

1 **New tricks for old guys:**
2 **recent developments in the chemistry, biochemistry, applications and**
3 **exploitation of selected species from the Lamiaceae family**

4 Edoardo Napoli, Laura Siracusa, Giuseppe Ruberto*

5 *Istituto del CNR di Chimica Biomolecolare, Via Paolo Gaifami, 18*

6 *95126 Catania, Italy*

7 giuseppe.ruberto@icb.cnr.it

8 *In memory of Carmela Spatafora*

9
10
11 **Abstract**

12 Lamiaceae is one of the largest families of flowering plants comprising about 250 genera
13 and over 7,000 species. The majority of the plants of this family are aromatic and
14 therefore important source of essential oils. Lamiaceae are widely used as culinary herbs
15 and reported as medicinal plants in several folk traditions. In the Mediterranean area
16 oregano, sage, rosemary, thyme and lavender stand out for geographical diffusion and
17 variety of uses. The aim of this review is to provide recent data dealing with the
18 phytochemical and pharmacological studies, and the more recent applications of the
19 essential oils and the non-volatile phytocomplexes. This literature survey suggests how
20 the deeper understanding of biomolecular processes in the health and food sectors as per
21 as pest control bioremediation of cultural heritage, or interaction with human
22 microbiome, fields, leads to the rediscovery and new potential applications of well-known
23 plants.

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26 **Keywords:** Lamiaceae; essential oils; extracts; phytocomplexes; biological activity.
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71 **1.Introduction**

72 Lamiaceae family belongs to the Lamiales order, also known as mint order, which
73 comprises 24 families, 1,059 genera and more than 23,800 species. The Lamiaceae is the
74 main family of this order together with Verbenaceae, Plantaginaceae, Scrophulariaceae,
75 Orobanchaceae, Acanthaceae, Gesneriaceae, Bignoniaceae, Oleaceae, Pedaliaceae, and
76 the small carnivorous families Lentibulariaceae and Byblidaceae.

77 Initially this family has been known for long time as Labiatae (*nomen*
78 *conservandum*) or mint family; only in the 1820 the name was definitely changed into
79 Lamiaceae. Without to deepen the taxonomic aspects, which are outside the aims of this
80 review, the Lamiaceae comprises seven sub-families: Ajugoideae, Lamioideae,
81 Nepetoideae, Prostantheroideae, Scutellarioideae, Symphorematoideae and Viticoideae,
82 which in turn enclose several tribes and sub-tribes. On the whole, the genera belonging
83 to this family are 245, which from a first evaluation lists 22,576 species, but only 7,886
84 have been accepted as true species removing a great number of synonym uses or wrong
85 attribution. ^[1] The main genera of this family are *Salvia*, *Scutellaria*, *Stachys*,
86 *Plectranthus*, *Hyptis*, *Teucrium*, *Vitex*, *Thymus* and *Nepeta* (**Figure 1**).

87 **INSERT FIGURE 1**

88 Lamiaceae is the family with the highest number of aromatic plants, even though
89 these are present in many other families, such as Abietaceae, Apiaceae, Asteraceae,
90 Cupressaceae, Luraceae, Myrtaceae, Rutaceae, etc. to quote the most important. For
91 aromatic plants we mean those that by a physical process, namely steam distillation of
92 hydrodistillation, give an essential oil, a lipidic mixture of a high number of different
93 chemical components whose peculiar characteristic is the high volatility and aroma
94 production. Most of the aromatic plants are cultivated and have a growing commercial
95 interest for their fragrance and flavour due to their essential oils, and for their medicinal
96 properties. Despite today more than 3,000 aromatic plants (belonging to the aforesaid
97 families) able to yield an essential oil are known, only three hundred essential oils have a
98 commercial interest. ^[2]

99 Furthermore, Lamiaceae is one of the most studied families for the content of a
100 high number of components endowed with a large spectrum of biological activities; as
101 previously mentioned most of the essential oils belong to this category of components,

102 but another consistent portion of not volatile bio-active secondary metabolites is present
103 in this family.

