of ‘Juanita’ tomatoes. In fact, such effect was not observed for the larger
254 ‘Dometica’ fruits.

255 The PLS calibration models explained 77% or more of Y-variance by using just a single factor for
256 LED-sensor data and IS-sensor ‘Dometica’ data. Two factors were instead necessary for IS-sensor
257 ‘Juanita’ data (Table 1). This could be due to the skewed distribution of the spectral data, which
258 tend to reduce the representativeness of estimated statistical parameters.

259 The PLS regression coefficients spectra for lycopene prediction are reported in Figure 4. Regression
260 coefficients for ‘Dometica’ derived from both LED- and IS- sensors data were quite similar (Fig.
261 4A), showing a negative peak at around 675 nm corresponding to the chlorophyll absorbance
262 maximum in the red, and a positive peak at around 560 nm often used in the past for
263 spectrophotometric determination of lycopene content in turbid media (Davis et al., 2003b). For
264 ‘Juanita’ fruits (Fig. 4B), the regression coefficients pattern for data from LED-sensor was
265 comparable to the ‘Dometica’ ones, while the 2-factors coefficients from IS-sensor data showed a
266 sharp positive peak at around 600 nm, beside the negative chlorophyll peak at 675 nm.

267 The performance of PLS regression models to predict lycopene from directional and total
268 reflectance spectroscopy can be evaluated from the scatter plots reported in Figures 5 and 6 for the
269 ‘Dometica’ and ‘Juanita’ varieties, respectively, and from the statistics data reported in Table 1.

270 For ‘Dometica’, lycopene prediction was slightly better by using directional than total reflectance,
271 while the opposite occurred for ‘Juanita’. Considering the prediction relative error, calculated as the
272 ratio between RMSECV and the mean lycopene content found in full ripe “red” tomatoes, it was
273 similar and around 16-17% for both varieties and sensors. These data suggested that both spectral
274 sensors utilized to estimate tomato lycopene performed similarly and that the prediction models
275 were not significantly affected by the size of the fruits.

276 The accuracy of the prediction model can be limited because of the heterogeneity of lycopene
277 distribution on fruits, especially for the early stages of ripening, when the reflectance spectrum
278 taken as average on four equatorial points might not be representative of the whole fruit. A
279 saturation of reflectance at the very full ripe stage may also occur. This could be related to the skin
280 to pulp distribution of lycopene. In fact, the ratio of lycopene content between skin and pulp
281 depends on the cultivar and changes during ripening (Kaur et al., 2006; Toor and Savage, 2005).

282 Since visible light does not penetrate much in highly pigmented samples, it is possible that the
283 correlation between optical signals and lycopene concentration can markedly change according to
284 the tomato fraction considered. This aspect will be investigated in future work.

Several different non-destructive spectroscopic techniques along with chemometrics have been used in the past to predict lycopene content in tomato fruits. Liu et al. (Liu et al., 2015) used multispectral imaging and multivariate analyses on cv. Wanza 15 tomatoes, finding higher or lower precision in assessing lycopene according to different quantitative models. PLS regression gave RMSEP of 6.5 mg/kg over the highest values of lycopene of about 36 mg/kg (that is about 18%). The error reduced to 2.3-2.6 mg/kg (6.4-7.2%) when new chemometric analysis such as least square-support vector machine (LS-SVM) and back propagation neural network (BPNN) were used. Clement et al. (Clément et al., 2008) presented an accurate prediction of lycopene ($R^2 = 0.98$, relative error at maturity of about 5%) on different (var. Trust, Blitz and DRK 453) not completely identified tomato varieties by using a bench spectrophotometer equipped with an integrating sphere and PLS analysis with 12 latent variables. A relative error of about 21% of prediction on the most red tomatoes cv. Momotaro was found by using a portable NIR sensor in the 600-1100 nm spectral range (Kusumiyati et al., 2008). Similar results were obtained by measuring the fruits directly on the plant or at postharvest. Imaging spectrometry between 396 and 736 nm along with PLS regression analysis was used on greenhouse cultivated tomatoes cv. Capita obtaining 8-9% relative error of prediction at the full ripe stage (Polder et al., 2004).

The level of performance of the different techniques depend on several aspects, such as the instrumentation used to obtain spectral data, more or less sophisticated algorithms of data elaboration, the spectral range considered, tomato cultivars and growing conditions, the destructive analysis for lycopene determination. For this, comparison of results from different methods is difficult and a conclusive indication of the best procedure to be used is still lacking. Yet, for practical purposes, it is often useful to find an optimal compromise between the precision of the techniques and the cost and user-friendliness of it.

