

**Preferential removal of pesticides from water by molecular  
imprinting on TiO<sub>2</sub> photocatalysts**

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**Abstract**

In order to achieve a selective removal of specific pesticides from water, we synthesized, through the sol-gel technique, molecularly imprinted TiO<sub>2</sub> photocatalysts with the only use of the standard reactants for the TiO<sub>2</sub> sol-gel synthesis together with the pesticide molecules, without any addition of further reactants, supports or matrices. It is a new, easy, smart and scalable method that avoid the multistep and solvent-consuming procedures, typical of the molecular imprinting. Two widely-used pesticides, i.e. the herbicide 2,4D, and the insecticide imidacloprid, were chosen as template for the molecular imprinting and as contaminants target for the photocatalytic tests. A remarkable enhancement of the photocatalytic activity was verified with the TiO<sub>2</sub> imprinted with the corresponding pesticide-target. The selectivity of the photodegradation process was verified thanks to the comparison with the degradation of pesticides not-used as template. Furthermore, the eventual toxic effects of the molecularly imprinted materials were evaluated by biological tests.

The combination of molecular imprinting with photocatalysis, here investigated for the first time with pesticides, it is a promising strategy to selectively catch (through the molecular imprinting process) and degrade specific organic contaminants from water (through the photocatalysis).

**Keywords:** Photocatalysis, Titanium dioxide, Water purification, Pesticides, Molecular Imprinting.

## 1. Introduction

The intensification of agricultural practises to fulfil the ever-increasing global demand of food has generated, as side effect, an enhanced contamination of air, water, soil and aquatic ecosystems [1,2]. In this contest, one of the major problems of the last years is the pollution of surface and ground waters by pesticides. The harmful effect of these chemical compounds arises from their toxicity and from the high mobility and persistence in the aqueous media [3,4]. Generally, in fact, only a low percentage of the applied pesticides is able to protect the agriculture products, while the major portion of pesticides is lost in the environment through volatilization, hydrolysis, degradation by photolysis or by microbial action [5]. The negative consequence is the high cost of agricultural production with a heavily-polluted environment.

Heterogeneous photocatalysis is recently emerged as a totally-green technology for environmental applications [6-10]. In particular, titanium dioxide ( $\text{TiO}_2$ )-based photocatalysis is considered today an efficient methodology in the area of wastewater treatment [11-17] and for hydrogen production by photocatalytic water splitting [18,19].

$\text{TiO}_2$  (commonly called *titania*) is an abundant, inexpensive, and nontoxic semiconductor material. The production of hydroxyl radicals, formed by the interaction of oxygen and water with the photoelectron-hole pairs (generated after the exposure of  $\text{TiO}_2$  to UV irradiation), allows to degrade a wide variety of bio-recalcitrant compounds [20,21]. Despite the countless advantages of the photocatalysis, one drawback is the non-selectivity of the process. Usually indeed, the water effluent streams contain a mixture of high toxic organic and non-biodegradable pollutants and less toxic organic and biodegradable compounds. Normally, the high toxic compounds are present in low concentrations, whereas the less toxic and biodegradable compounds are the major

components [22,23]. When TiO<sub>2</sub> is used to purify this type of water, the low and/or non-toxic organic contaminants at high concentrations are firstly removed, while the highly toxic organic pollutants at low concentrations are difficult to degrade. Furthermore, the selectivity of the photocatalytic process could be important in some industrial applications dealing with aqueous mixture containing compounds that should be removed and other compounds that should be recovered [24]. Therefore, the achievement of a selective TiO<sub>2</sub>-based photocatalysis process is a fundamental task to obtain an efficient water remediation treatment.

Different methodologies have been investigated to enhance the selectivity of TiO<sub>2</sub>, such as the pH variation [25,26] to control the surface electric charge, the surface functionalization with specific molecules that can afford a selective and effective adsorption of specific organic pollutants [27,28], the doping process [29, 30], or the modification of the crystallinity of the photocatalysts [31]. An innovative and effective strategy to design TiO<sub>2</sub> photocatalyst able to enhance the selectivity degradation of the organic contaminants can be obtained by the molecular imprinting (MI) process.

Since the first work of M. V. Polyakov in 1931 [32], the molecular imprinting technique has been recognized as a performing method to produce materials with a memory of the size, shape, and chemical composition of the template molecules [33-37]. The synthesis method consists in favouring the interaction of the template molecules onto an organic or inorganic matrix during its preparation step; a subsequent removal of the imprinted molecule (i.e., the template) leads to cavities complementary to the template. The combined approach between molecular imprinting and photocatalysis leads to obtain a selective photodegradation; in detail, the molecular imprinting process promotes a selective interaction between the TiO<sub>2</sub> and the dangerous contaminant,

whereas the photocatalytic process, activated by the titanium dioxide, efficiently degrades the specific pollutant.

Until now, there are some studies in the literature that analyse the performance of TiO<sub>2</sub> nanoparticles coated or supported with a layer of molecularly imprinted polymers (MIP). These materials exhibited specific affinity toward various target pollutants [38-43]. However, this approach has still some drawbacks. First of all, the synthesis of these imprinted photocatalysts usually required multistep procedures, needed for the preparation of the polymer matrix, which involve a relevant consumption of time and solvents; additionally, the polymeric matrix can cover the surface active sites of TiO<sub>2</sub>, hindering the light absorption and thus decreasing the photocatalytic activity; finally, the removal of the template from the polymer matrix can be sometimes difficult, due to the strong chemical interaction that can be formed between the polymer and the target molecules. To avoid such disadvantages, the development of materials with a molecularly imprinted inorganic framework (namely, without the utilize of organic polymers) can be a successfully strategy. Silica (SiO<sub>2</sub>) has been used as the inorganic matrix [44-47]. An alternative methodology consists in synthesizing TiO<sub>2</sub> without the use of a supporting matrix, adding in the first step of the titania synthesis the target pollutant and then removing it with a thermal treatment. This process allows to obtain a molecularly imprinted TiO<sub>2</sub>. This procedure is simple and environmental-friendly: the imprinted cavities can be shaped during the synthesis of titania, and the template molecules can be totally removed by calcination. To our knowledge, only few papers have reported this approach for the synthesis of imprinted TiO<sub>2</sub> photocatalysts as nanoparticles or as films [48-51], and no one has investigated the photocatalytic water purification from pesticides. Additionally, the selection of pesticides as target pollutants in the photocatalytic experiments are less investigated in the literature compared to other

molecules (such as, dyes or phenolic compounds), due the harder reaction conditions required for these type of contaminants [52, 53].

On the above considerations, we here report for the first time the experimental results of the preferential photodegradation of two widely used pesticides: the herbicide 2,4-dichlorophenoxyacetic acid (2,4D), and the insecticide imidacloprid (1-(6-chloro-3-pyridinylmethyl)-N-nitro-2-imidazolidinimine), using molecularly imprinted TiO<sub>2</sub> catalysts synthesized towards the sol-gel technique without the addition of organic or inorganic matrix.

The 2,4-D (Fig. 1, molecular length: 1.002 nm, estimated by ChemDraw<sup>®</sup>) belongs to the group of synthetic chlorophenoxy herbicides. It can be also used in combination with other herbicides for the post-emergence control of broad-leaved weeds [54]. It persists in the environment for a long time [55], and it has been identified by the International Agency for Research on Cancer as a possible human carcinogen (class 2B-carcinogen) [56]. The imidacloprid (Fig. 1, molecular length: 1.078 nm, estimated by ChemDraw<sup>®</sup>) is a neonicotinoid insecticide having outstanding effectiveness and systemic action for crop protection against piercing-sucking pests. The wide use of imidacloprid causes, for its high leaching potential [57], soil pollution, surface/underground water contamination, and destruction of biological systems [58]. Severe exposure to imidacloprid can cause thyroid lesions, lethargy and spasms, and it can harm the mammalian reproduction [59,60].

The photocatalysts were characterized by scanning electron microscopy (SEM), transmission electron microscopy (TEM), X-ray diffraction (XRD), N<sub>2</sub> adsorption-desorption measurements, Fourier Transform Infrared (FTIR), X-ray photoelectron (XPS), UV-vis diffuse reflectance spectroscopy (DRS), and point of zero charge (pzc) determination.

The toxicity of the samples was assessed by the zebrafish embriotoxicity test (ZFET), according to the Organization for Economic Co-operation and Development (OECD, 2013) [61]. ZFET is a modern non-animal test, representing an effective alternative to acute test with adult fish. [62] Moreover, it is an excellent test for the valuation of toxicity of micro/nanoparticles [63-65]. The metallothioneins 1 (MTs1), a group of low molecular mass proteins, were analysed as biomarkers of exposure. These proteins are involved in heavy metal ion homeostasis and detoxification [66].

## **2. Experimental**

### ***2.1. Catalyst preparation***

Titanium(IV) butoxide, 2,4-dichlorophenoxyacetic acid (2,4D) and imidacloprid, were purchased from Sigma-Aldrich; ethanol and glacial acetic acid from VWR. All the chemical substances were used without any further purification. The molecularly imprinted TiO<sub>2</sub> samples were synthesized using the sol-gel technique [67]. The stoichiometric amount of pesticide required to obtain a molar ratio of 5:1 between TiO<sub>2</sub> and 2,4D or imidacloprid was dissolved in a solution of 0.5 ml of acetic acid and 1.5 ml of ethanol; then the solution was stirred at room temperature until a complete solubilization of the pesticide. Successively, 2 ml of titanium butoxide was added in the mixture and it was stirred for further 10 min. Then, a solution containing 2 ml of demineralized water, 2 ml of acetic acid and 2 ml of ethanol was added dropwise to the above solution. The resultant slurry was stirred for 3 hrs and aged for 24 hrs. The obtained wet gel was dried at 100 °C for 12 hrs and then calcined in air at 500 °C for 6 hrs (heating ramp 2 °C/min) to remove the template/pesticide molecules and the organic substances formed during the synthesis. The samples will be hereafter called “TiO<sub>2</sub> MI/2,4 D” and “TiO<sub>2</sub> MI/Imid”. The imprinted titania without the calcinations process



will be identified as “TiO<sub>2</sub> MI/2,4 D Not Removed” and “TiO<sub>2</sub> MI/Imid. Not Removed”. The bare TiO<sub>2</sub> (simply called “TiO<sub>2</sub>”) was prepared with the same procedure described above without the use of pesticides.