104 These plants, which enclose the so-called medicinal and aromatic plants, are
105 altogether defined as officinal plants possessing specific sensorial, biological and
106 pharmacological properties. They cannot be classified according to the classic botanic
107 categories, namely herbaceous, arboreous, woody, etc., but the aforesaid characteristics
108 make these plants transversal from the botanic, agronomic and ecological point of views.
109 This large variability accounts for a very wide range of classification producing an
110 estimation ranging between 10,000 and 50,000 botanic species comprehensible in this
111 category. [3]

112 The term ‘officinal’, which characterizes these plants, derives from the Latin name
113 “*officina*”, namely the back of the medieval chemist’s shop, where the plants were
114 subjected to different manufactures (drying, grinding, soaking, essence production, etc.)
115 to obtain several marketable products.

116 According to a recent survey of World Health Organization issued in 2016, [4] the
117 main human pathologies and the leading causes of deaths were: ischemic heart disease,
118 stroke, obstructive pulmonary disease, respiratory infections, Alzheimer and central
119 nervous system (CNS) diseases, lung cancer, diabetes, road injury, diarrhoeal diseases,
120 tuberculosis. To-day also according to the World Health Organization (WHO) about 3.5
121 billion of people rely on medicinal plants for their healthcare requirements (WHO Global
122 report 2019). [5] This is particularly true in the developing countries of Asia, South
123 America and mainly Africa, where about 80% of population is based on this practice. [6]
124 Traditional Chinese Medicine (TCM), Ayurvedic medicine (India), South and Central
125 America, Middle East and Asia, as well as many other traditional popular treatments
126 based on medicinal plants in many countries are the expression of the large use of these
127 natural resources and a witness of a cultural heritage, which dates back to several
128 thousands of years.

129 In these last decades these typologies of medical remedies have assumed an even
130 more important role also in the western population, where the traditional uses of plants
131 have been lost, overshadowed or forgotten. In fact, according to the reports of WHO the
132 global market for herbal medicine raised from US\$17 billion in 2000, to US\$60 billion
133 in 2003; in particular, only the industry related to TCM in 2012 reached a value of UD\$83

134 billion, ^[7] a value of US\$115 has been estimated for 2020. ^[8] A little less than 30,000
135 plants are recorded as medicinal, but only about 4,500 have been phytochemically studied
136 and are used currently used in plant-based medicines. Therefore, a large portion of these
137 plants is still unexplored, representing a very important patrimony of new potential
138 bioactive compounds. ^[7,9]

139 As previously mentioned medicinal and aromatic plants, or in general officinal
140 plants, are not ascribable to a particular botanical or agronomic category, being
141 distributed among different orders, families and genera. The **Figure 2** shows the main
142 families in which are present the highest number of medicinal plants. As it can be seen
143 the Fabaceae or Leguminosae family contains the highest number of these species,
144 whereas the Lamiaceae family is in the second position.

145 **INSERT FIGURE 2**

146 Lamiaceae species are distributed nearly worldwide, however, the Mediterranean
147 basin is the region where this family found the best environmental conditions for its
148 development and differentiation. ^[10,11] The so-called “Mediterranean scrub” is in fact
149 characterized by a large number of Lamiaceae species, which form coastal and
150 mountainous scrublands (garrigues) with rocky and sandy soil, being the typical and
151 unique environmental peculiarity of a large portion of the Mediterranean area.

152 The discovery of new natural products and the more detailed studies of
153 phytocomplexes, here intended as a complex mixture of phytochemicals, represent the
154 more promising perspectives of this sector. As previously mentioned also in this family
155 many species have not yet been studied and therefore a lot of work must be still done.
156 However, *conditio sine qua non* that this knowledge became a shared patrimony is the
157 conservation, the cure and the attention towards the genetic resources and biodiversity.
158 This is strictly connected with the actions of the human beings and with the climate
159 change which already are producing serious problems to the plant survival. A study of
160 some years ago confirmed that this trend based on our incapacity to invert this course and
161 to make the politically correct choices could be the cause of the vulnerability and the
162 serious threat of more than half of species in Europe by 2080. ^[12]

163 Plants have ‘accompanied’ the evolution of the human beings supplying an
164 essential portion of foods, and a series of ‘tools’ able to fight several pathologies or
165 generically speaking many health problems. As previously mentioned to-day a large

166 portion of the world population uses still plants and their derivatives as medicinal
167 remedies, however, it also true that many drugs sold in our pharmacies and used against
168 a large spectrum of pathologies (chronic and/or epidemic), have a natural origin.

