

# SUPPLEMENTARY MATERIAL

## From waste to function: valorization of collagen-based wastes with Natural Deep Eutectic Solvents for bioadhesive applications

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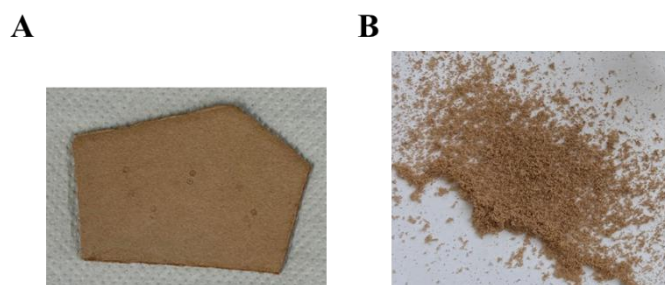


Figure S 1: Vegetable-tanned leather as supplied by the tannery (A) and after grating into small pieces (B).

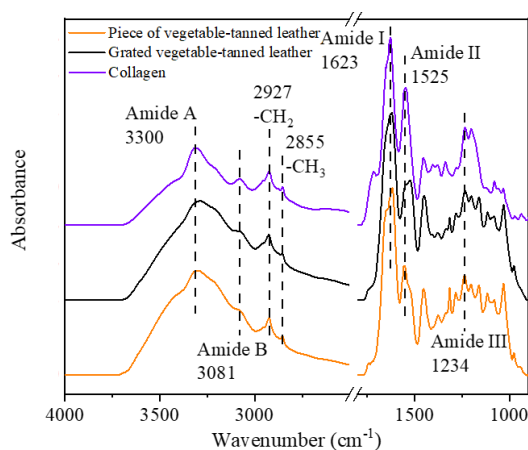


Figure S 2: ATR-FTIR spectrum of the collagen (purple line), vegetable-tanned leather sample as provided by the supplier (orange line) and grated vegetable-tanned leather (black line). Characteristic bands associated with collagen structure are highlighted.

Table S 4: TGA analysis of the samples: collagen, as-received tanned leather and grated tanned leather.

Sample	T <sub>onset</sub> (°C)	T (1° Peak) (°C)	T (2° Peak) (°C)	T max (°C)	Residue Mass % (T=900°C)	Mass loss (wt%)		
						Stage I (25-200°C)	Stage II (200-600°C)	Stage III (600-900°C)
Collagen	299.3	38.3	169.7	326.1	13	32	52	3
Leather as recived	268.8	46.2	/	309.6	26	15	54	5
Grated leather	263.7	39.1	/	310.3	23	11	59	7

\*Error (expressed as ± standard deviation) was evaluated by performing in triplicate representative samples. More in detail, error in the temperatures was ± 0.3°C, and in the mass losses was ± 4%.