## ***2.2. Catalyst characterization***

The morphology of the synthesized samples was investigated by SEM with a field emission Zeiss Supra 25 microscope, and by TEM with a JEOL ARM200F Cs-corrected, operated at 200 keV. In more detail, the TEM images were acquired in Bright Field TEM mode (BF-TEM) and high resolution mode (HR-TEM). The X-ray diffraction (XRD) analyses were performed with a Bruker D-500 diffractometer, operating with a parallel Cu-K $\alpha$  radiation at 40 kV and 40 mA,  $2\theta$  from 20° to 60°, in grazing incidence mode (0.8°). The XRD patterns are reported in counts per second (cps) versus  $2\theta$ , and they were examined by the Bruker software suite, including ICSD structure database. The textural properties of the materials were determined by Brunauer-Emmett-Teller (BET) adsorption-desorption of N<sub>2</sub> at -196 °C using a Micromeritics Tristar II Plus 3020 with a pre-treatment of out-gassing at 100 °C overnight. The FTIR spectra were obtained through a Perkin-Elmer Spectrum 1000 spectrometer (accuracy of wavenumber  $\pm 0.01$  cm<sup>-1</sup>, resolution 4 cm<sup>-1</sup>). The investigated samples, in the form of tables, were obtained by mixing the catalysts powders (1 mg) with KBr powders (300 mg) in an Agate mortar and pressing with a press. The XPS analyses were performed at 45° take-off angle relative to the surface plane with a PHI 5600 Multi Technique System (base pressure of the main chamber:  $1 \times 10^{-8}$  Pa) [68, 69]. Samples were excited with Al K $\alpha$  X-ray radiation using a pass energy of 5.85 eV. Structures due to the K $\alpha$  satellite radiations were subtracted from the spectra prior to data processing. The XPS peak intensities were obtained after Shirley background removal [68,69]. The atomic concentration analysis was performed by taking into

account the relevant atomic sensitivity factors. The instrumental energy resolution was  $\leq$  0.5 eV. Spectra calibration was achieved by fixing the main C 1s signal at 285.0 eV. The point of zero charge (pzc) of the catalysts was established with the Noh-Schwarz method of mass titration [70]. This method involves to find the asymptotic value of the pH of an oxide/water slurry as the oxide mass content is increased. The typical values of oxide/water by weight were 0.1, 1, 5, 10, 20, 40%. DRS spectra were recorded in the 200-700 nm wavelength range, using a Perkin-Elmer Lambda 40 UV-vis spectrophotometer.

### ***2.3. Photocatalytic activity experiments***

The selective photodegradation of 2,4D and imidacloprid pesticides were investigated under UV irradiation using an UV lamp centred at 368 nm (emission spectrum showed in the Fig. S1C), with a full width at half maximum lower than 10 nm, and an irradiance of 4 mW/cm<sup>2</sup>, which simulate the UV irradiance of the sun on the Earth. Prior to carry out the measurements, the catalysts were irradiated by the UV lamp for 60 min in order to remove the hydrocarbons and the adventitious carbon from the samples' surface [71]. A quantity of 1 mg of catalyst powders was immersed in 4 ml of aqueous solution of 2,4D or imidacloprid in quartz cuvettes. The starting concentration of 2,4D and imidacloprid was fixed to  $5 \times 10^{-5}$  M for both organic contaminants. The solutions were treated by ultrasonic irradiations with a frequency of 50 Hz for 5 min, to favour the dispersion of the powders. Control experiments were carried out in the dark for 90 min to ensure the adsorption-desorption equilibrium of the pesticides on the beaker's surface and on the samples' surface. At regular time intervals (for a total time of 3 hrs), 2 ml of the irradiated solutions were measured with an UV-vis spectrophotometer (Lambda 45, Perkin-Elmer), in a wavelength range of 200-700 nm. Before any measurements, the samples were centrifuged for 3 min with a spin of 1500

rpm, to remove the powders and reduce the light scattering phenomena during the spectrophotometric measurements. The degradation process of 2,4D and imidacloprid was evaluated by the absorbance peaks at 229 nm and 270 nm (Figs. S1A-B), respectively, in the Lambert-Beer regime [72]. The degradations of 2,4D and imidacloprid in the absence of any photocatalyst were also evaluated as references.

The photocatalytic tests in presence of a mixture solution of 2,4D and imidacloprid (2 ml 2,4D  $5 \times 10^{-5}$  M + 2 ml imidacloprid  $5 \times 10^{-5}$  M) were also investigated, following the same experimental conditions reported above.

The selectivity of the photodegradation process was verified thanks to the comparison with the degradation of the pesticides not-used as template, and also using another pesticide: the fungicide ortho-phenylphenol ( $5 \times 10^{-5}$  M).

The mineralization of the 2,4D and imidacloprid pesticides after the photocatalytic process was evaluated by measuring the total organic carbon (TOC) content with a Shimadzu TOC-LCSH analyzer (tested volume: 3 ml) equipped with a non-dispersive infrared detector. The inorganic carbon was removed from the tested solutions using the Non-Purgeable Organic Carbon (NPOC) method by adding phosphoric acid and successively purging by dry air, then the TOC content was measured after a high-temperature catalytic oxidation at 680 °C.

The photodegradation products were determined by the Electrospray Ionization (ESI) mass analysis. The measurements were performed by a Thermo Finnigan LCQ-DECA ion trap mass spectrometer (Thermo Fischer Scientific, San Jose, CA), equipped with an Atmospheric Pressure Ionization (API) source. Mass spectra were acquired in negative ion mode in the  $m/z$  range 150-225 under the following conditions: capillary temperature 130 °C nebulizer gas (nitrogen) with a flow rate of 15 arbitrary units; source voltage 4 kV; capillary voltage 46 V; tube lens voltage 90 V. After

photocatalysis, each solution was injected (direct infusion) into the mass spectrometer, at a flow rate of 10  $\mu\text{L}/\text{min}$ . The mass spectrometer was calibrated using a standard mixture sodium dodecyl sulfate (265.17 Da), Ultramark (1621 Da) and sodium taurocholate (514,42 Da).

#### **2.4. Zebrafish embryotoxicity test**

In order to test the eventual toxicity of the synthesized materials, zebrafish eggs fertilized within 4 hrs post fertilization (hpf) were provided from the Center of Experimental Ichthyopathology of Sicily (CISS), University of Messina (Italy). Zebrafish embryos were exposed to bare  $\text{TiO}_2$ , and to test the eventual toxicity changes due to the molecular imprinting process, to the  $\text{TiO}_2$  MI/Imid, used as model sample for the molecularly imprinted  $\text{TiO}_2$  materials. Two concentrations ( $14 \times 10^{-5}$  and  $14 \times 10^{-6}$  mg/ml) in 5 ml of freshwater for 4-96 hpf were measured for evaluating the possible toxic effects of the synthesized powders on the zebrafishes. The solutions were renewed and embryonic/larval mortality and hatching rate were evaluated every 24 hrs. According to *Ref.* [65], healthy embryos were placed in 24-well culture plates (10 embryos in 5 ml solution/well). Each group had five replicate wells. Each experiment was repeated four times. Two larvae of each group were used for immunodetection of biomarkers by immunofluorescence according to *Ref.* [73]. The primary antibody anti-mouse-methallotionein 1 (1:500, from Abcam) and secondary antibody with fluorescein tetramethylrhodamine (TRITC) conjugated goat anti-mouse IgG (1:1,000, form Sigma-Aldrich) were used. Negative controls were performed by incubation with anti-mouse sera without antibodies. Observations were carried out using confocal laser scanning microscopy (CLSM; Zeiss LSM 700), equipped with the ZEN-2011 software and with a red fluorescence filter set.

### 3. Results and discussion

The SEM images of TiO<sub>2</sub>, TiO<sub>2</sub> MI/2,4D and TiO<sub>2</sub> MI/Imid powders are reported in **Fig. 2**. All the samples showed a rough morphology with heterogeneous shaped particles. The non-homogenous morphology is typical of the TiO<sub>2</sub> matrix-free sol-gel synthesis [74-76]. The inset of **Fig. 2A** depicts a high magnification of the material.

The TEM analyses (**Fig. 3 and S2**) performed on TiO<sub>2</sub> and TiO<sub>2</sub> MI/2,4D samples evidenced a granular morphology, irrespective of the molecular imprinting process. The diffraction pattern of the TiO<sub>2</sub> sample (**inset of Fig. 3A**) unequivocally showed a polycrystalline anatase phase of the synthesized titania, in good agreement with the literature [77]. The diffraction pattern obtained for the TiO<sub>2</sub> MI/2,4D (**inset of Fig. 3B**) does not show any change in the crystalline structure, due to the molecular imprinting process.

XRD analyses are reported in **Fig. 4**. All the XRD patterns are typical of the TiO<sub>2</sub> in the anatase phase (JCPDS number 21-1272), as confirmed by the signal at  $2\theta = 25.3^\circ$  (101),  $37.8^\circ$  (004),  $38.5^\circ$  (112),  $48.0^\circ$  (200),  $54.0^\circ$  (105), and  $55.1^\circ$  (211), and in agreement with the TEM analyses (**insets of Fig. 3**), consequently the diffraction analyses do not evidence substantial modifies in the crystalline structure of molecularly imprinted samples with respect to the reference one (i.e., TiO<sub>2</sub>).

Some interesting variations in the textural properties were highlighted through the N<sub>2</sub> adsorption-desorption measurements. All of the samples exhibited the type-IV isotherm (not shown) typical of a mesoporous structure, as expected [78]. **Table 1** reported the BET surface area ( $S_{\text{BET}}$ ), the mean pore diameter ( $d_p$ ), and the pore volume ( $V_p$ ) of the investigated samples. The BET surface area of the bare TiO<sub>2</sub> is 59 m<sup>2</sup>/g and the molecular imprinting process caused a decrease of surface area of 4 m<sup>2</sup>/g for the TiO<sub>2</sub>

MI/2,4D and of 6 m<sup>2</sup>/g for the TiO<sub>2</sub> MI/Imid. Consequently, an increase in the mean pore diameter and in the pore volume was detected compared to un-modified TiO<sub>2</sub> (Table 1). The BJH (Barrett, Joyner and Halenda) method [79] was used to calculate the pore size distributions of the samples from experimental isotherms. Fig. 5 reports the differential variation of the pore volume per pore unit as a function of the pore diameter. A clear shift towards larger pores is detected due to the molecular imprinting process. In detail, the TiO<sub>2</sub> MI/Imid catalysts exhibited the largest distribution, with a mean pore diameter of 9.7 nm (Tab. 1), about 4 nm larger than the mean pore diameter of the TiO<sub>2</sub> MI/2,4D catalyst, and about 5 nm larger than the mean pore diameter of bare TiO<sub>2</sub>. The differences in the textural properties between the bare TiO<sub>2</sub> and the molecularly imprinted samples can be reasonably due to the molecular imprinting process which induces the modifications of the pores already existing in the structure of the not-imprinted TiO<sub>2</sub>.