311 **4. Conclusion**

312 Our results showed that portable, low-cost and compact LED-based sensors measuring directional
313 reflectance can be used with a fair precision for the non-destructive assessment of lycopene in
314 tomatoes. Although this approach can be limited in accuracy with respect to more sophisticated
315 spectroscopic techniques and statistical analyses, it can be useful for practical applications and to
316 facilitate transfer of the methodology to tomato producers. Calibrated portable sensors can be
317 employed directly into the field (or greenhouses) to predict the best time of harvesting in order to
318 get the highest level of lycopene. This is an important issue considering the marked dependence of
319 lycopene content on the environmental conditions (Brandt et al., 2006).
320 Furthermore, there will be interest in applying the reflectance technique to control the level of
321 lycopene in tomatoes during their travelling from producers to consumers and during storage
322 conditions (Schouten et al., 2014; Verheul et al., 2015), or to evaluate its seasonal variation (Toor et
323 al., 2006). The spread of non-destructive tools, as those presented here, will have also a positive
324 environmental impact reducing the use of time-consuming and costly destructive analyses. Lastly,
325 another remarkable application of reflectance sensors could concern the monitoring of lycopene
326 stability during tomato processing (Seybold et al., 2004; Shi and Maguer, 2000).

327

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338
339

340 **References**

341

- 342 Baranska, M., Schütze, W., Schulz, H., 2006. Determination of lycopene and β -carotene content in
343 tomato fruits and related products: comparison of FT-Raman, ATR-IR, and NIR spectroscopy.
344 Anal. Chem. 78 (24), 8456-8461.
- 345 Brandt, S., Pék, Z., Barna, É., Lugasi, A., Helyes, L., 2006. Lycopene content and colour of
346 ripening tomatoes as affected by environmental conditions. J. Sci. Food Agric. 86 (4), 568-572.
- 347 Choudhary, R., Bowser, T.J., Weckler, P., Maness, N.O., McGlynn, W., 2009. Rapid estimation of
348 lycopene concentration in watermelon and tomato puree by fiber optic visible reflectance
349 spectroscopy. Postharvest Biol. Technol. 52 (1), 103-109.
- 350 Clément, A., Dorais, M., Vernon, M., 2008. Nondestructive measurement of fresh tomato lycopene
351 content and other physicochemical characteristics using visible-NIR spectroscopy. J. Agric. Food
352 Chem. 56 (21), 9813-9818.
- 353 Combes, D., Bousquet, L., Jacquemoud, S., Sinoquet, H., Varlet-Grancher, C., Moya, I., 2007. A
354 new spectrogoniophotometer to measure leaf spectral and directional optical properties. Remote
355 Sens. Environ. 109 (1), 107-117.
- 356 Cozzolino, D., 2014. Use of infrared spectroscopy for in-field measurement and phenotyping of
357 plant properties: instrumentation, data analysis, and examples. Applied Spectroscopy Reviews 49
358 (7), 564-584.
- 359 Davis, A.R., Fish, W.W., Perkins-Veazie, P., 2003a. A rapid hexane-free method for analyzing
360 lycopene content in watermelon. J. Food Sci. 68 (1), 328-332.

361 Davis, A.R., Fish, W.W., Perkins-Veazie, P., 2003b. A rapid spectrophotometric method for
362 analyzing lycopene content in tomato and tomato products. *Postharvest Biol. Technol.* 28 (3), 425-
363 430.

364 Esbensen, K.H., Guyot, D., Westad, F., Houmoller, L.P. (2002). *Multivariate data analysis: in*
365 *practice: an introduction to multivariate data analysis and experimental design*, Oslo, Norway:
366 CAMO Software AS.

367 Fish, W.W., Perkins-Veazie, P., Collins, J.K., 2002. A quantitative assay for lycopene that utilizes
368 reduced volumes of organic solvents. *J. Food Compos. Anal.* 15 (3), 309-317.

369 Hallmann, E., 2012. The influence of organic and conventional cultivation systems on the
370 nutritional value and content of bioactive compounds in selected tomato types. *J. Sci. Food Agr.* 92
371 (14), 2840-2848.

372 Kaur, D., Sharma, R., Abas Wani, A., Singh Gill, B., Sogi, D., 2006. Physicochemical changes in
373 seven tomato (*Lycopersicon esculentum*) cultivars during ripening. *Int. J. Food Prop.* 9 (4), 747-
374 757.

375 Kusumiyati, Akinaga, T., Tanaka, M., Kawasaki, S., 2008. On-tree and after-harvesting evaluation
376 of firmness, color and lycopene content of tomato fruit using portable NIR spectroscopy. *Journal of*
377 *Food, Agriculture & Environnement* 6 (2), 327-332.

378 Latimer, G.W., Jr. (2012). *Official Methods of Analysis of AOAC INTERNATIONAL*, 19th Edition,
379 Gaithersburg, MD, USA: AOAC INTERNATIONAL.