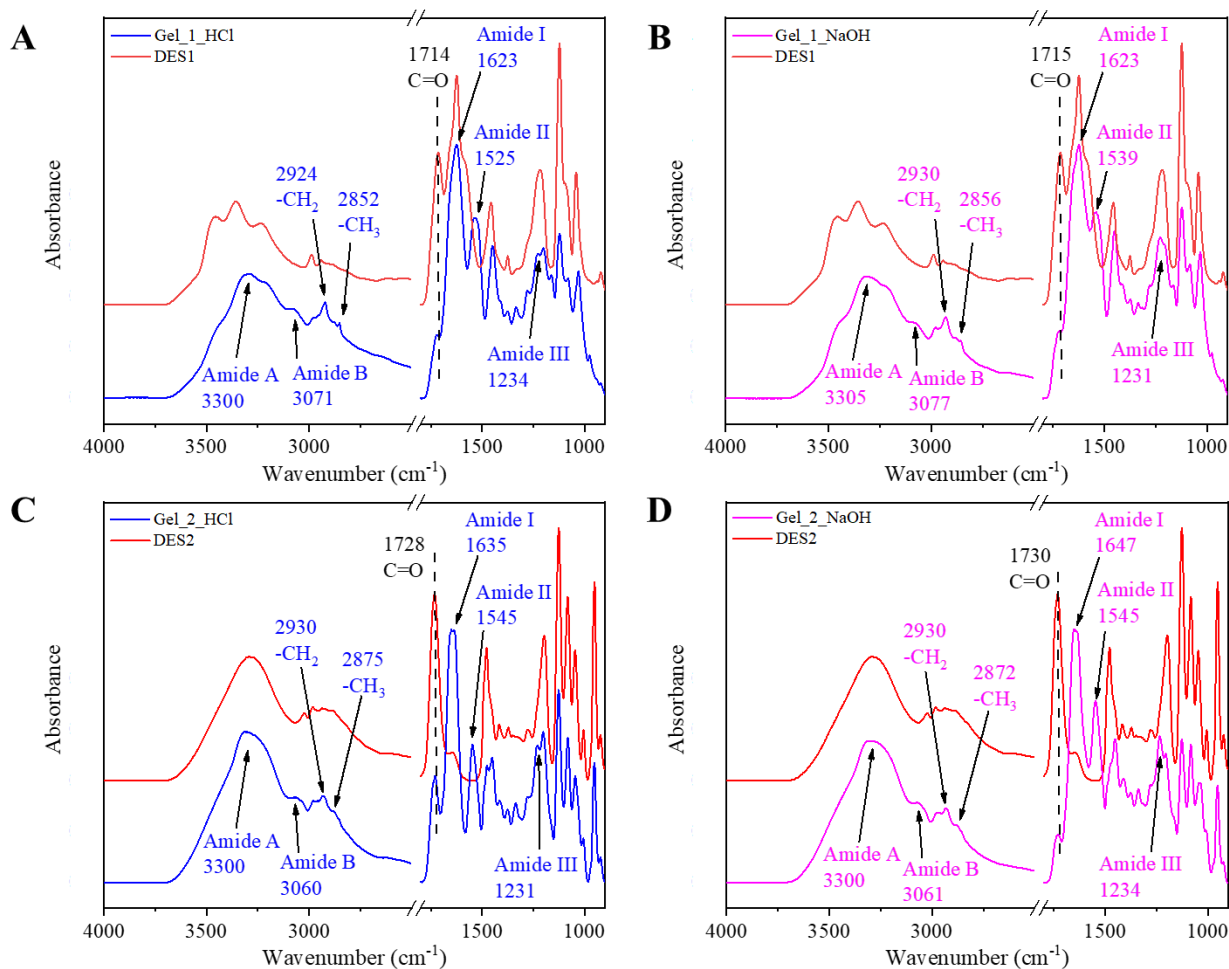


Figure S 3: ATR-FTIR spectra of samples analyzed.

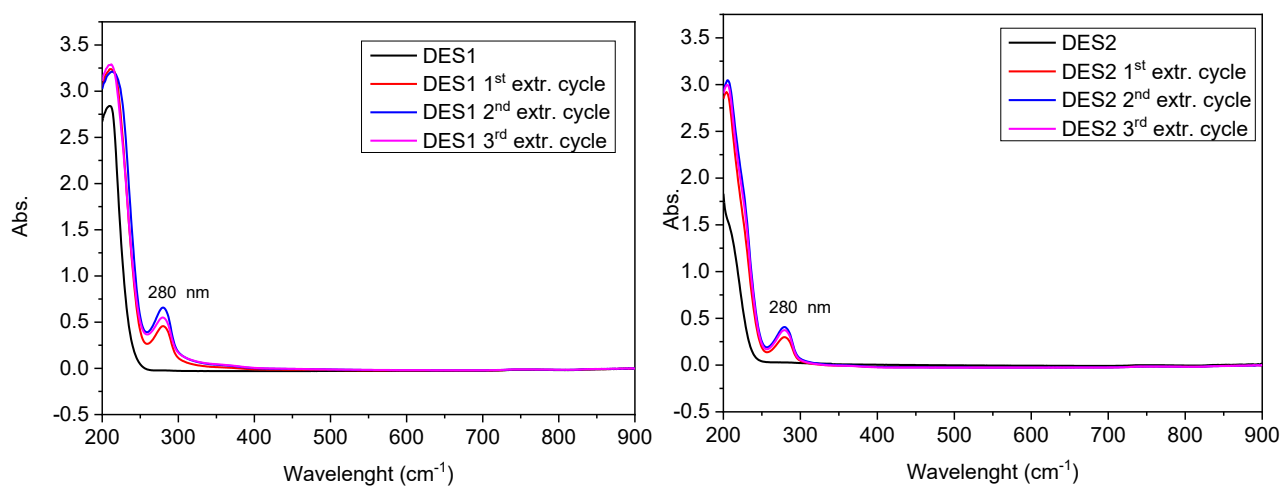


Figure S 4: UV-Vis of DES 1 (on the left) and DES 2 (on the right) used in subsequent cycles.

Samples are diluted 1:300 in water. The signal at 280 nm of mimosa and quebracho extracts containing tannins are visible (DOI: <http://dx.doi.org/10.1016/j.indcrop.2016.07.022>) while the signal of chestnuts extracts and the other signals of previous extracts are covered by the signal of DES.