FTIR measurements were carried out to check the effectiveness of the molecular imprinting process. The FTIR results are reported in Fig. 6. In particular, Figure 6A displays the FTIR spectra of 2,4D, TiO<sub>2</sub> MI/2,4D Not Removed (i.e. before the calcination), TiO<sub>2</sub> MI/2,4D, and TiO<sub>2</sub>. The 2,4D pesticide showed a peak at 1736 cm<sup>-1</sup> which is related to the presence of C=O of the carboxyl group [80]. The peaks at 1478 cm<sup>-1</sup> and 1431 cm<sup>-1</sup> indicates the C=C vibrations of the aromatic ring and the CH<sub>2</sub> vibrations of alkanes, respectively [80,81]. The antisymmetric and symmetric vibrations of C–O–C correspond to the bands at 1312 cm<sup>-1</sup> and 1092 cm<sup>-1</sup>, whereas the band at 1230 cm<sup>-1</sup> is due to O–H vibration coupled with C–O stretching [82]. The peak at 642 cm<sup>-1</sup> is attributed to C–Cl stretching [83]. The TiO<sub>2</sub> MI/2,4D Not Removed exhibited some of the typical bands of 2,4D which indicate the successful attachment of the molecule target (2,4D) to the titanium dioxide. Specifically, as reported in Table 2, the

bands at  $1736\text{ cm}^{-1}$ ,  $1312\text{ cm}^{-1}$ , and  $1092\text{ cm}^{-1}$  of 2,4D are shifted to different wavenumbers on the  $\text{TiO}_2$  MI/2,4D Not Removed. These shifts can be correlated to the formation of hydrogen bonding and to the electrostatic interactions originated during the first steps of the sol-gel synthesis, i.e. the hydrolysis and the successively condensation of titanium butoxide [49]. Indeed, the first step of sol-gel synthesis involved the hydrolysis of titanium butoxide with the consequent condensation to form the Ti-O-Ti network. Meanwhile, the formation of the complex between  $\text{TiO}_2$  and the pesticide (added together to titanium butoxide) occurs by electrostatic interaction and hydrogen bonding. The calcination treatment leads to the complete removal of the pesticide and the consequently formation of the molecularly imprinted sample, as confirmed by FTIR spectrum of the  $\text{TiO}_2$  MI/2,4D sample, in which the bands related to the 2,4D are absent. Indeed, the spectrum of the  $\text{TiO}_2$  MI/2,4D catalyst is almost similar to that of bare  $\text{TiO}_2$ . The wide band in the range  $700\text{-}500\text{ cm}^{-1}$  observed for the  $\text{TiO}_2$  MI/2,4D and  $\text{TiO}_2$  samples can be assigned to the Ti-O-Ti stretching vibration modes [82]. The band at  $1628\text{ cm}^{-1}$  of un-modified  $\text{TiO}_2$  is assigned to the bending vibration of the O-H group of residual water molecules.

The FTIR spectra of imidacloprid,  $\text{TiO}_2$  MI/Imid Not Removed (i.e., before the calcination),  $\text{TiO}_2$  MI/Imid, and  $\text{TiO}_2$  are shown in Fig. 6B. The bands at  $3363\text{ cm}^{-1}$  and  $1288\text{ cm}^{-1}$  of imidacloprid are assigned to the stretching of N-H and N-C of secondary ammine, respectively [83]. The peak at  $1569\text{ cm}^{-1}$  characterizes the stretching vibration of pyridine ring, whereas the band at  $1445\text{ cm}^{-1}$  are attributed to the C=N ring in-plane vibrations and to the deformation modes of =CH- on the pyridine ring [84]. The symmetric vibration of  $\text{NO}_2$  group can be identified with the band at  $1230\text{ cm}^{-1}$ , whereas the asymmetric vibrations that are usually associated to the bands in the  $1530\text{-}1630\text{ cm}^{-1}$  region overlap with the other bands related to the stretching vibration of pyridine ring

(reported in the 1500-1300  $\text{cm}^{-1}$  region) [85]. Finally, the band at 1051  $\text{cm}^{-1}$  is assigned to pyridine C–Cl stretching [82]. Also in this case the  $\text{TiO}_2$  Imid Not Removed sample (i.e., without calcination treatment) showed some fingerprint bands of the pesticide used as template, whereas as expected after the calcination ( $\text{TiO}_2$  MI/Imid sample) the peaks due to the presence of imidacloprid disappeared. Table 3 indicates a shift from 1569  $\text{cm}^{-1}$  for imidacloprid to 1539  $\text{cm}^{-1}$  for  $\text{TiO}_2$  MI/Imid Not Removed, due to the electrostatic interactions and the hydrogen bonds with the nitrogen of pyridine of imidacloprid, confirming also for these samples the successful attachment of the target molecule (i.e., imidacloprid) to the Ti–O–Ti network in the first steps of sol-gel route. The wide band at 3358  $\text{cm}^{-1}$  of  $\text{TiO}_2$  MI/Imid Not Removed could be due to the overlap of the peaks of the stretching of amine group and the peaks of the stretching of the adsorbed water. The  $\text{TiO}_2$  MI/Imid sample displayed the typical FTIR spectrum of titanium dioxide, unambiguously demonstrating the removal of the template thanks to the calcination process.

In order to have further validations of the successful interaction between the pesticides and the  $\text{TiO}_2$  and to exclude the eventual presence of other elements/impurities on the samples' surface, XPS measurements were carried out (Fig. 7). Stoichiometric  $\text{TiO}_2$  samples are white in colour, while the eventual presence of oxygen deficient  $\text{TiO}_{2-x}$  species causes changes in colouration from white up to deep blue in  $\text{TiO}_2$  ceramic samples [86,87]. All the samples prepared in this study were white. As an initial check control, we performed XPS analysis of bare  $\text{TiO}_2$  anatase powders. XPS results show the Ti  $2p_{3/2, 1/2}$  spin-orbit components at 459.9 and 465.6 eV (Tab. 4, Fig. S3A) and a rather symmetric O 1s band at 531.1 eV (Fig. S3B), in agreement with already reported literature data for the anatase phase [88]. No evidence of low binding energy broadening of the doublet evident for  $\text{TiO}_2$  itself was observed. This fact



suggests the substantial absence of  $\text{TiO}_{2-x}$  species on the XPS probed depth [86, 87]. In fact, the eventual presence of Ti ions in a 3+ valence state should give origin to an XPS spectrum consisting of a pair of overlapping Ti 2p doublets, because of the competition of various final state screening effects [89-91].

The XPS spectrum of the  $\text{TiO}_2$  MI/2,4D Not Removed sample (i.e., before the calcination) shows the Ti 2p  $3/2$  and  $1/2$  spin-orbit components at 459.3 and 465.2 eV (Tab. 4, Fig. S3C), and a rather broad O 1s band centred at 531.7 eV (Tab. 4, Fig. S3D). The Ti 2p levels are at 0.6-0.4 eV lower binding energies with respect to those observed for the not-imprinted  $\text{TiO}_2$ , thus indicating the donation electron capability of the 2,4D molecule. In contrast, the O 1s spectral profile clearly suggests the presence of different electronic states consistent with the presence of  $\text{TiO}_2$ , oxygen atoms belonging to the 2,4D molecule, water and, finally, Ti-OH hydroxide species.

Fig. 7A shows the XPS spectrum of the  $\text{TiO}_2$  MI/2,4D sample (i.e after calcination at 500 °C) in the Ti 2p energy region. The Ti 2p features are at 459.4 eV and 465.2 eV (Tab. 4). These XPS values are almost coincident with those observed for the  $\text{TiO}_2$  MI/2,4D Not Removed sample, thus suggesting that the calcination process does not significantly modify the electronic states of the imprinted system. It seems that after calcination a “memory” effect maintains the overall electronic structure, thus confirming the retention of the structural microstructure of the pesticides molecules. Worthy of note, while the  $\text{TiO}_2$  MI/2,4D Not Removed sample clearly shows (Fig. S4A) the Cl 2p spin-orbit components at 200.2 and 201.7 eV (Tab. 4), (in close agreement with the XPS values observed for the 2,4D molecule), in contrast there is no XPS trace of chlorine (Cl 2p states) after the calcination. These observations demonstrate the absence of the 2,4D molecule (Fig. S4A) in the  $\text{TiO}_2$  MI/2,4D sample. Moreover, Fig. 7B illustrates the highly symmetric O 1s XPS band of the  $\text{TiO}_2$  MI/2,4D catalyst. It is evident that the

calcination procedure removes also both water and Ti-OH hydroxide surface contaminants.

XPS ionization energies, related to the TiO<sub>2</sub> MI/Imid Not Removed (Figs. S4 B-C) and TiO<sub>2</sub> MI/Imid (Figs. 7C-D) samples, are strongly reminiscent of those of the TiO<sub>2</sub> MI/2,4D system, being the former ionization just about 0.3 eV at lower binding energies (Tab. 4). This evidence demonstrates the electron donating capability of this further molecule (imidacloprid) and the fact that the calcination procedure does not significantly modify the electronic states of the imprinted samples (see Tab. 4). Furthermore, in the TiO<sub>2</sub> MI/Imid Not Removed sample the XPS N 1s band (Fig. S4D) clearly consists of two major evident features at 399.6 eV with a shoulder (due to the amine and imine moieties) at 405.3 eV (due to the -NO<sub>2</sub> group). In tune with the behaviour already observed for the TiO<sub>2</sub> MI/2,4 D catalyst, also in this case the calcination treatment totally removes the imidacloprid molecule from the TiO<sub>2</sub> matrix, since no XPS N 1s signal can be found in the related spectrum (Fig. S4D).

The results of the atomic concentration calculated by XPS analysis indicate that for all the calcined samples the O/Ti ratios are  $2.2 \pm 0.1$ , in agreement with the expected stoichiometry ratio. In contrast, the two not-treated samples (before calcination) show O/Ti ratios of 2.5 (TiO<sub>2</sub> MI/2,4D Not Removed) and 2.6 (TiO<sub>2</sub> MI/Imid Not Removed). These ratios are in agreement with the presence of oxygen atoms in both the 2,4D and imidacloprid pesticides. The nitrogen atomic concentration observed in the TiO<sub>2</sub> MI/Imid Not Removed catalyst gave a N/Ti ratio of 0.56, thus about a half with respect to the nominal concentration of nitrogen considering the 5 nitrogen of imidacloprid and the 5:1 molar ratio between the TiO<sub>2</sub> and the pesticides. Similar observations can be done for the TiO<sub>2</sub> MI/2,4D Not Removed sample that shows a Cl/Ti ratio of 0.32 close to the nominal ratio once the fact that the 2,4D molecule contains 2 chlorine atoms. On

the contrary carbon is present in all samples. This is expected, since it is well known that atomically pure TiO<sub>2</sub> samples can only be obtained by firing overnight at 1000 °C directly in the XPS main chamber, to avoid any contact with air before the XPS analysis [86,87]. We cannot perform this procedure since firing of TiO<sub>2</sub> at temperatures equal or above 600 °C causes the transformation of the anatase in the rutile phase. Moreover, the adventitious carbon contamination, due to the hydrocarbons present in air, is currently used to calibrate the XPS spectra [68, 69]. As a consequence, all the calcined samples contain a certain amount of carbon in the 5-10 % range. However, in accordance with the procedure reported in the literature [71] the catalysts were irradiated with UV radiation before the photocatalytic tests for 60 min, to remove the hydrocarbons and the adventitious carbon from their surface.