380 Liu, C., Liu, W., Chen, W., Yang, J., Zheng, L., 2015. Feasibility in multispectral imaging for
381 predicting the content of bioactive compounds in intact tomato fruit. *Food Chem.* 173, 482-488.

382 Mignani, A.G., Ciaccheri, L., Mencaglia, A.A., Tuccio, L., Agati, G., (2015). Application of a
383 LED-based reflectance sensor for assessing in situ the lycopene content of tomatoes (*Lycopersicon*
384 *esculentum* Mill.), in: Kim, M.S., Chao, K., Chin, B.A. (Eds.), *Sensing for Agriculture and Food*
385 *Quality and Safety VII*. Spie-Int Soc Optical Engineering, Bellingham, pp. 9488061-9488066.

386 Müller, L., Caris-veyrat, C., Lowe, G., Böhm, V., 2016. Lycopene and its antioxidant role in the
387 prevention of cardiovascular diseases - A critical review. *Crit. Rev. Food Sci.* 56, 1868–1879.

388 Nassar, A.M., Kubow, S., Donnelly, D.J., 2015. High-throughput screening of sensory and
389 nutritional characteristics for cultivar selection in commercial hydroponic greenhouse crop
390 production. *Int. J. Agron.* Article ID 376417, 28 pages.

391 Nicolai, B.M., Beullens, K., Bobelyn, E., Peirs, A., Saeys, W., Theron, K.I., Lammertyn, J., 2007.
392 Nondestructive measurement of fruit and vegetable quality by means of NIR spectroscopy: A
393 review. *Postharvest Biol. Technol.* 46 (2), 99-118.

394 Polder, G., Van der Heijden, G., Van der Voet, H., Young, I., 2004. Measuring surface distribution
395 of carotenes and chlorophyll in ripening tomatoes using imaging spectrometry. *Postharvest Biol.*
396 *Technol.* 34 (2), 117-129.

397 Rao, A.V., Agarwal, S., 2000. Role of antioxidant lycopene in cancer and heart disease. *J. Am.*
398 *Coll. Nutr.* 19 (5), 563-569.

399 Schouten, R.E., Farneti, B., Tijskens, L., Alarcón, A.A., Woltering, E.J., 2014. Quantifying
400 lycopene synthesis and chlorophyll breakdown in tomato fruit using remittance VIS spectroscopy.
401 *Postharvest Biol. Technol.* 96, 53-63.

402 Seybold, C., Fröhlich, K., Bitsch, R., Otto, K., Böhm, V., 2004. Changes in contents of carotenoids
403 and vitamin E during tomato processing. *J. Agric. Food Chem.* 52 (23), 7005-7010.

404 Shi, J., Maguer, M.L., 2000. Lycopene in tomatoes: chemical and physical properties affected by
405 food processing. *Crit. Rev. Food Sci.* 40 (1), 1-42.

406 Toor, R., Savage, G., Lister, C., 2006. Seasonal variations in the antioxidant composition of
407 greenhouse grown tomatoes. 19 (1), 1-10.

408 Toor, R.K., Savage, G.P., 2005. Antioxidant activity in different fractions of tomatoes. *Food Res.*
409 *Int.* 38 (5), 487-494.

410 USDA, 1975. Tomato Ripeness Color Chart. U.S. Department of Agriculture: Washington, DC.

411 Verheul, M.J. (2012). Effects of plant density, leaf removal and light intensity on tomato quality
412 and yield. In *Proceedings of the VII International Symposium on Light in Horticultural Systems* 956
413 (pp. 365-372). International Society for Horticultural Science (ISHS): Leuven, Belgium.

414 Verheul, M.I.J., Slimestad, R., Tjøstheim, I.H., 2015. From producer to consumer: greenhouse
415 tomato quality as affected by variety, maturity stage at harvest, transport conditions, and
416 supermarket storage. *J. Agric. Food Chem.* 63 (20), 5026-5034.

417 Wang, H., Peng, J., Xie, C., Bao, Y., He, Y., 2015. Fruit quality evaluation using spectroscopy
418 technology: a review. *Sensors* 15 (5), 11889-11927.

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423 **Figure Captions**

424

425 Figure 1. Tomato fruits of the ‘Dometica’ and ‘Juanita’ variety used in the study.

426

427 Figure 2. Representative spectra of directional (A,C) and total reflectance (B,D) acquired on
428 ‘Dometica’ (A,B) and ‘Juanita’ (C,D) fruits with different lycopene content by the LED-sensor and
429 the IS-sensor.