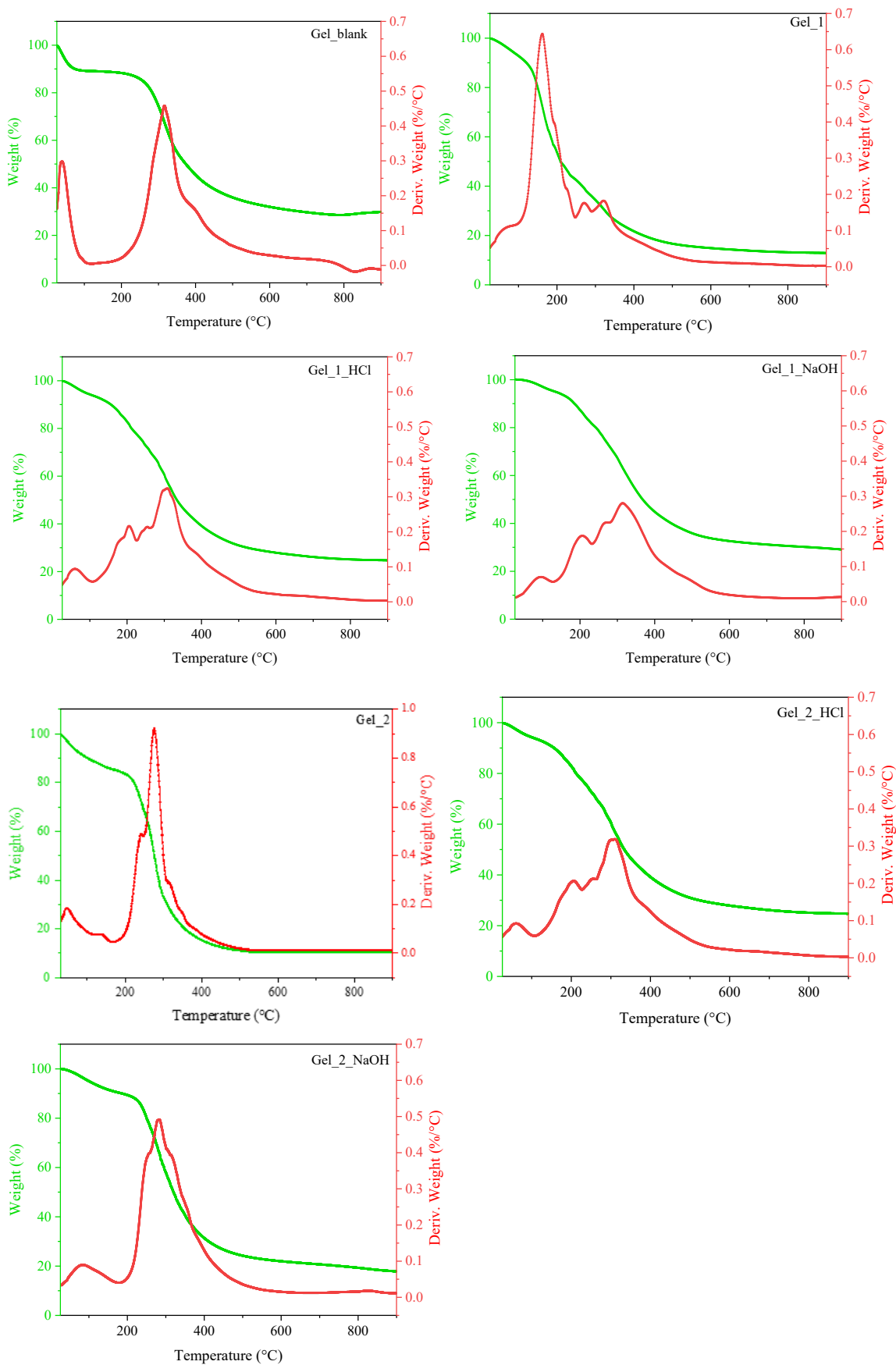


Figure S 5: TGA of the samples Gel\_blank, Gel\_1, Gel\_1 HCl, Gel\_1\_NaOH, Gel\_2, Gel\_2 HCl, Gel\_2\_NaOH.

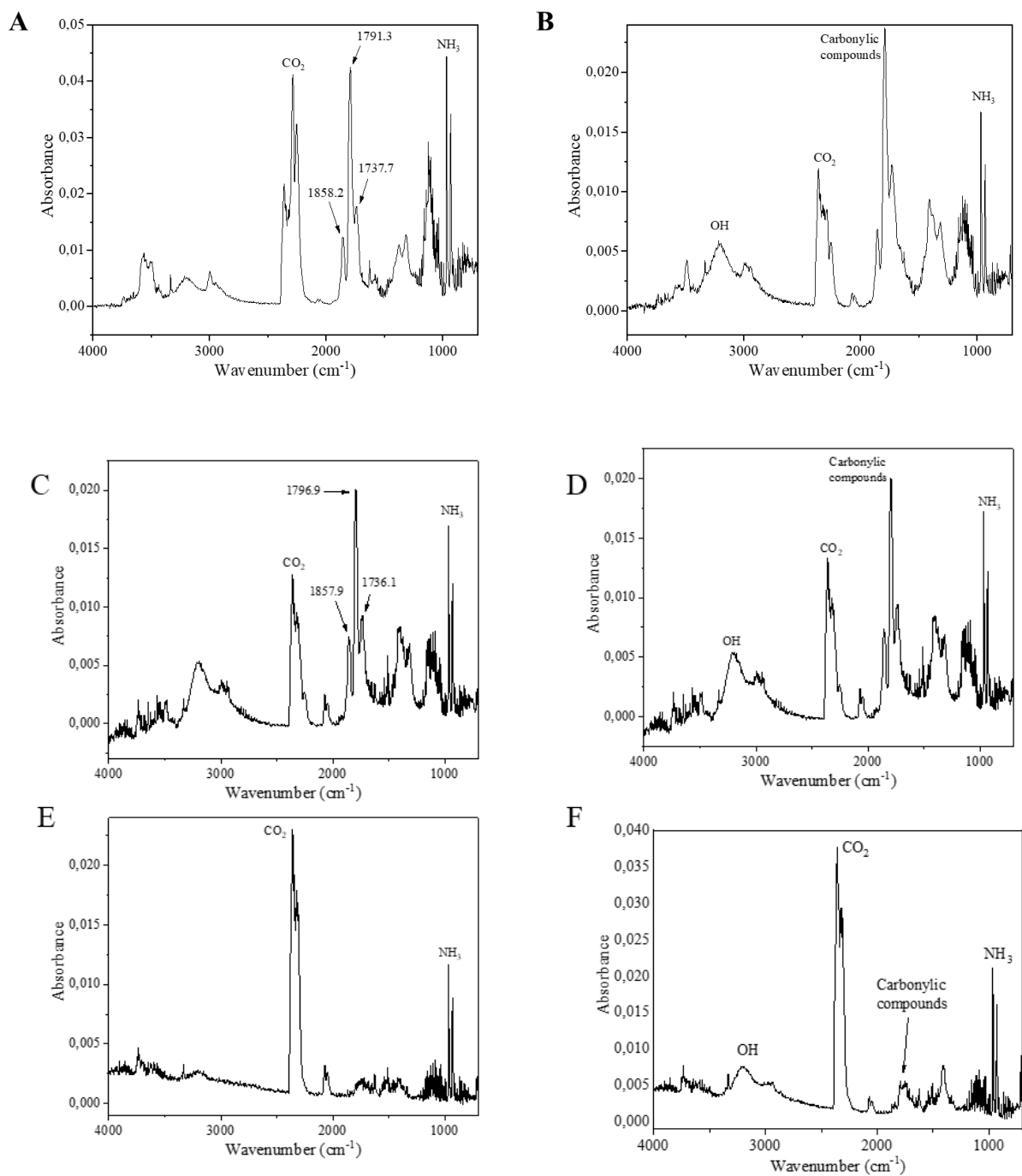


Figure S 6: FTIR spectrum of the gases evolved during thermal degradation of Gel\_1 at 163°C (A) and at 323°C (B), Gel\_1\_HCl at 210°C (C) and at 310°C (D) and Gel\_1\_NaOH at 210°C (E) and at 315°C (F), under nitrogen flow at a heating rate of 20 °C/min.