The absence of other impurities after the calcination process on the imprinted samples were further assed by the point of zero charge determination and the UV-vis DRS measurements. In particular, as showed in the Fig. S5 our materials did not exhibit any substantial variation of the point of zero charge due to the imprinting process, being the pzc between  $6.7 \pm 0.3$  and  $6.9 \pm 0.3$ , also comparable to the values reported in the literature for the TiO<sub>2</sub> synthetized by sol-gel method [92, 93]. Similarly, only slight variations of the reflectance were detected in the imprinting samples compared to unmodified TiO<sub>2</sub> (Fig. S6), and the differences of band-gap values (from  $3.0 \pm 0.3$  to  $3.1 \pm 0.3$  eV) estimated by the Kubelka-Munk method [94] are included in the experimental error, suggesting that the molecular imprinting process did not modify the band-gap structure of TiO<sub>2</sub>.

To test the photocatalytic efficiency of the synthetized samples, the degradation of 2,4D (Fig. 8A) and of imidacloprid (Fig.8B) pesticides under UV light irradiation was measured. The graphs report  $C/C_0$  ratio as a function of time  $t$ , where  $C$  is the

concentration of the pollutant after UV light irradiation at the time  $t$ , while  $C_0$  is the starting concentration of the pesticide (the variations of absorbance with the time are displayed in Fig. S7). The experimental error of the photodegradation measurements is 1%. Before the catalytic measurements, control experiments without irradiation, were performed. The results obtained in the dark did not indicate any relevant contribution of the adsorption process; this is probably related to the formation of weak interactions between the pesticides and the molecularly imprinted samples in the aqueous solution. Similar results, concerning the lack of contribution of the adsorption process, are reported in the literature for molecularly imprinted ZnO and TiO<sub>2</sub> [36, 48].

Fig. 8A is focused on the degradation of the 2,4D herbicide. In absence of any catalysts there is not a detectable degradation of 2,4D, as expected. Differently, bare TiO<sub>2</sub> sample leads to a degradation of the 2,4D pesticide of  $\sim 15\%$ . A significant enhancement of the 2,4D conversion, till to  $\sim 47\%$ , was obtained through the TiO<sub>2</sub> MI/2,4D catalyst. Thus, the combination of molecular imprinting with the photocatalytic process allowed to efficiently promote the 2,4D degradation compared to bare TiO<sub>2</sub> by a factor of  $\sim 3$ . A similar catalytic behaviour can be noted between the TiO<sub>2</sub> MI/Imid, and the un-modified TiO<sub>2</sub>, that is an indication of an achieved selectivity in the photocatalytic process. Indeed, only the sample imprinted with the same pollutant that should be degraded (in this case the 2,4D pesticide that was imprinted on the TiO<sub>2</sub> MI/2,4D catalyst) can efficiently remove the 2,4D herbicide from the contaminated water. The calculated kinetic rate constants for all the investigated samples (considering a first order kinetic) [95,96] are reported in Tab. 5. The kinetic constant of TiO<sub>2</sub> MI/2,4D is  $\sim 6$  times higher compared to the not-imprinted TiO<sub>2</sub> samples; this higher degradation rate of TiO<sub>2</sub> MI/2,4D is an evidence of the synergism between the molecular imprinting and the photocatalytic process: the former leads to selectively

catch (through the molecular imprinting process) the pesticide, the latter to effectively degrade it from water (through the photocatalysis).

It is important to remark that the high performance of the TiO<sub>2</sub> MI/2,4D catalyst cannot be related to any variation of the BET surface area. Indeed, the data reported in Tab. 1 indicate a similar surface area between bare TiO<sub>2</sub> ( $59 \pm 1 \text{ m}^2 \text{ g}^{-1}$ ) and molecularly imprinted materials: TiO<sub>2</sub> MI/2,4D ( $55 \pm 1 \text{ m}^2/\text{g}$ ) and TiO<sub>2</sub> MI/Imid ( $53 \pm 1 \text{ m}^2/\text{g}$ ). Consequently, the key factor in the preferential photodegradation of the target pollutant is due to the molecular imprinting process.

The high performance through the photocatalytic oxidation of 2,4D pesticide showed by the TiO<sub>2</sub> MI/2,4D catalyst is promising to effectively remove this harmful compound from water. Underlining that the photo-oxidation can be considered an alternative solution together with adsorption [97, 98], the selective separation and sensing [99-101] obtained with other molecularly imprinted 2,4D materials.

The selectivity of the process was further confirmed considering the photoactivity of TiO<sub>2</sub> MI/2,4D towards the degradation of another pesticide: the imidacloprid insecticide. The result is reported in Fig. 8B. A comparable photoactivity was measured between the TiO<sub>2</sub> MI/2,4D and the un-modified TiO<sub>2</sub>, highlighting the selectivity of the process. The TiO<sub>2</sub> MI/2,4D sample, in fact, due to the different pores/cavities and the dissimilar chemical composition of the 2,4D respect to imidacloprid, it is not selective for the imidacloprid degradation, showing the same catalytic activity of not-imprinted TiO<sub>2</sub>.

Fig. 8B reports the degradation of the imidacloprid insecticide. Also for the photodegradation of the imidacloprid the molecular imprinting process allows to achieve a selective removal of this pesticide from water (the variations of absorbance with the time are showed in Fig. S8). The concentration of imidacloprid remains substantially the

same in the test without catalyst, while the TiO<sub>2</sub> degrades ~ 19% of the initial concentration of the imidacloprid pesticide. The TiO<sub>2</sub> MI/Imid showed the best activity, with a degradation of ~ 35%. **Tab. 6** indicates a 2 times higher kinetic constant for TiO<sub>2</sub> MI/Imid compared to bare TiO<sub>2</sub>. Only the TiO<sub>2</sub> MI/Imid catalyst (sample imprinted with the same pesticide/target to be degraded) allows to significantly increase the photocatalytic degradation of imidacloprid, confirming the attained selectivity of the process and the synergism between the molecular imprinting and the photocatalysis. Also in this case, the high performance of the TiO<sub>2</sub> MI/Imid catalyst cannot be related to any variation of the BET surface area (see **Tab. 1**). The modification, with the molecular imprinting process of the pores/cavities of TiO<sub>2</sub>, is the key factor to obtain selective and performing materials. The modified pores/cavities are, indeed, the recognition sites for the selective interaction between the TiO<sub>2</sub> and the pesticides that favour an increased photoactivity.

In order to evaluate the ability in removing organic pollutants from the solution, we have measured the TOC amount of the pesticides solutions after 3 hours under UV irradiation. **Figs. 8C and D** suggested a partial mineralization of the solutions. These data are in good agreement with the photocatalytic mechanisms reported in the literature for the degradation of 2,4D and imidacloprid [95,96,102-104]. In particular, for the 2,4D pesticide, the hydroxyl radicals ( $\bullet$ OH) (formed after the UV irradiation of TiO<sub>2</sub>) react with the herbicide to yield mainly 2,4-Dichlorophenol and 2-chloro benzoquinone. This was confirmed by ESI-MS measurement reported in **Fig. S9**.

Regarding the TiO<sub>2</sub>-driven imidacloprid photodegradation, it is expected that the interaction of the hydroxyl radicals with the imidacloprid gives the cleavage of C–N and the N–N bonds with the formation of amine and chloro-pyridine species [96,104].

Further oxidation processes lead to the formation of small molecules as H<sub>2</sub>O, CO<sub>2</sub>, chlorine dioxide and nitrogen oxides species [96,104].

The synthesized TiO<sub>2</sub>-based photocatalysts exhibited a good reusability property (Fig. 9). Indeed, both the 2,4D photodegradation kinetic constant (Fig.9A) and the imidacloprid photodegradation kinetic constant (Fig.9B) remain essentially the same after five consecutive photocatalytic tests. The slight decrease of the kinetic constants in the last runs can be reasonably ascribed to a small loss of the powder during the centrifugation processes. Furthermore, no substantial variations were detected from the XPS and the FTIR analyses of the used (i.e after the photocatalytic tests) TiO<sub>2</sub> MI/2,4D and TiO<sub>2</sub> MI/Imid. (Figs. S10-S11) materials, confirming the stability of the investigated samples.

An additional photocatalytic test was performed, employing a mixed aqueous solution of 2,4D and imidacloprid. The results of this test are shown in Fig. 10 (the absorption spectra of the catalysts as a function of the time are reported in Fig. S12) Fig. 10A demonstrates that also in this case only the TiO<sub>2</sub> MI/2,4D leads to a higher performance in the photo-oxidation of the herbicide 2,4D. In detail, the kinetic constant of the 2,4D degradation is about 5 times higher compared to TiO<sub>2</sub> and TiO<sub>2</sub> MI/Imid (Tab. 7). Analogously, following the imidacloprid degradation in the mixed solution (Fig. 10B) it is clear that the TiO<sub>2</sub> MI/2,4D and TiO<sub>2</sub> exhibited similar photoactivity, whereas the TiO<sub>2</sub> MI/Imid effectively degrades the corresponding pesticide/template with a kinetic constant which is about 2 times higher compared to the ones of the other TiO<sub>2</sub> photocatalysts.

We performed a photocatalytic test with another pesticide: the fungicide ortho-phenylphenol. The molecular structure of the fungicide is reported in Fig. 11A. Fig. 11B shows that the imprinted samples displayed a similar catalytic behaviour of bare TiO<sub>2</sub>

with comparable kinetic constants (Tab. 8). Indeed, being the molecular structure of the fungicide (Fig. 11A) completely different from the ones of 2,4D and imidacloprid (Fig. 1), no particular interactions between the imprinted samples and the pesticide are favoured, leading to a degradation rate similar to bare TiO<sub>2</sub>.

Finally, to have further proofs of the selectivity of the synthesized TiO<sub>2</sub> molecularly imprinted samples, we carried out a photocatalytic test using the 1,4-dichlorobenzene pollutant, that has a similar chemical structure of 2,4D pesticide (Fig. 12A). It is possible to notice (Fig. 12B) that, also in this case, all the investigated catalysts showed similar performance, with a slightly better degradation efficiency displayed by the TiO<sub>2</sub> MI/2,4D (42%) compared to TiO<sub>2</sub> (37%) and TiO<sub>2</sub> MI/Imid (35%). The small increase of the photoactivity of the TiO<sub>2</sub> MI/2,4D can be due to the weak interactions occurring between this photocatalyst and the 1,4-dichlorobenzene caused by the chemical similarity between 1,4-dichlorobenzene and 2,4D. While the TiO<sub>2</sub> MI/Imid, owing completely different recognition sites/pores, exhibited the same catalytic behaviour of the un-imprinted TiO<sub>2</sub>. As clearly visible from the Fig. 12B a small photodegradation due to the photolysis (i.e. degradation without catalysts), under UV irradiation, characterized the 1,4-dichlorobenzene, this is in accordance with the literature [105].