169 In recent years the use (and the abuse) of food supplements has grown
170 exponentially. To these products which can not be classified as food and/or drugs, the
171 term ‘nutraceutic’ has been associated, while a food containing some of these components
172 has been defined as ‘functional food’, namely a food able to help an organism to keep a
173 state of well-being, even though it is not generally accepted that such ‘functionality’, in
174 nutritional and healthy sense, can be associated only to a selected number of foods. [3,5,6]

175 The Lamiaceae family, object of this review, represents a small portion of the
176 important natural resources that the human beings should have carefully to protect.
177 Besides the classic and still effective studies aimed to the discover of new natural
178 products, several new tools are available helping the fight against the new healthy
179 emergencies of this millennium. One of these is the so-called multi-resistance of several
180 microbial strains towards a large number of antibiotics, which until few years ago were
181 able to contrast many types of infections. Unfortunately, owing to a large abuse of these
182 drugs against microbial strains, these have developed more or less marked resistance
183 making useless or little effective their action in the anti-infections therapies. This problem
184 is particularly acute in the hospitals and for some particular and very dangerous bacterial
185 strains as *Mycobacterium tuberculosis*, the etiological agent of tuberculosis (TB), which
186 in these last years, showed a broad recrudescence becoming one the main cause of death.
187 [13] New strategies such as the use of a multi-components therapy in the hope to exploit
188 synergic and/or additive effects, or the analyses of new and little explored new sources
189 has been taken in consideration. [14-16] Very recently some cyclic heptapeptideptides
190 have been synthesized taking paradoxically inspiration from a *Staphylococcus aureus*
191 toxin, showing interesting activities against antibiotic-resistant gram-positive and –
192 negative pathogens. [17]

193 A further and promising tool for the development of the natural products and
194 herbal extracts is the so called “drug repurposing”, which has been defined as the
195 “Studying the drugs that are already approved to treat one disease or condition to see if
196 they are safe and effective for treating other diseases” as elaborated by the National
197 Centre for Advancing Translational Sciences (NCATS, Bethesda, Maryland, USA) of the

198 National Institute of Health (NIH, Bethesda, Maryland, USA).^[18] This approach applied
199 in these last years in the pharmaceutical sector has been mainly developed for economic
200 reasons (but not only) because the production of a new drug is a very long (13-15 years)
201 and very expensive (2-3 US\$ billion) process, whereas the cost of a pharmaceutical
202 repurposing of an old drug is about 300 million of US\$, since in this case the toxicity and
203 clinical studies have been already carried out. As previously mentioned this approach is
204 applied also to natural products and plant extracts,^[19,20] and recently it has been
205 supported by several tools such as the Connectivity Map (CMap), the Library Integrated
206 Network based Cellular Signatures (LINCS), the Genome Wide Association Studies
207 (GWAS), the Side Effect Resource (SIDER), and the Directionality Map (DMAP), which
208 have significantly reinforced the drug repurposing applications.^[21-25]

209 These new studies besides to be applied to fight the main pathologies previously
210 described appear particularly promising in the research of therapeutic tools against the
211 so-called rare or orphan diseases, which are often chronic and progressive without an
212 appropriate drug equipment as well as focused pharmaceutical studies.

213 Aim of this review is to give an up-dated vision of the most recent studies on some
214 selected Lamiaceae plants, such as lavender, oregano, sage, rosemary, and thyme, which
215 characterize the patrimony of the Mediterranean area, with particular attention to southern
216 Italy regions. The increasing cultivation of these species and their growing diffusion as
217 new crops, as well as the evaluation of their new applications of the extracts and essential
218 oils in new sectors of their potential bioactivities will be discussed.

219

220 **2. Essential oils: very old “guys” with great prospects**

221 Essential oils (EOs) are volatile and complex mixtures of several compounds belonging
222 to different chemical families characterized by strong odor synthesized by aromatic plants
223 as secondary metabolites. These phytocomplexes should be obtained by physical
224 processes involving steam distillation, hydro or dry distillation with the sole exception of
225 *Citrus* EOs which are mechanically obtained by peel expression.^[5] After the culinary use
226 as dried or fresh spice, the extraction of EOs is the second main use for rosemary, sage,
227 oregano and thyme, while for lavender the production of the essences is probably the
228 main scope of its cultivation due the additional interest of the cosmetic industry. For the
229 above mentioned reasons several studies were conducted in the recent past with the aim

230 to optimize or increase the yields of EOs using novel techniques in addition or as “pre-
231 treatments” of the traditional way of production. Examples of these novel techniques are
232 microwave-assisted hydrodistillation, [26,27] solvent-free microwave extraction, [28]
233 ultrasounds, [29,30] ultrasonic-microwave assisted preceded by enzymolysis. [31] From
234 these studies, it emerges that the combination of advanced and traditional techniques leads
235 to a variable increase in yields and a significant reduction in times with consequent energy
236 savings. However, this approach also brings with it a further qualitative and quantitative
237 variability of essential oils that is added to that present in nature (conditioned by climatic
238 and environmental factors) with consequent variation of the biological activities of the
239 oils themselves. This parameter must be evaluated every time we want to produce an
240 essential oil that has a particular chemical composition using a combination of extraction
241 techniques.