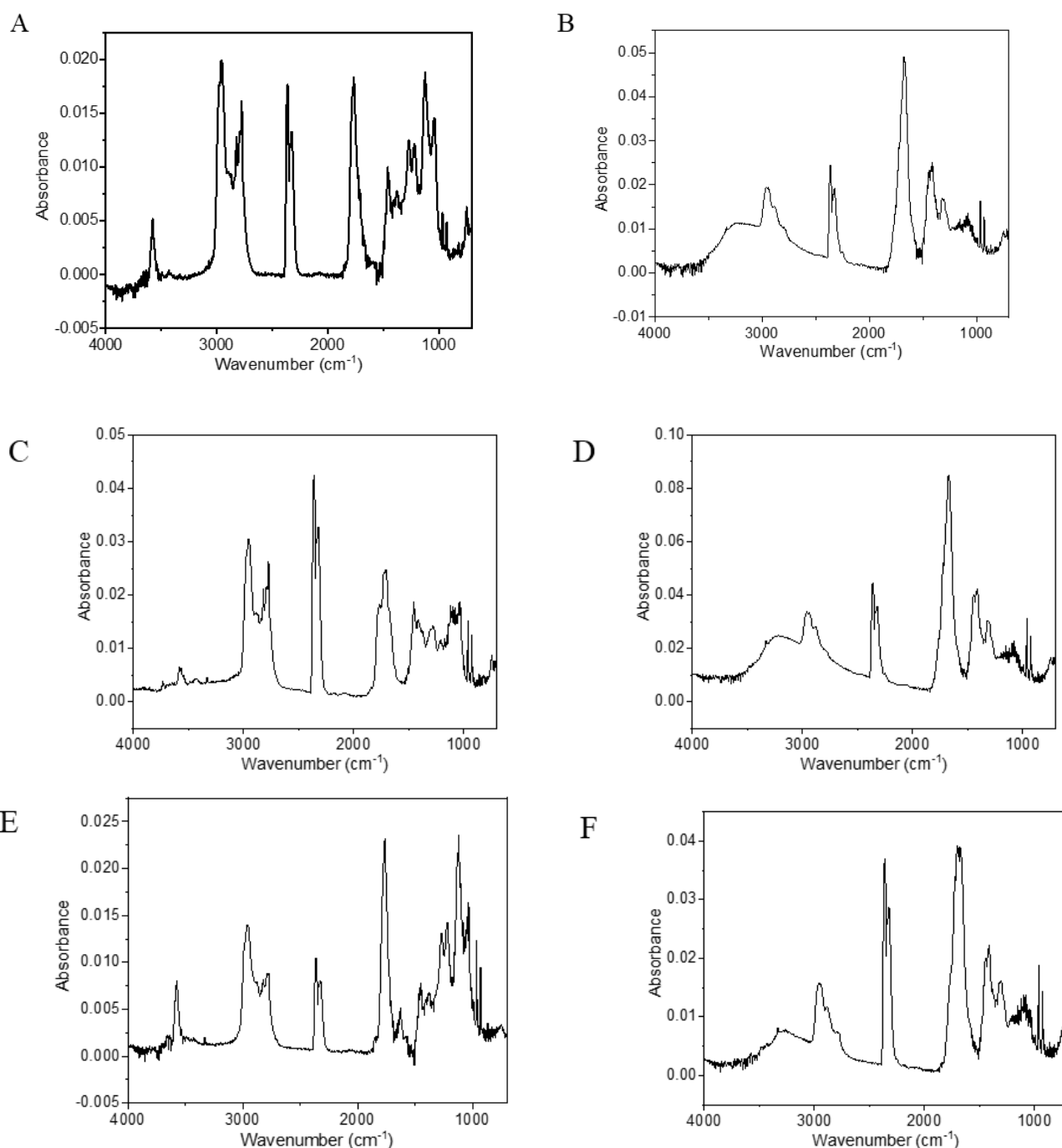


Figure S 7: FTIR spectrum of the gases evolved during thermal degradation of Gel\_2 at 220°C (A) and at 280°C (B), Gel\_2\_HCl at 220°C (C), and at 280°C (D) and Gel\_2\_NaOH at 220°C (E) and at 280°C (F), under nitrogen flow at a heating rate of 20 °C/min.

Table S 5: Maximum temperature in DTG associated with collagen degradation.

Sample	Collagen	Leather as recived	Grate d leathe r	Gel-blank	Gel_1	Gel_1_HCl	Gel_1_NaO H	Gel_2	Gel_2_HCl	Gel_2_NaOH
T max DTG (°C)	326	310	310	317	302	301	314	275	283	282

\*Error (<math>\pm 2^{\circ}\text{C}</math>, expressed as  $\pm$  standard deviation) was evaluated by performing in triplicate representative samples.