430

431 Figure 3. Correlation between scores of the PLS regression first factors from reflectance (R) data
432 recorded by the LED-sensor and the IS-sensor for ‘Dometica’ (A) and ‘Juanita’ (B) tomatoes.
433 Different symbols and colors indicate different ranges of the lycopene content (mg/100g DW): for
434 ‘Dometica’, 0-15 (circles, green); 15-30 (squares, yellow); 30-50 (triangles, orange); 50-79
435 (diamonds, red). For ‘Juanita’, 0-10 (circles, green); 10-15 (squares, yellow); 15-32 (triangles,
436 orange); 32-49 (diamonds, red).

437

438 Figure 4. Regression coefficients spectra for prediction of lycopene content in ‘Dometica’ (A) and
439 ‘Juanita’ (B) tomatoes. ‘Dometica’ data and LED-sensor ‘Juanita’ data refer to PLS models with 1
440 factor, IS-sensor data refer to a PLS model with 2 factors.

441

442 Figure 5. PLS prediction of lycopene content from reflectance spectroscopy in the 500-750 nm
443 range on ‘Dometica’ large round tomatoes by using directional reflectance (A, LED-sensor) or total
444 reflectance (B, IS-sensor). Dotted lines represent the identity lines.

445

446 Figure 6. PLS prediction of lycopene content from reflectance spectroscopy in the 500-750 nm
447 range on ‘Juanita’ small cherry tomatoes by using directional reflectance (A, LED-sensor) or total
448 reflectance (B, IS-sensor). Dotted lines represent the identity lines.

Table 1. Regression statistics for the prediction of lycopene in tomatoes						
Variety/Device	PLS Factors	RMSEC (mg/100g DW)	R ² cal.	RMSECV (mg/100g DW)	R ² cross-val.	Relative error (%)*
‘Dometica’						
LED-sensor	1	10.0	0.79	11.0	0.77	16.2
IS-sensor	1	11.0	0.77	12.0	0.73	17.6
‘Juanita’						
LED-sensor	1	6.2	0.80	6.6	0.77	16.0
IS-sensor	2	5.7	0.83	6.5	0.81	15.8

*Relative error was calculated as ratio between RMSECV and the mean lycopene concentration of full ripe “red” tomatoes.

Figure 1
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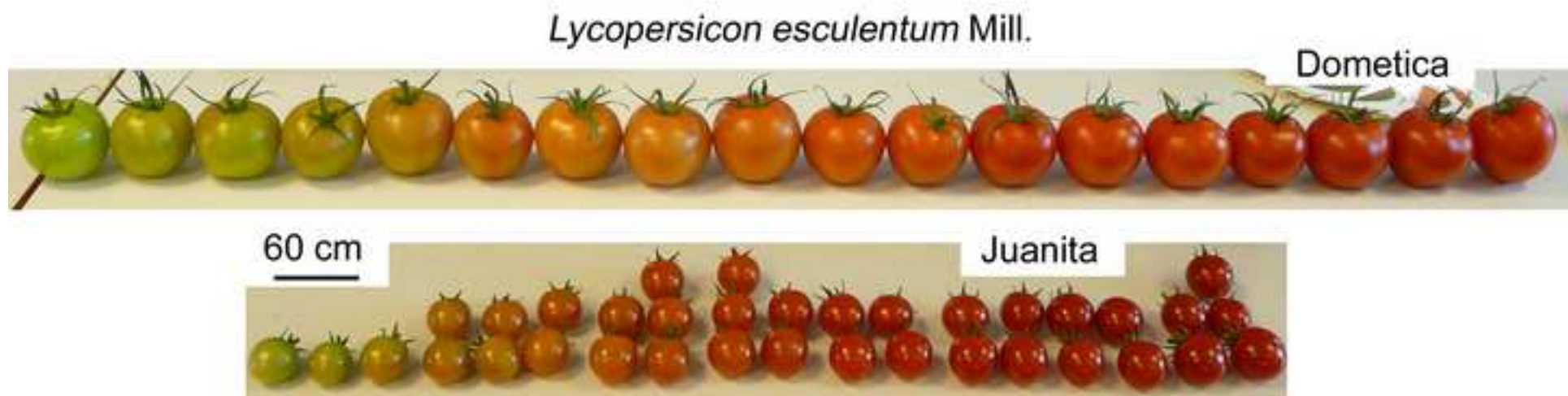