The reported photocatalytic tests indicate that the molecular imprinting process is as an effective methodology to achieve a selective degradation of pesticides from water by titanium dioxide. The samples synthesized using as templates the pesticides showed a remarkable catalytic activity towards the corresponding pesticide (i.e. TiO<sub>2</sub> MI/2,4D towards 2,4D; TiO<sub>2</sub> MI/Imid towards imidacloprid). This is due to the electrostatic interactions and hydrogen bonds formed during the sol-gel synthesis between the Ti–O–Ti network and the pesticide molecules (as confirmed by FTIR analyses reported in Fig. 6). After calcination, the pesticide is completely removed (as confirmed by FTIR



analyses depicted in Fig. 6, and confirmed by XPS results reported in Fig. 7) from the TiO<sub>2</sub>, favouring in this way the formation of a titania with a “memory” of the template molecules. The final result is a specific molecular recognition between the molecularly imprinted TiO<sub>2</sub> and the pesticide used during the synthesis step. This allows a selective and effective photo-degradation of the water contaminant used as a template.

In view of real applications, it is crucial to examine the eventual toxicity of the investigated materials. For this reason, we have tested the micro/nanopowders toxicity using the zebrafish as model organism (see details in the supporting information Fig. S13). The obtained positive results lead to exclude, with the investigated experimental conditions, a critical toxicity of the examined materials.

#### 4. Conclusions

Molecularly imprinted TiO<sub>2</sub> photocatalysts were synthesized using an easy and simple sol-gel methodology. The herbicide 2,4-D or the insecticide imidacloprid, two common pesticides in agriculture, were used as templates during the synthesis process, and then removed through a calcination step. The effectiveness of the molecular imprinting processes was confirmed by FTIR and XPS measurements, that highlighted the formation of electrostatic interaction and/or hydrogen bonds between the Ti–O–Ti network and the pesticide molecules in the first steps of the sol-gel synthesis. This approach induced an increased and selective photodegradation of the water contaminant used as a template, i.e. TiO<sub>2</sub> MI/2,4D sample towards the degradation of 2,4D, and TiO<sub>2</sub> MI/Imid sample towards the degradation of imidacloprid. Finally, the eventual toxicity of the synthesized TiO<sub>2</sub> powders was excluded thanks to the positive responses of the

zebrafish embryotoxicity tests. The approach described is a promising strategy to obtain safe and performing materials for environmental applications.

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**Table 1** BET Surface area ( $S_{\text{BET}}$ ), mean pore diameter ( $d_p$ ) and pore volume ( $V_p$ ) of the investigated samples.

Catalysts	$S_{\text{BET}}$ ( $\text{m}^2 \text{g}^{-1}$ )	$d_p$ (nm)	$V_p$ ( $\text{cm}^3 \text{g}^{-1}$ )
TiO <sub>2</sub>	59 ± 1	5.1 ± 0.2	0.05 ± 0.02
TiO <sub>2</sub> MI/2,4D	55 ± 1	6.4 ± 0.2	0.14 ± 0.02
TiO <sub>2</sub> MI/Imid.	53 ± 1	9.7 ± 0.2	0.19 ± 0.02

**Table 2** Vibrational frequencies of 2,4D and TiO<sub>2</sub> MI/2,4D Not Removed samples.

	2,4D	TiO <sub>2</sub> MI/2,4D Not Removed
v C=O	1736 $\text{cm}^{-1}$	1719 $\text{cm}^{-1}$
v C=C <sub>aromatic</sub>	1478 $\text{cm}^{-1}$	1476 $\text{cm}^{-1}$
v C-H <sub>alkanes</sub>	1431 $\text{cm}^{-1}$	1424 $\text{cm}^{-1}$
v C-O-C <sub>as</sub>	1312 $\text{cm}^{-1}$	1290 $\text{cm}^{-1}$
v O-H/C-O	1230 $\text{cm}^{-1}$	1230 $\text{cm}^{-1}$
v C-O-C <sub>s</sub>	1092 $\text{cm}^{-1}$	1081 $\text{cm}^{-1}$
v C-Cl	642 $\text{cm}^{-1}$	642 $\text{cm}^{-1}$

**Table 3** Vibrational frequencies of Imidacloprid and TiO<sub>2</sub> MI/Imid Not Removed samples.

	Imidacloprid	TiO <sub>2</sub> MI/Imid Not Removed
v pyridine	1569 $\text{cm}^{-1}$	1539 $\text{cm}^{-1}$
v C=N/C-H <sub>pyridine</sub>	1445 $\text{cm}^{-1}$	1443 $\text{cm}^{-1}$
v N-C <sub>secondary amine</sub>	1288 $\text{cm}^{-1}$	1286 $\text{cm}^{-1}$
v NO <sub>2</sub> s	1230 $\text{cm}^{-1}$	1230 $\text{cm}^{-1}$
v C-Cl <sub>pyridine</sub>	1051 $\text{cm}^{-1}$	1051 $\text{cm}^{-1}$

**Table 4** XPS Binding Energies (eV) for the investigated TiO<sub>2</sub> samples.

Sample	C 1s	O 1s	Ti 2p <sub>3/2,1/2</sub>	Cl 2p <sub>3/2,1/2</sub>	N 1s
TiO <sub>2</sub>	258.0	531.1	459.9 465.6	-	-
TiO <sub>2</sub> MI/2,4D Not Removed	285.0 289.2	531.7	459.3 465.2	200.3 201.9	-
TiO <sub>2</sub> MI/2,4D	285.0 289.1	530.8	459.4 465.2	-	-
TiO <sub>2</sub> MI/Imid. Not Removed	285.0 288.9	531.4	459.1 464.9	200.2 201.8	399.6 405.3
TiO <sub>2</sub> MI/Imid.	285.0 289.1	530.6	459.1 464.9	-	-

**Table 5** Kinetic constants of the photocatalytic degradations of 2,4D.

Sample	k (min <sup>-1</sup> ) · 10 <sup>-4</sup>
2,4D	1.01 ± 0.06
TiO <sub>2</sub>	6.09 ± 0.06
TiO <sub>2</sub> MI/2,4D	36.15 ± 0.06
TiO <sub>2</sub> MI/Imid	6.08 ± 0.06

**Table 6** Kinetic constants of the photocatalytic degradations of imidacloprid.

Sample	k (min <sup>-1</sup> ) · 10 <sup>-4</sup>
Imidacloprid	1.00 ± 0.06
TiO <sub>2</sub>	11.05 ± 0.06
TiO <sub>2</sub> MI/2,4D	11.09 ± 0.06
TiO <sub>2</sub> MI/Imid	22.45 ± 0.06

**Table 7** Kinetic constants of the photocatalytic degradations of 2,4D and imidacloprid in the mixed solution (2,4D + imidacloprid).

<b>Sample</b>	<b>k 2,4D (min<sup>-1</sup>) · 10<sup>-4</sup></b>	<b>k imid. (min<sup>-1</sup>) 10<sup>-4</sup></b>
2,4D + imid.	1.01 ± 0.06	0.99 ± 0.06
TiO <sub>2</sub>	7.03 ± 0.06	11.07 ± 0.06
TiO <sub>2</sub> MI/2,4D	39.05 ± 0.06	12.03 ± 0.06
TiO <sub>2</sub> MI/Imid	8.06 ± 0.06	20.09 ± 0.06

**Table 8** Kinetic constants of the photocatalytic degradations of o-phenylphenol pesticide.

<b>Sample</b>	<b>k o-phenylphenol (min<sup>-1</sup>) · 10<sup>-4</sup></b>
O-phenylphenol	0.98 ± 0.06
TiO <sub>2</sub>	7.85 ± 0.06
TiO <sub>2</sub> MI/2,4D	8.71 ± 0.06
TiO <sub>2</sub> MI/Imid	9.81 ± 0.06

**Captions to figures**

- Fig. 1** Molecular structures of 2,4D and imidacloprid.
- Fig. 2** SEM images of TiO<sub>2</sub>-based materials: (A) TiO<sub>2</sub> together with a high magnification image reported as inset, (B) TiO<sub>2</sub> MI/2,4D, (C) TiO<sub>2</sub> MI/Imid.
- Fig. 3** TEM images of TiO<sub>2</sub> (A) and TiO<sub>2</sub> MI/2,4D (B). The diffraction patterns of the samples are reported in the insets.
- Fig. 4** XRD patterns of TiO<sub>2</sub>, TiO<sub>2</sub> MI/2,4D, and TiO<sub>2</sub> MI/Imid samples (from the bottom to the top).
- Fig. 5** Pore size distribution of TiO<sub>2</sub>, TiO<sub>2</sub> MI/2,4D, and TiO<sub>2</sub> MI/Imid samples (from the bottom to the top).
- Fig. 6** FTIR spectra of (A) 2,4D, TiO<sub>2</sub> MI/2,4D Not Removed, TiO<sub>2</sub> MI/2,4D and TiO<sub>2</sub> (from the bottom to the top) and of (B) imidacloprid TiO<sub>2</sub> MI/Imid. Not Removed, TiO<sub>2</sub> MI/Imid and TiO<sub>2</sub>. The dashed lines are the main bands of 2,4D and imidacloprid molecules.
- Fig. 7** (A) Al K $\alpha$  excited XPS of the TiO<sub>2</sub> MI/2,4D sample measured in the Ti 2p binding energy region, (B) Al K $\alpha$  excited XPS of the TiO<sub>2</sub> MI/2,4D sample measured in the O 1s binding energy region, (C) Al K $\alpha$  excited XPS of the TiO<sub>2</sub> MI/Imid sample measured in the Ti 2p binding energy region, (D) Al K $\alpha$  excited XPS of TiO<sub>2</sub> MI/Imid sample, measured in the O 1s binding energy region. In all the spectra the structure due to satellite radiation has been subtracted.
- Fig. 8** (A) Photocatalytic degradation of 2,4D under UV light irradiation for 2,4 D (down triangles), TiO<sub>2</sub> (triangles), TiO<sub>2</sub> MI/Imid. (circles) and TiO<sub>2</sub> MI/2,4D

(squares). (B) Photocatalytic degradation of imidacloprid under UV light irradiation for imidacloprid (down triangles), TiO<sub>2</sub> (triangles), TiO<sub>2</sub> MI/Imid (circles) and TiO<sub>2</sub> MI/2,4D (squares). The experimental errors are included in the symbols. (C) Comparison between the measured TOC and UV degradation of 2,4D after 3 h of irradiation for TiO<sub>2</sub>, TiO<sub>2</sub> MI/ Imid, and TiO<sub>2</sub> MI/2,4D samples. (D) Comparison between the measured TOC and UV degradation of imidacloprid after 3 h of irradiation for TiO<sub>2</sub>, TiO<sub>2</sub> MI/ Imid, and TiO<sub>2</sub> MI/2,4D sample.