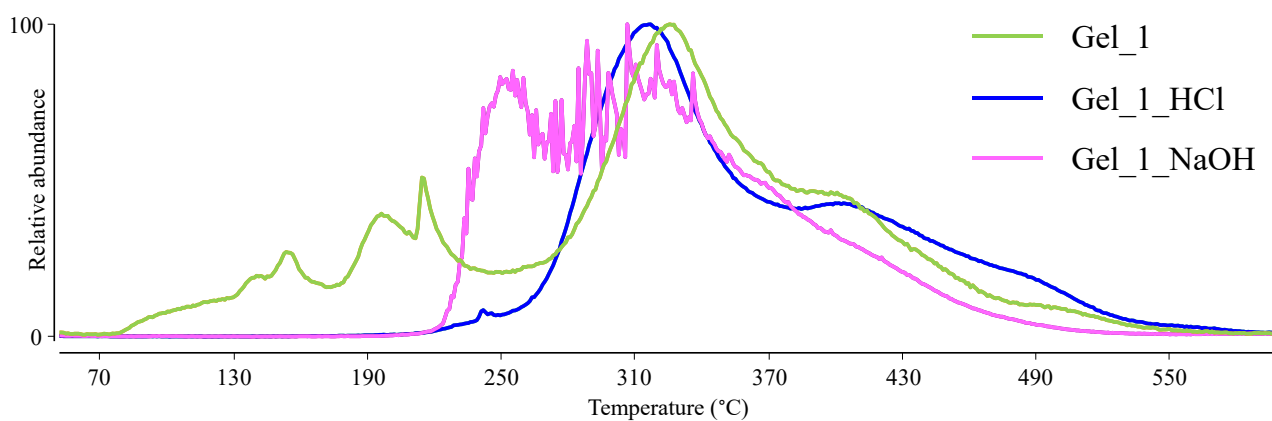


Figure S 8: Total Ion Thermograms (TITs) of Gel\_1 (green), Gel\_1\_HCl (blue) and Gel\_1\_NaOH (pink).

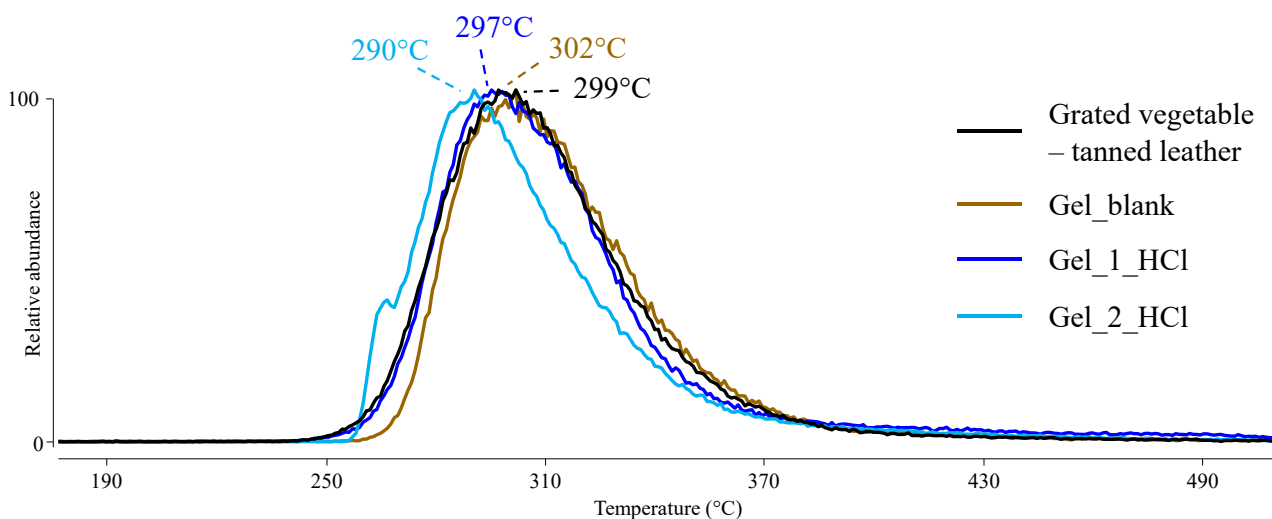


Figure S 9: Extracted Ion Thermograms (EITs) of fragment ion  $m/z$  154 of grated vegetable – tanned leather (black), Gel\_blank (brown), Gel\_1\_HCl (blue) and Gel\_2\_HCl (light blue).