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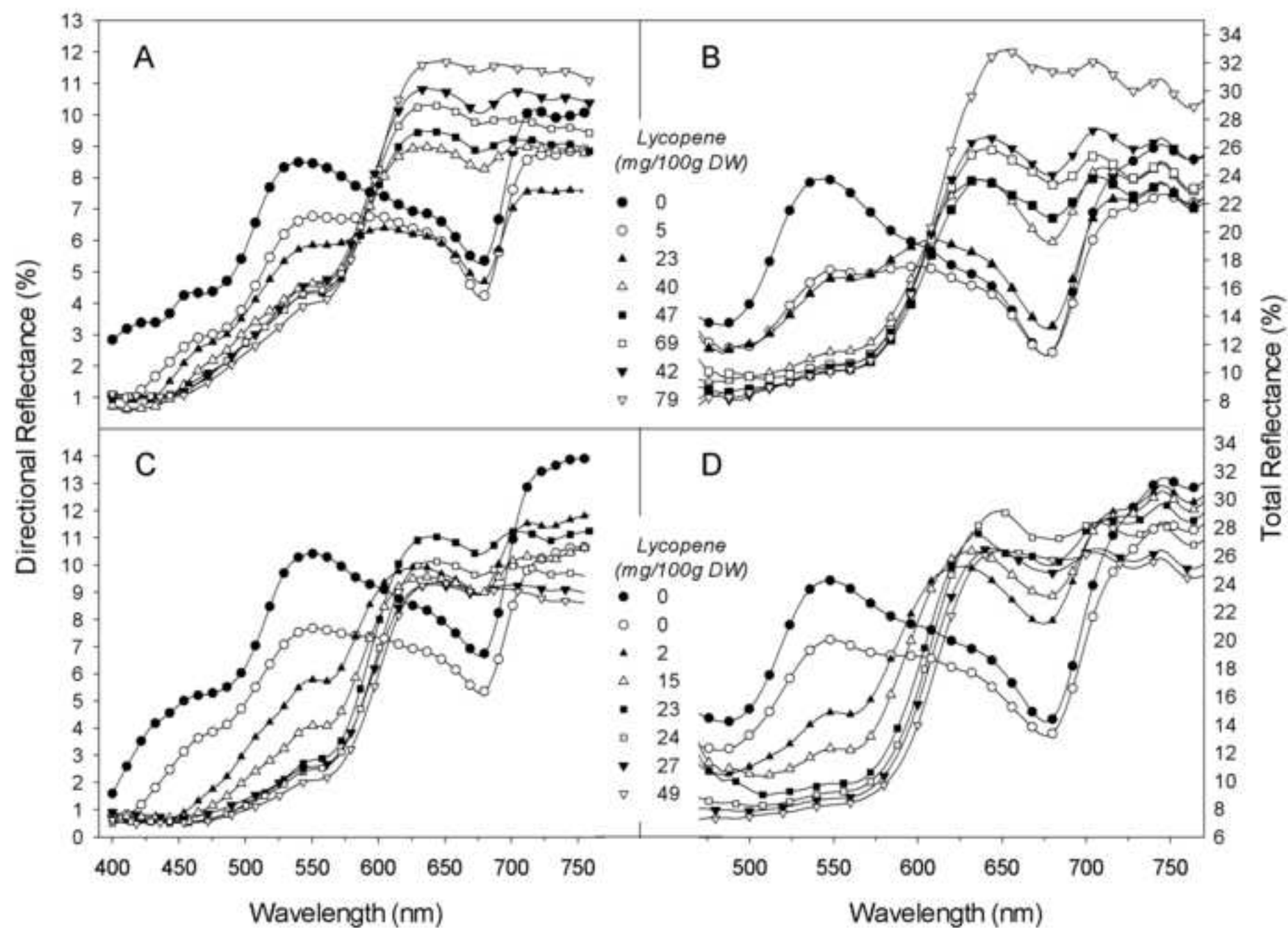