242 From the point of view of biological properties and new applications what are
243 the most interesting prospects concerning essential oils and their production with
244 particular reference to the five species of Lamiaceae covered by this review? What
245 emerges from the literature analysis of the last 5-10 years is that the interest on essential
246 oils still remains the exploitation and optimization of the broad-spectrum antimicrobial
247 activity that these phytocomplexes possess. Innovation lies in the application of this
248 activity, which is now declined in the most varied industrial and human and animal health
249 fields. There is also a growing interest in the enhancement of peculiar endemisms and
250 chemotypes capable of enhancing a certain area of production, together with the
251 rationalization of crop management under stress conditions and agronomic techniques
252 able to optimize the productions in terms of biomass and essential oils yields and adapt
253 them even in difficult or extreme environmental conditions. Another aspect of great
254 interest is the research through nanoformulation techniques to overcome the problem of
255 the high volatility and poor solubility in water of essential oils that often limits its
256 potential use at agronomic and industrial level.

257 Progress has also been made in the knowledge and understanding of the
258 potential of Lamiaceae essential oils as antigerminative and phytotoxic agents capable of
259 being used as an eco-sustainable alternative in the fight against weeds. Finally, there are
260 still numerous studies on the potential repellency and toxicity of these essential oils
261 against the harmful insects of agricultural production or vectors of potential diseases for

262 men or animals. Also in these cases, the combination with nanoformulation techniques
263 seems to be the most promising route.

264 *2.1 Enhancement of endemisms and chemical variations within populations*

265 The chemical variability of essential oils is a peculiar character of these natural matrices
266 being important to understand their biological properties, as well as for their
267 industrialization and commercialization. Owing to their compositional variability the
268 essential oils are able to perform several biological actions, however, at the same time
269 this variability can represent an obstacle for their industrialization processes. This
270 dichotomy has led on the one hand to the search for a rationalization and extremization
271 of the "chemotype" concept, on the other a search for particular endemisms and
272 compositional variations capable of preserving biodiversity and enhancing particular
273 production areas.

274 An exhaustive treatment of the chemotypes and endemisms of the five
275 Lamiaceae covered by this study is out of the scope of this review. Rather we will try to
276 emphasize the research published in recent years by providing a very general picture of
277 the most recent knowledge in this field.

278 *2.1.1 Lavender*

279 A very interesting dissertation on current lavender and lavender EOs market with an
280 evaluation of the economic impact of these productions in an officinal plants producer
281 country as Turkey was published recently by Giray, ^[32] concluding that the sector of
282 aromatic and medicinal plants and their essential oils is very promising at the socio-
283 economic level for developing countries. An exhaustive work on the chemistry of
284 *Lavandula* genus was made by Aprotosoai and colleagues ^[33] covering the chemistry of
285 17 species from very different geographical origins. What emerges is that the chemical
286 composition of lavender essential oils have a huge range of variability in terms of volatile
287 constituents, concluding that more research is needed in terms of the knowledge on the
288 genotypic and environmental events that produce the lavender chemical biodiversity,
289 recalling for the standardization of methodology for the analysis and extraction. A
290 complementary list of recent publications is reported in **Table 1**. Among studies on *L.*
291 *angustifolia*, a recent work of Smigielski et al. ^[34] compares fresh and dried plant material
292 finding no significant differences in terms of chemical composition of essential oils but
293 rather a higher antioxidant activity in the oil from fresh aerial parts. Recent works ^[35-38]

294 report a linalool predominance in *L. angustifolia* essential oils from Italy, Croatia, Turkey
295 and Spain respectively. Kivrak and Singh et al., [39-40] report two cases of linalyl acetate
296 chemotypes from Turkey and India respectively, while Behladj et al., [41] reports 1,8-
297 cineole chemotype in Algerian populations.