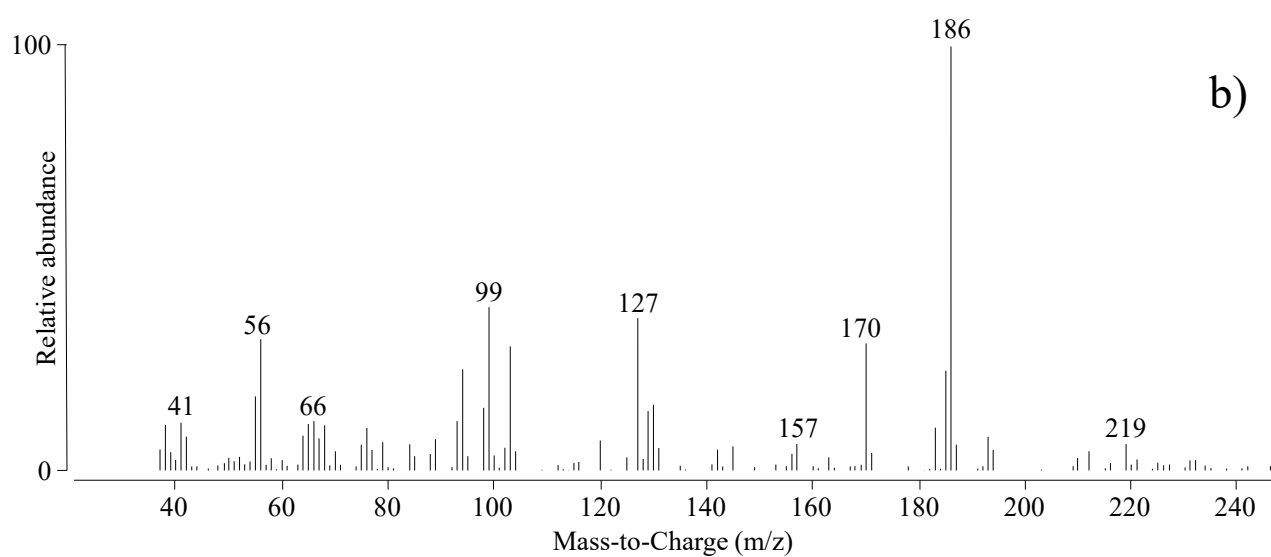
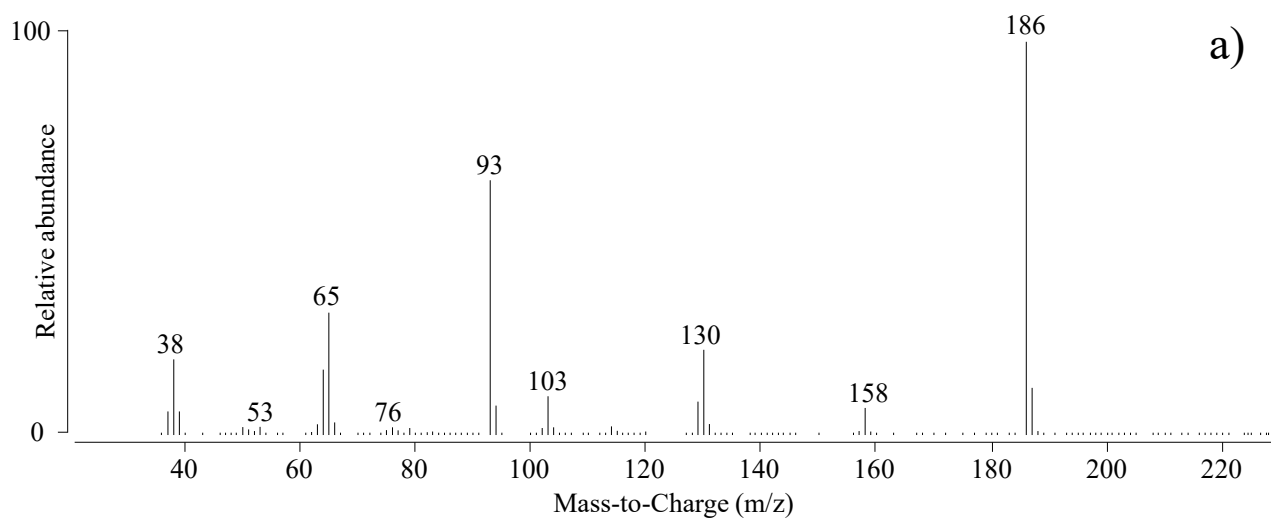


Figure S 10: Mass spectra of heteroaromatic compounds from EICs in Fig 4 at a) 19.1 min and b) 22.9 min.

**Table S 6:** Summary of protein identification results.

For each protein, data shows number of identified peptides, percentage of sequence coverage, number of peptide isoforms, and counts of unmodified and modified peptides. Modified peptide counts represent the number of peptide isoforms containing the specific modification used for calculating modification extent.