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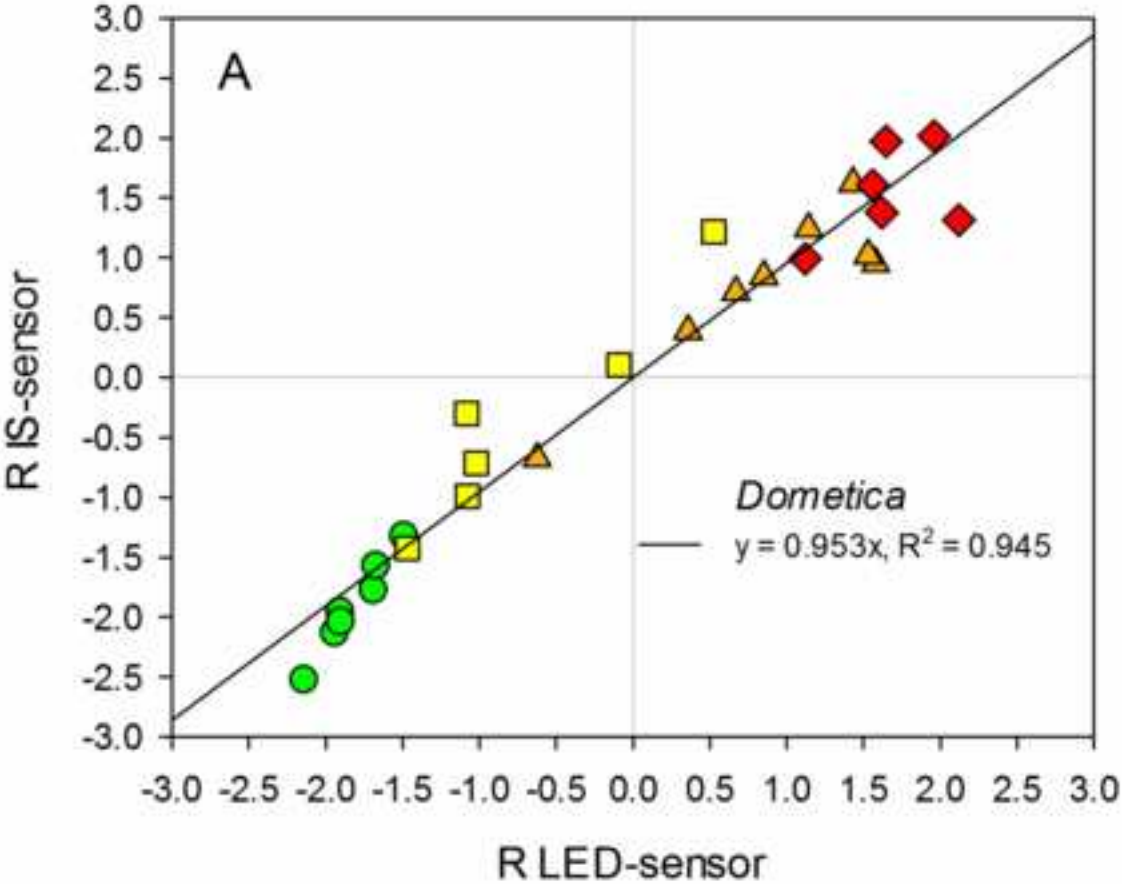


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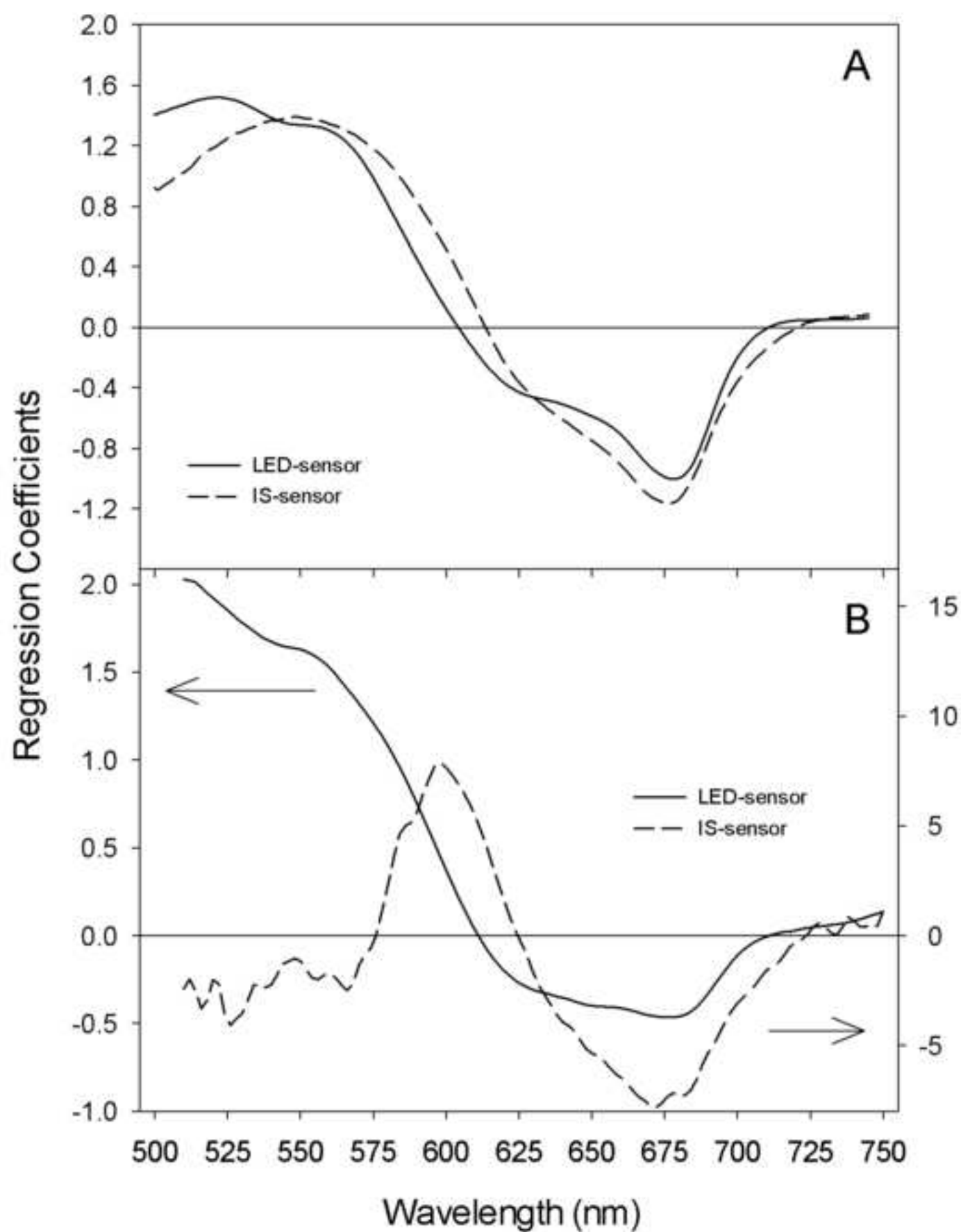