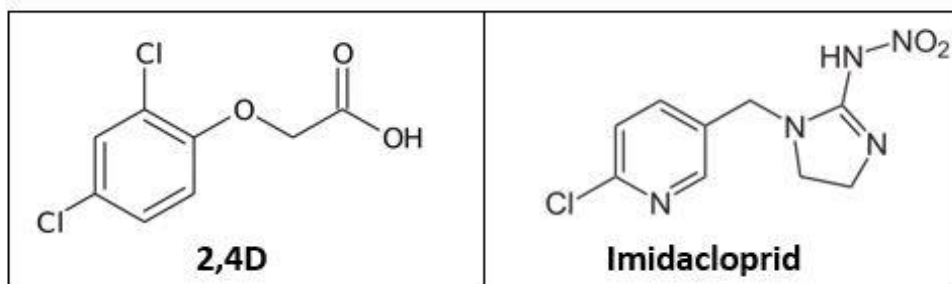
**Fig. 9** (A) 2,4D photodegradation kinetic constants for TiO<sub>2</sub>, TiO<sub>2</sub> MI/Imid and TiO<sub>2</sub> MI/2,4D samples for 5 runs; (B) Imidacloprid photodegradation kinetic constants for TiO<sub>2</sub>, TiO<sub>2</sub> MI/Imid and TiO<sub>2</sub> MI/2,4D samples for 5 runs.

**Fig. 10** (A) Photocatalytic degradation of 2,4D in the mixed 2,4D + imidacloprid solution under UV light irradiation, for 2,4 D (down triangles), TiO<sub>2</sub> (triangles), TiO<sub>2</sub> MI/Imid. (circles) and TiO<sub>2</sub> MI/2,4D (squares). (B) Photocatalytic degradation of imidacloprid in the mixed 2,4D + imidacloprid solution under UV light irradiation, for imidacloprid (down triangles), TiO<sub>2</sub> (triangle), TiO<sub>2</sub> MI/Imid (circle), and TiO<sub>2</sub> MI/2,4D (square). The experimental errors are within the symbols.

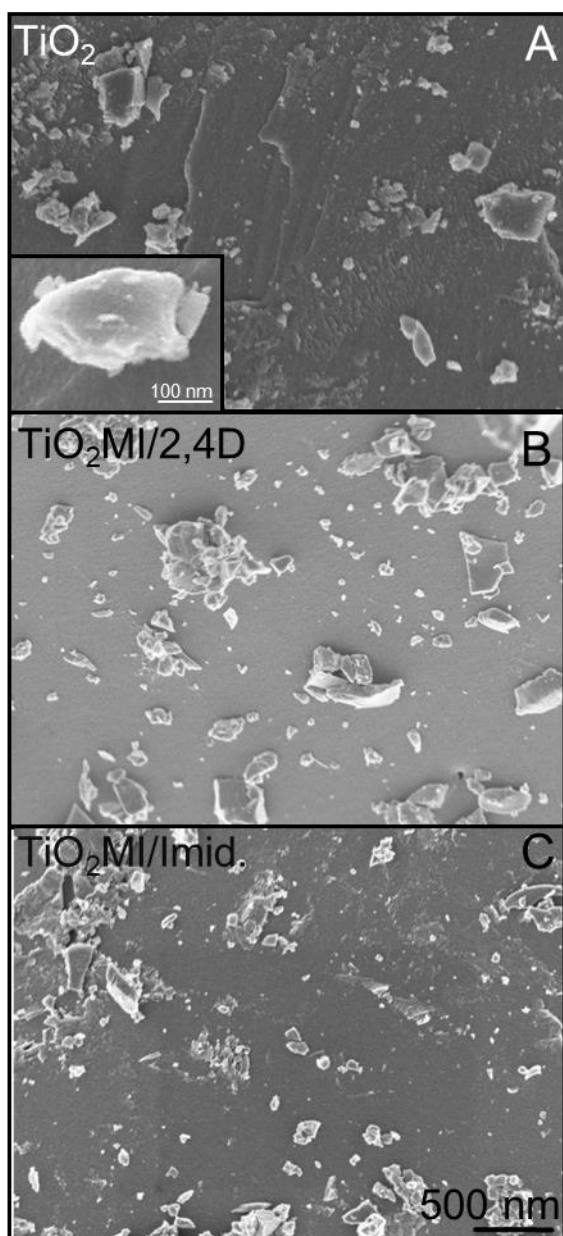
**Fig. 11** (A) Molecular structure of ortho-phenylphenol, (B) Photocatalytic degradation of ortho-phenylphenol under UV light irradiation for o-phenylphenol (down triangles), TiO<sub>2</sub> (triangles), TiO<sub>2</sub> MI/Imid. (circles), and TiO<sub>2</sub> MI/2,4D (squares). The experimental errors are within the symbols.

**Fig. 12** (A) Molecular structure of 1,4-dichlorobenzene, (B) Photocatalytic degradation of 1,4-dichlorobenzene under UV light irradiation for 1,4-dichlorobenzene (down triangles), TiO<sub>2</sub> (triangles), TiO<sub>2</sub> MI/Imid. (circles),

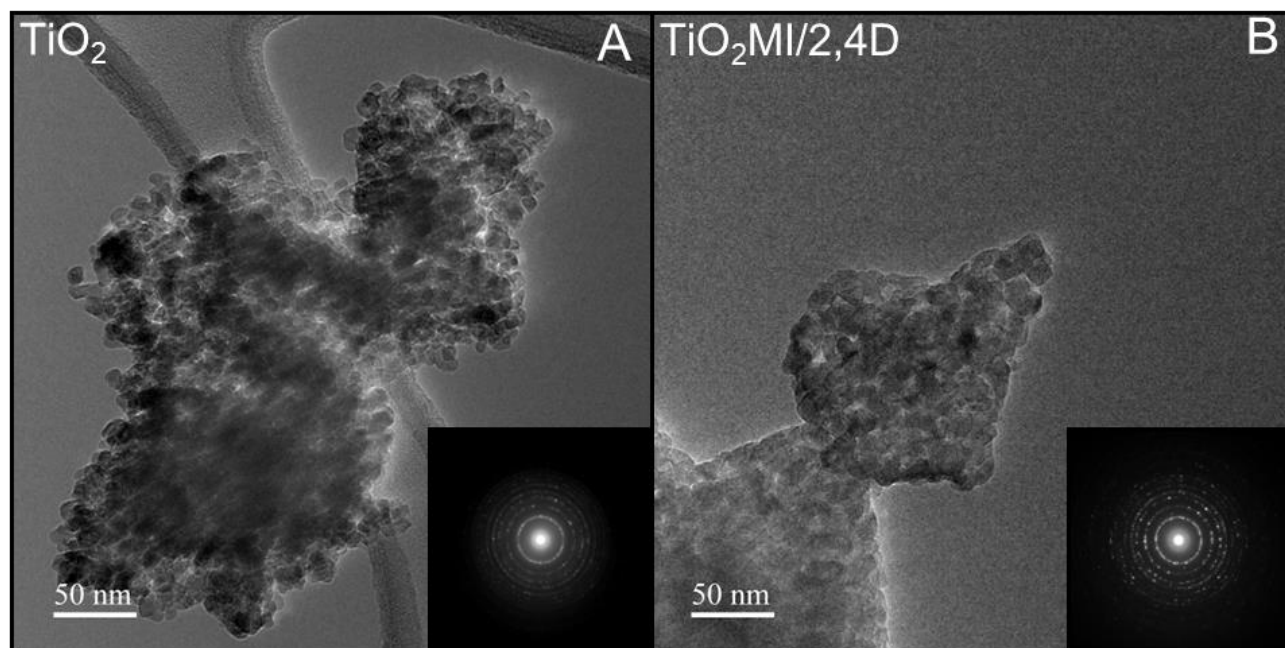
and TiO<sub>2</sub> MI/2,4D (squares). The experimental errors are within the symbols.