<i>Sample</i>	<i>Protein name (Uniprot ID)</i>	<i>Identified peptides</i>	<i>Sequence coverage(%)</i>	<i>Peptide isoforms</i>	<i>Unmodified Peptides</i>	<i>Oxidized M</i>	<i>Deamidated N</i>	<i>Deamidated Q</i>
<i>Gel_1_REP1</i>		77	58.8	255	31	14	24	33
<i>Gel_1_REP2</i>		46	42.9	145	11	16	16	15
<i>Gel_1_HCl_REP1</i>		72	59.3	289	30	20	37	53
<i>Gel_1_HCl_REP2</i>		57	53.4	297	25	14	89	49
<i>Gel_1_NaOH_REP1</i>		74	59.1	297	57	14	33	33
<i>Gel_1_NaOH_REP2</i>		68	57.3	363	47	14	55	41
<i>Gel_2_REP1</i>		74	57.4	300	42	20	23	35
<i>Gel_2_REP2</i>	<i>Collagen 1(I) (P02453)</i>	65	54.0	227	16	21	17	25
<i>Gel_2_HCl_REP1</i>	<i>alpha-chain</i>	78	61.0	310	50	19	29	48
<i>Gel_2_HCl_REP2</i>		80	59.3	346	55	18	33	44
<i>Gel_2_NaOH_REP1</i>		76	61.2	316	47	20	21	37
<i>Gel_2_NaOH_REP2</i>		75	60.4	326	53	21	28	38
<i>Gel_blank_REP1</i>		63	56.2	256	28	17	26	30
<i>Gel_blank_REP2</i>		46	44.4	105	14	14	9	15
<i>Grated_leather_REP1</i>		52	49.9	137	16	17	9	19
<i>Grated_leather_REP2</i>		30	31.6	52	10	11	6	4
<i>Gel_1_REP1</i>		53	54.9	108	8	5	28	7
<i>Gel_1_REP2</i>		28	33.6	56	4	0	13	4
<i>Gel_1_HCl_REP1</i>		62	66.9	165	9	8	43	14
<i>Gel_1_HCl_REP2</i>		51	56.0	140	6	7	43	14
<i>Gel_1_NaOH_REP1</i>		55	59.1	166	12	9	47	14
<i>Gel_1_NaOH_REP2</i>		54	56.6	182	17	8	49	12
<i>Gel_2_REP1</i>		55	58.3	132	10	10	36	7
<i>Gel_2_REP2</i>	<i>Collagen 2(I) (P02465)</i>	48	51.8	109	10	8	33	6
<i>Gel_2_HCl_REP1</i>	<i>alpha-chain</i>	62	64.9	171	14	9	42	13
<i>Gel_2_HCl_REP2</i>		64	68.9	210	11	12	52	13
<i>Gel_2_NaOH_REP1</i>		66	67.8	213	14	8	44	10
<i>Gel_2_NaOH_REP2</i>		63	66.7	203	11	12	41	8
<i>Gel_blank_REP1</i>		51	57.0	144	9	13	45	6
<i>Gel_blank_REP2</i>		33	36.6	63	4	5	19	5
<i>Grated_leather_REP1</i>		43	50.6	75	6	5	28	3
<i>Grated_leather_REP2</i>		22	26.6	33	2	1	7	2
<i>Gel_1_REP1</i>		55	70.1	168	7	7	42	21
<i>Gel_1_REP2</i>		33	50.5	70	4	8	21	8
<i>Gel_1_HCl_REP1</i>		56	66.9	168	6	16	48	20
<i>Gel_1_HCl_REP2</i>		53	65.9	195	8	13	68	29
<i>Gel_1_NaOH_REP1</i>		56	66.9	177	7	17	47	18
<i>Gel_1_NaOH_REP2</i>	<i>Collagen 1(III) (P04258)</i>	57	65.7	173	6	14	51	21
<i>Gel_2_REP1</i>	<i>alpha-chain</i>	59	70.9	188	3	18	48	25
<i>Gel_2_REP2</i>		46	61.2	125	4	13	37	12
<i>Gel_2_HCl_REP1</i>		64	70.2	197	7	20	56	18
<i>Gel_2_HCl_REP2</i>		59	68.6	192	8	24	47	20
<i>Gel_2_NaOH_REP1</i>		56	67.2	167	6	19	42	12
<i>Gel_2_NaOH_REP2</i>		59	68.9	185	8	17	45	21

<i>Gel_blank_REP1</i>	48	62.8	150	5	10	43	16
<i>Gel_blank_REP2</i>	36	44.7	70	3	8	22	6
<i>Grated_leather_REP1</i>	37	50.1	87	3	11	27	10
<i>Grated_leather_REP2</i>	20	26.9	35	1	5	9	3

**Table S 4:** Overall extent of backbone cleavage.

Data shows the mean ratio of semitryptic to total peptides across samples along with its standard error. Semitryptic/TOT ratios indicate the extent of backbone cleavage in samples, where higher ratios reflect increased protein degradation. Total counts of identified tryptic and semitryptic peptides backbone cleavage data is based on are reported for each sample replicate.

<i>Sample</i>	<i>Experiment</i>	<i>Total Tryptic</i>	<i>Total Semi-tryptic</i>	<i>Semitryptic/TOT (Mean)</i>	<i>Std Error</i>
<i>Gel_1</i>	<i>Gel_1_REP1</i>	199	25	0.11	0.00
	<i>Gel_1_REP2</i>	120	16		
<i>Gel_1_HCl</i>	<i>Gel_1_HCl_REP1</i>	454	146	0.25	0.01
	<i>Gel_1_HCl_REP2</i>	466	164		
<i>Gel_1_NaOH</i>	<i>Gel_1_NaOH_REP1</i>	480	68	0.14	0.01
	<i>Gel_1_NaOH_REP2</i>	470	81		
<i>Gel_2</i>	<i>Gel_2_REP1</i>	317	33	0.11	0.02
	<i>Gel_2_REP2</i>	305	45		
<i>Gel_2_HCl</i>	<i>Gel_2_HCl_REP1</i>	477	83	0.15	0.00
	<i>Gel_2_HCl_REP2</i>	517	86		
<i>Gel_2_NaOH</i>	<i>Gel_2_NaOH_REP1</i>	486	102	0.15	0.02
	<i>Gel_2_NaOH_REP2</i>	515	81		
<i>Gel_blank</i>	<i>Gel_blank_REP1</i>	282	22	0.07	0.00
	<i>Gel_blank_REP2</i>	99	7		
<i>Grated_leather</i>	<i>Grated_leather_REP1</i>	62	5	0.08	0.01
	<i>Grated_leather_REP2</i>	38	4		

**Table S 5:** Analysis of protein modification extent.

Mean values and standard errors for the extent of methionine (M) oxidation and deamidation (N|Q) are presented by sample, along with the number of detected and modified amino acid sites the data is based on. N|Q represents combined deamidation at both asparagine (N) and glutamine (Q) residues.