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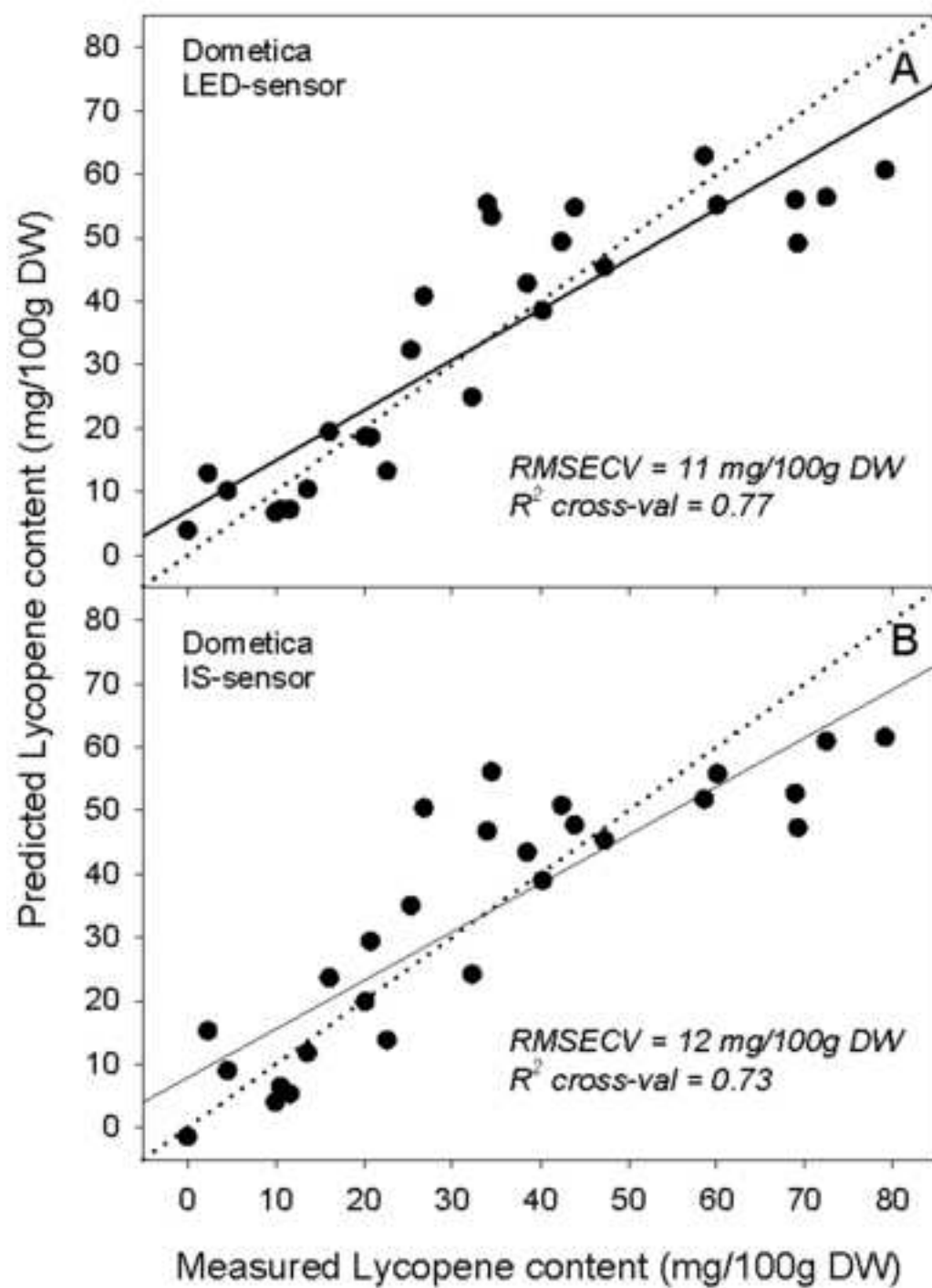
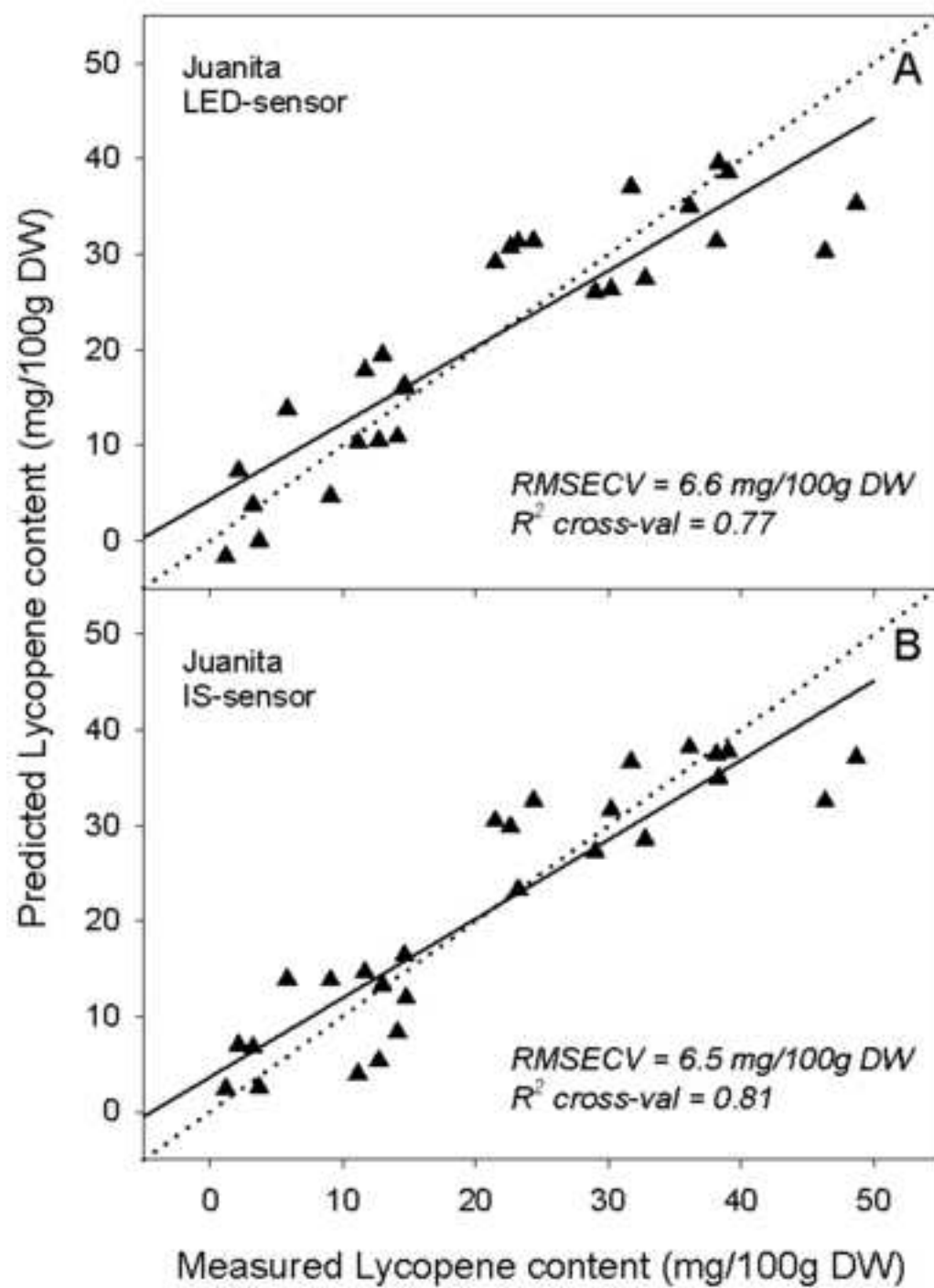


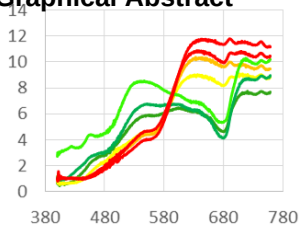
Figure 6
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*Highlights

- Integrating-sphere and LED-based sensors for tomato spectral reflectance detection
- Directional and total reflectance spectra of whole tomatoes are highly correlated
- Tomato lycopene was predicted non-destructively by reflectance and PLS regression
- Directional and total reflectance similarly predict tomato lycopene (16% error)

Graphical Abstract



Reflectance predicted Lycopene