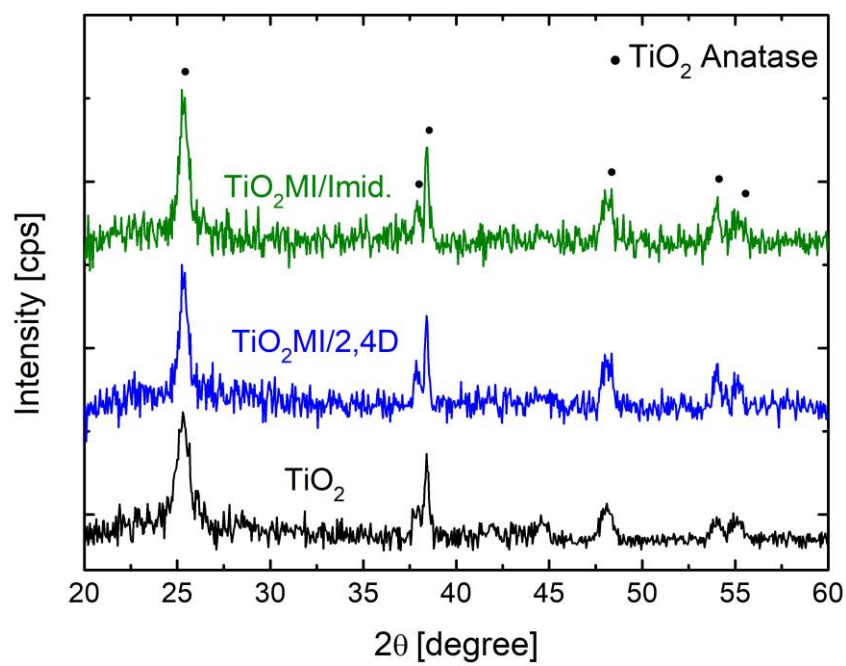


**Fig.1**

**Fig. 2**



**Fig. 3**

**Fig. 4**

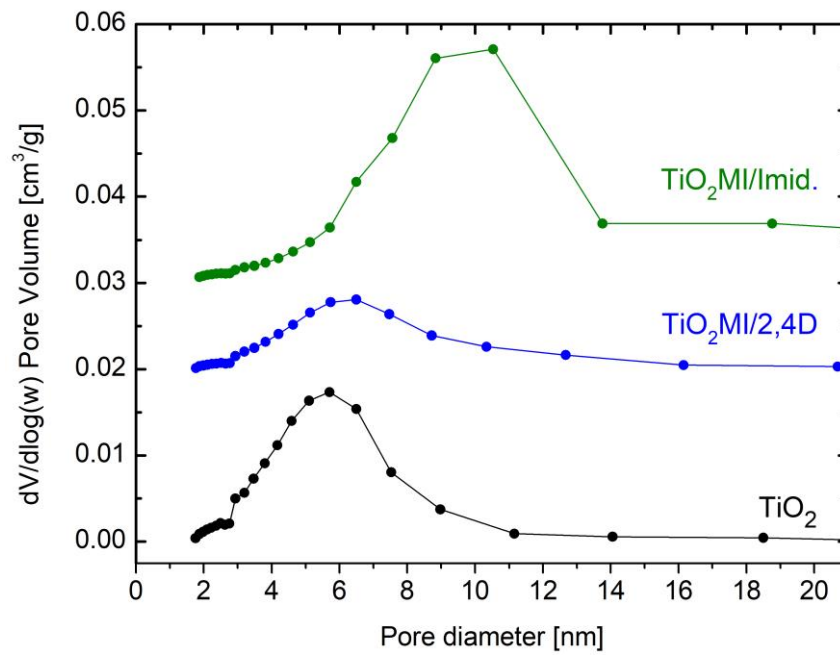
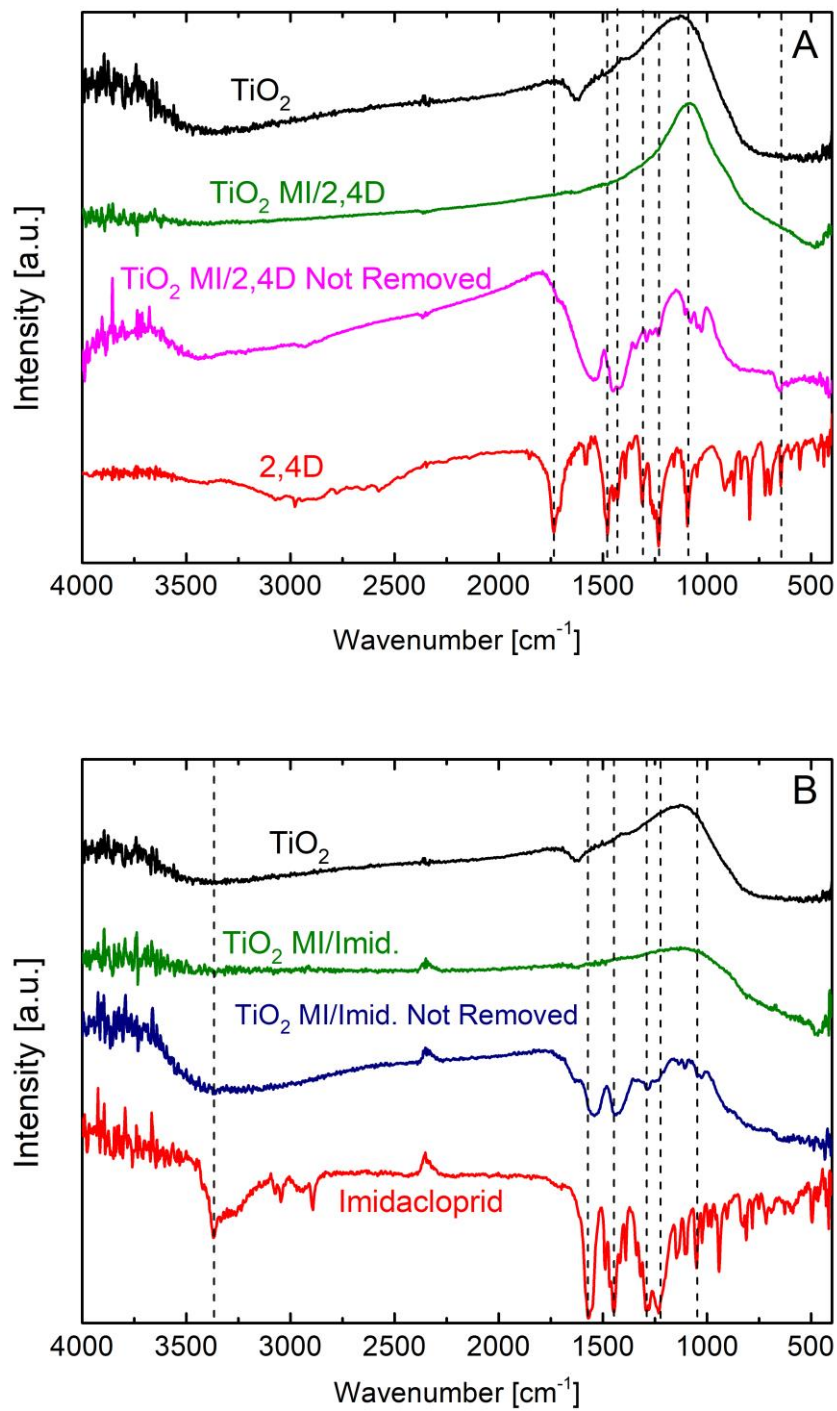
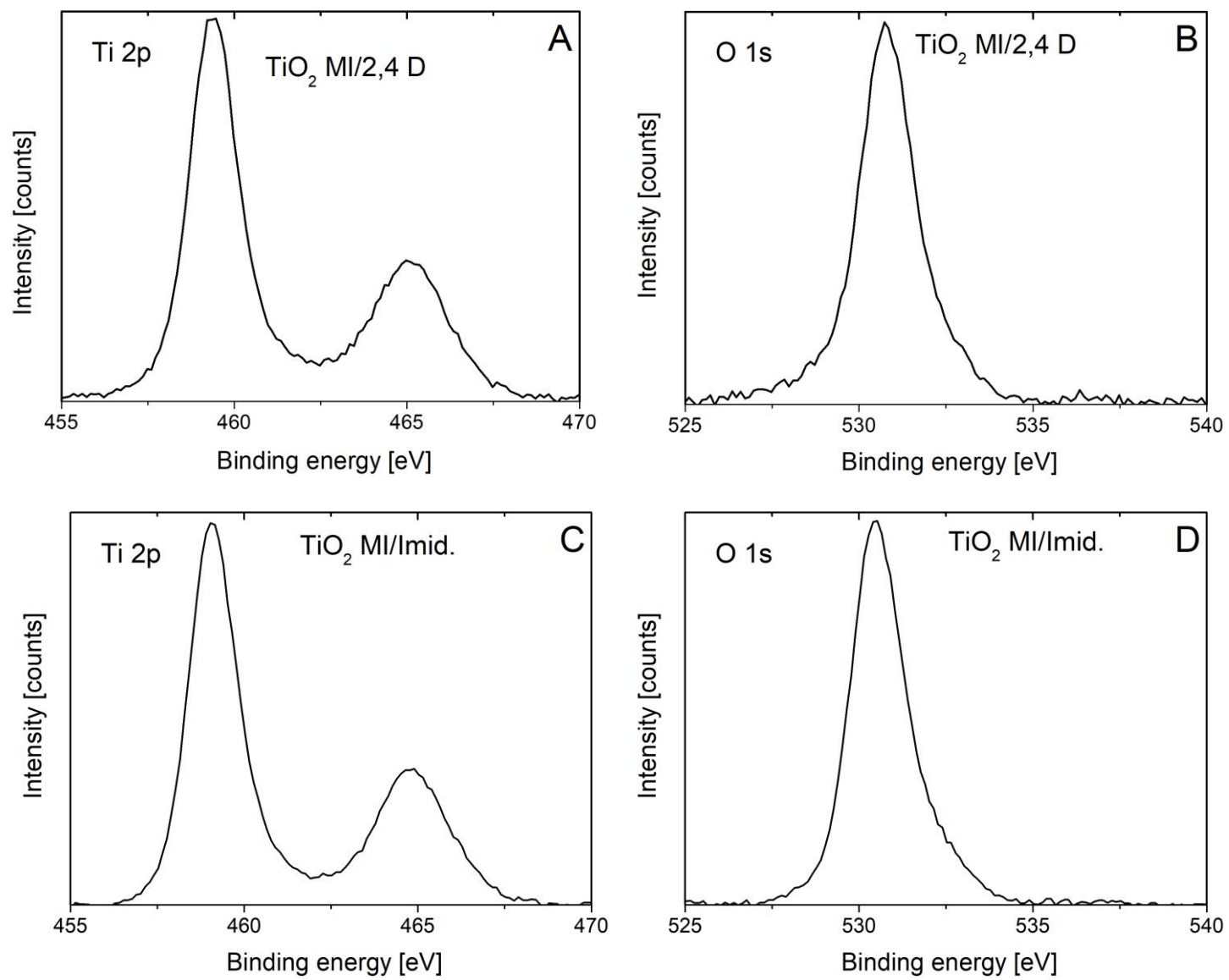