<i>Sample</i>	<i>Oxidation (M)</i>				<i>Deamidation (N Q)</i>			
	<i>Detected M</i>	<i>Modified M</i>	<i>Fraction of modified/TOT</i>	<i>Std Error</i>	<i>Detected N Q</i>	<i>Modified N Q</i>	<i>Fraction of modified/TOT</i>	<i>Std Error</i>
<i>Gel_1</i>	77	67	0.86	0.01	1094	485	0.33	0.02
<i>Gel_1_HCl</i>	76	65	0.86	0.01	1092	394	0.44	0.06
<i>Gel_1_NaOH</i>	49	42	0.86	0.03	624	198	0.36	0.00
<i>Gel_2</i>	99	90	0.89	0.04	1168	403	0.31	0.00
<i>Gel_2_HCl</i>	106	92	0.90	0.00	1085	334	0.34	0.00
<i>Gel_2_NaOH</i>	70	63	0.87	0.03	747	229	0.31	0.02
<i>Gel_blank</i>	58	48	0.85	0.10	509	177	0.37	0.05
<i>Grated_leather</i>	35	31	0.90	0.00	236	96	0.40	0.00

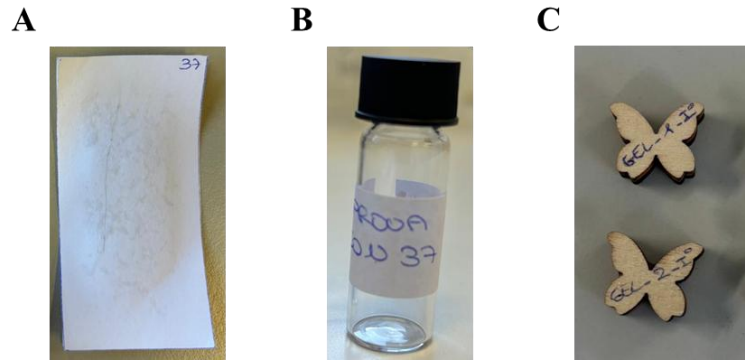


Figure S 11: Adhesion tests with Gel\_1 as bioadhesive. A) paper–paper, B) paper–glass, and C) wood–wood.

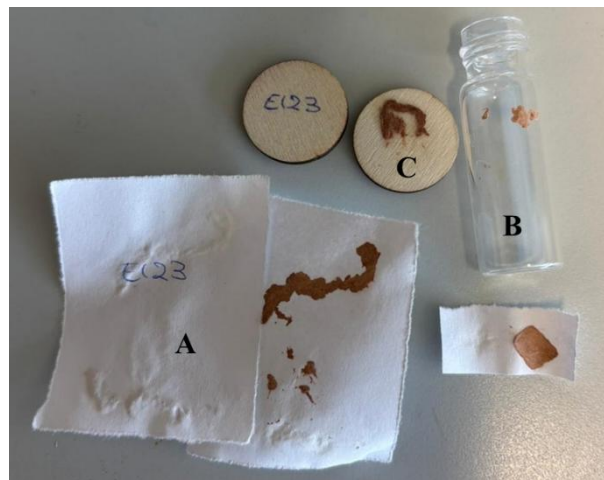


Figure S 12: Adhesion test with control experiment (Gel-blank). A) paper–paper, B) paper–glass, and C) wood–wood.

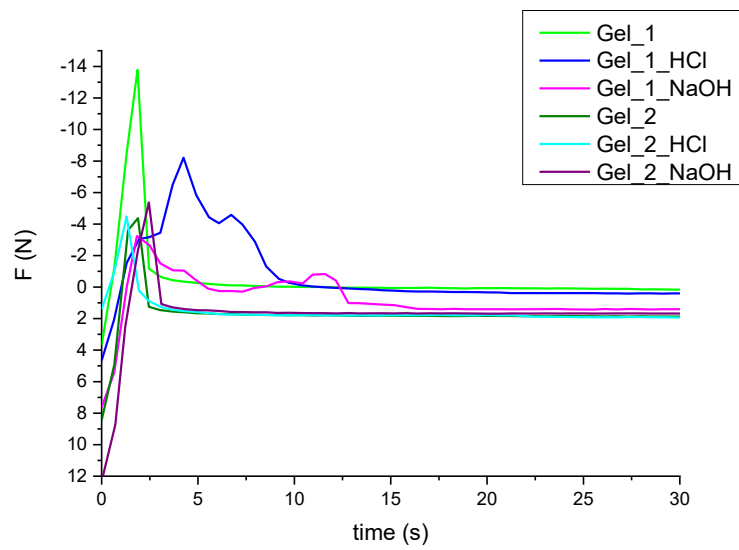


Figure S 13: tack test made on the samples to test the adhesive strength on steel–wood surfaces.

Table S 6: Scores attributed to each principle to evaluate the process greenness, made according to Path2Green application.

<b>Principles</b>		<b>Score*</b>	<b>Motivation</b>	<b>Weight**</b>
<b>1</b>	<b>Biomass</b>	+1	Waste biomasses	6
<b>2</b>	<b>Transport</b>	+0.5	40 km distance	5
<b>3</b>	<b>Pre-treatment</b>	-0.2	Physical pretreatment	2.5
<b>4</b>	<b>Solvent</b>	+1	Recommended solvents (DESs)	6
<b>5</b>	<b>Scaling</b>	-1	In batches	3
<b>6</b>	<b>Purification</b>	-0.5	Water	2.5
<b>7</b>	<b>Yield</b>	+1	Complete valorization of the biomass	4
<b>8</b>	<b>Post-treat.</b>	+1	Ready-to-use extract	2.5
<b>9</b>	<b>Energy</b>	+0.5	Low energy dependence	5
<b>10</b>	<b>Application</b>	+0.66	To be applied in four domains	4.5
<b>11</b>	<b>Repurposing</b>	+1	Non-Virgin raw materials are used	6
<b>12</b>	<b>Waste</b>	-1	Waste generated: 100% m/m	6

\*Scores and weights were chosen following the recommendations of (DOI: 10.1039/d4gc02512a).