Fig. 5

**Fig. 6**

**Fig. 7**

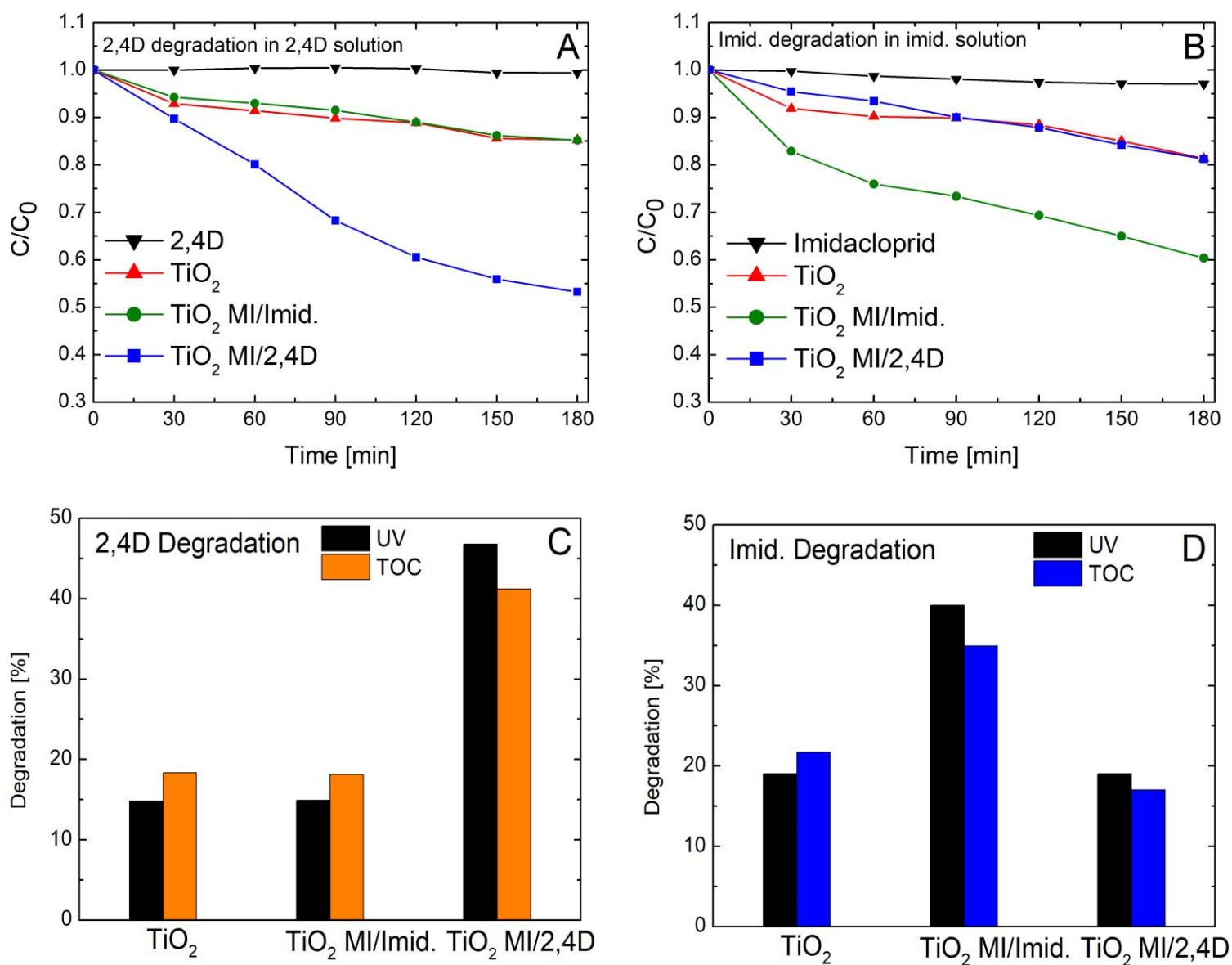


Fig. 8

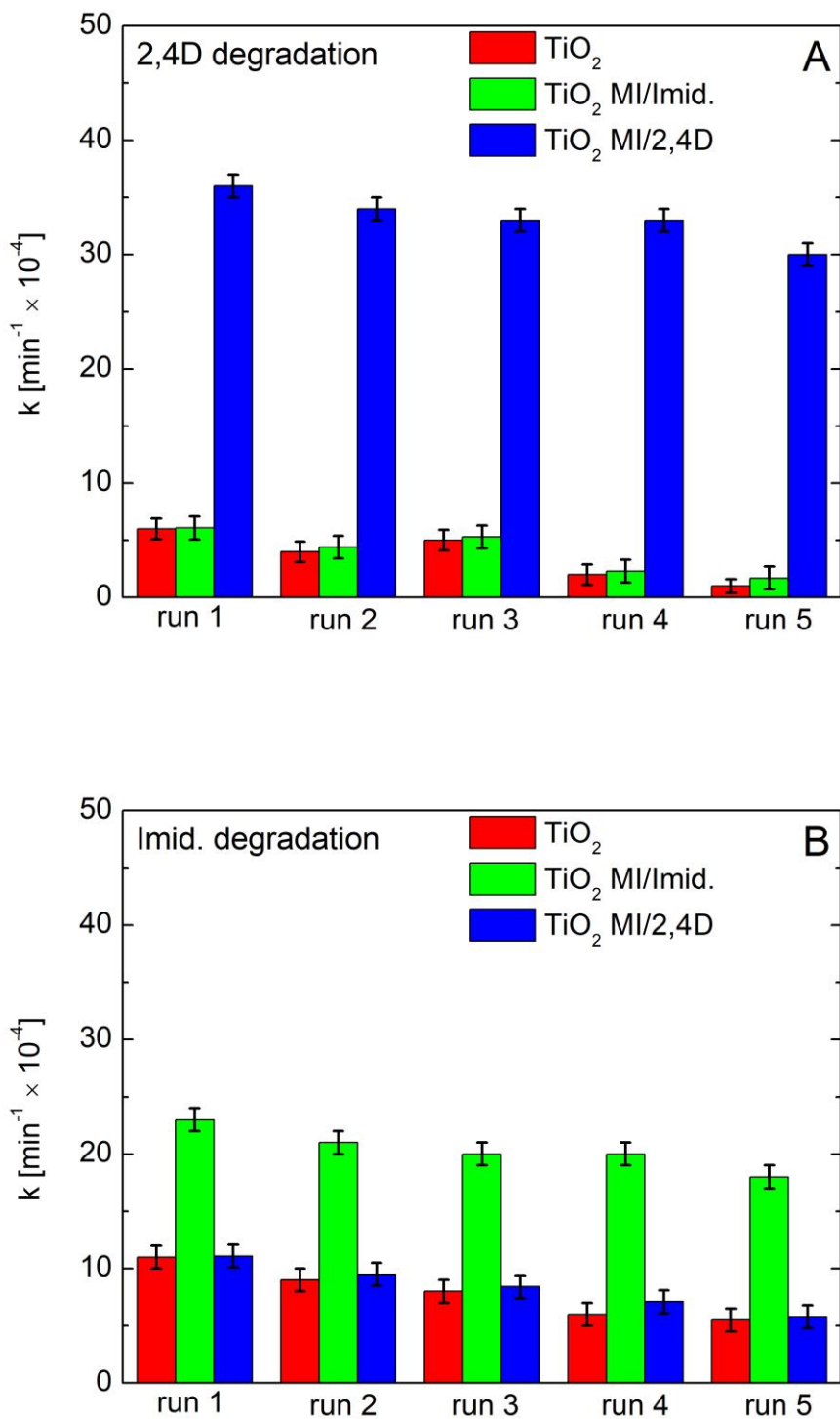


Fig. 9

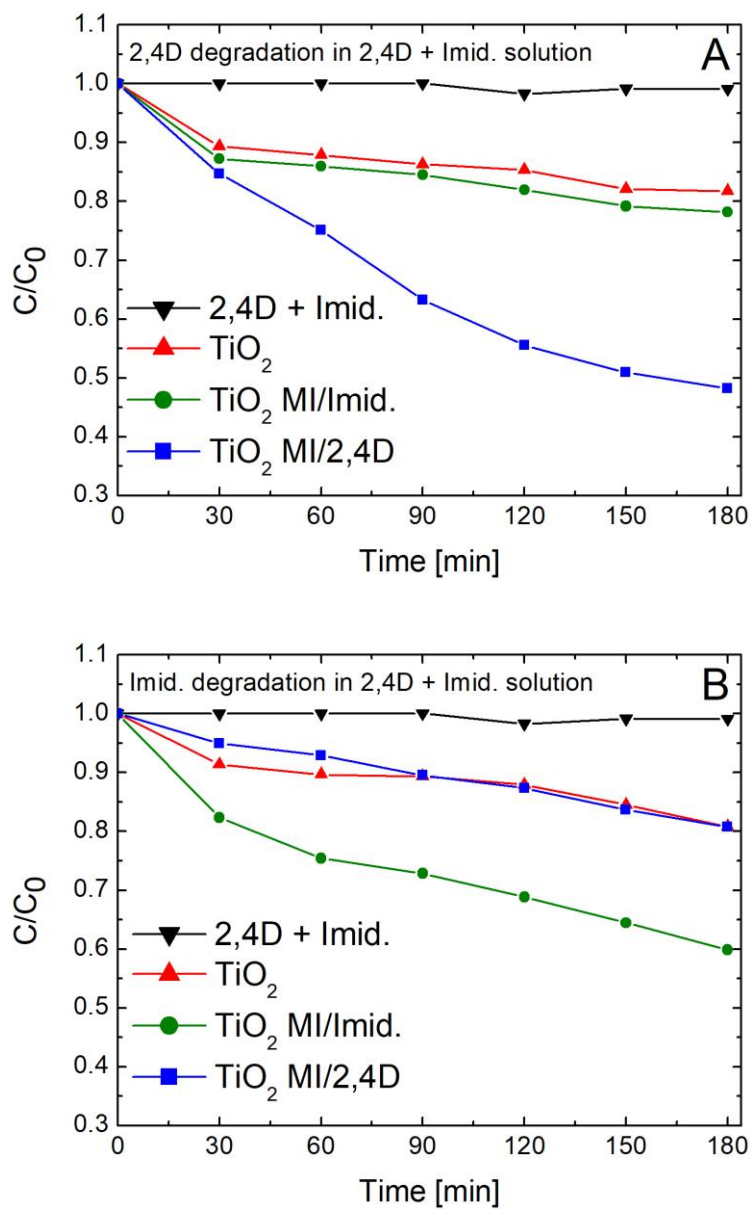
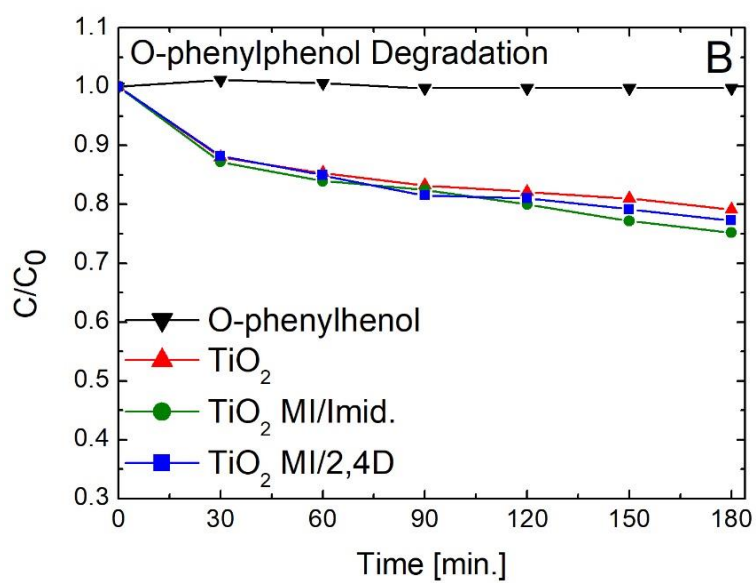
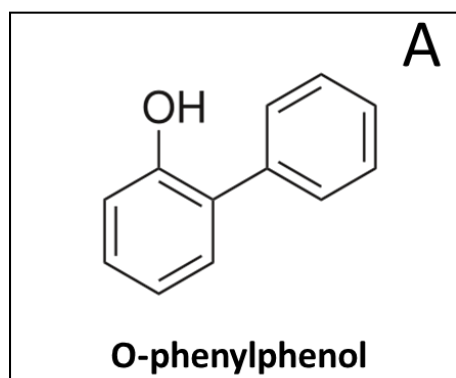
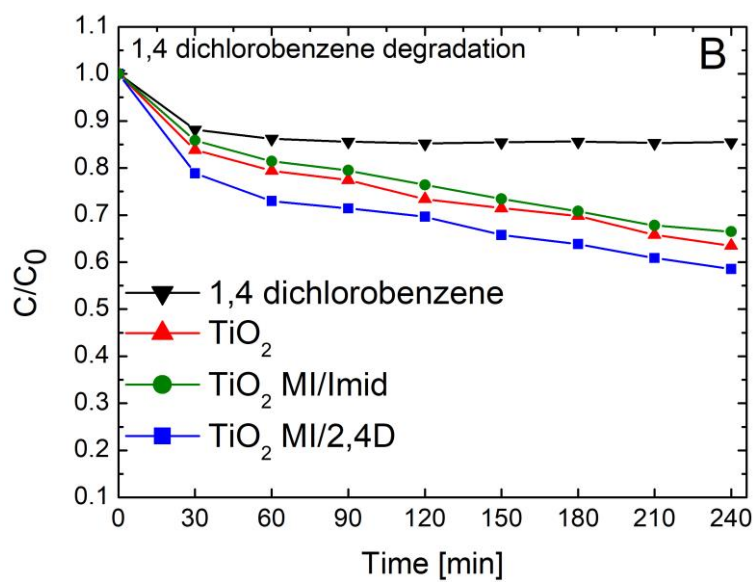
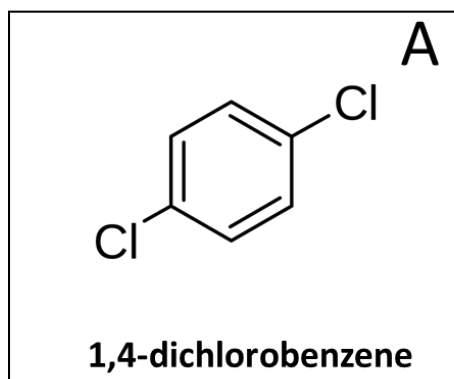


Fig. 10





**Fig. 11**



**Fig. 12**