298 Some reports on the evaluation of inflorescences, aerial part and whole plant
299 EOs from *L. stoechas* from Spain and Italy, reporting a fenchone chemotype followed by
300 camphor and 1,8-cineole as other main components. [42-44]

301 *L. hybrida* (several cultivars) from Italy and Turkey were extensively studied
302 by Pistelli et al., [35] and Kivrak et al., [39] while Robu et al. and Bajalan et al. published
303 analysis of plant material from Romania and Iran, respectively. [45,46] The picture that
304 emerges regarding *L. hybrida* is more varied, from a compositional point of view, than
305 that present for *L. angustifolia*.

306 Regarding the less studied species of lavender, very interesting is the work of
307 El Hamadouï et al., [47] which report the chemical composition and biological activities
308 of a carvacrol chemotype of *L. mairei*, an endemic and rare species from Morocco.
309 Another two endemic species are *L. pinnata* and *L. pubescens* collected in Spain and
310 Yemen, were studied by Argentieri et al. and Al Badani et al., [48-49] Both were carvacrol
311 chemotypes with a variable amount of carvacrol methyl ether and caryophyllene oxide as
312 other main compounds. Finally, an endemic and rare species namely *L. stricta*, was
313 studied by Alizadeh and Aghae reporting a rare case for this genus of α -pinene
314 chemotype EO. [50]

315 INSERT TABLE 1

316 2.1.2 Oregano

317 A recent work published by Leyva-López et al. [52] provides a broad overview of the major
318 oregano species and their chemical variability, relating them to their biological activities
319 reported in literature. The constituents and concentration of the compounds of the EOs
320 usually vary due to a great diversity of factors such as species, soil conditions, harvest
321 season, geographical location, climatic and growth conditions. A list of recent studies on
322 chemical composition of oregano species EOs is presented in **Table 2**.

323 Within oregano, *O. vulgare* is probably the more widespread specie worldwide
324 and great attention is still paid by researchers to it. A huge work was focused on the
325 qualitative and quantitative composition of essential oil compounds of European

326 *Origanum vulgare* analyzing more than 500 plants from 17 countries. [53] Their work
327 confirms a large variability in terms of yields (0.03-4.60%) and chemical composition,
328 which can be partially explained by different efficiency or activity (up-/down regulation)
329 of the cymyl-, sabinyl- and linalool/linalyl acetate pathway. Their very interesting
330 conclusion is that “the cymyl- and the acyclic pathway were usually active in plants from
331 the Mediterranean climate whereas an active sabinyl-pathway was a characteristic of
332 plants from the Continental climate”. The picture is completed by the works of De Mastro
333 et al. and Napoli et al., [54-55] for southern Italy populations reporting the familiar chemical
334 variability and by the carvacrol-chemotypes populations reported for Spanish [56] and
335 Iranian [57] *Origanum vulgare* L. Hatipi et al. report a case of sabinene chemotype *O.*
336 *vulgare* in Kosovo. [58] Argentinian oregano species were studied by Asensio et al., [59]
337 which reports *trans*-sabinene hydrate as main compound for *Origanum x Majoricum*, *O.*
338 *vulgare* ssp. *vulgare* and *O. vulgare* ssp. *hyrtum*. Within *O. vulgare* subspecies, *O.*
339 *vulgare* ssp. *hyrtum* is largely diffused worldwide and is very important under a
340 commercial point of view. It was widely studied by Tuttolomondo, Mancini and
341 collaborators reporting a prevalent thymol chemotype for Italian populations, [60-63]
342 (**Figure 3**) and by Stesevic et al. [64] for populations from Montenegro showing instead a
343 carvacrol chemotype. This differentiation is appreciable at the olfactory and organoleptic
344 level by consumers as shown by a recent comparative study of the chemical composition
345 and sensory analysis of Sicilian oregano and a commercial sample (**Figure 4**). [65] These
346 aspects have positively influenced a large increase of its production (**Figures 5 and 6**)
347 which has been favorably received by the market and consumer.

INSERT TABLE 2

INSERT FIGURES 3-6

350 Recently, many studies have been carried out to re-evaluate chemical
351 composition of endemic species of oregano in different countries such as *O. compactum*,
352 endemic of Maghreb, [66-68] *O. ehrenbergii* [69] and *O. libanoticum* [70,71] endemic of
353 Lebanon, *O. acutidens* endemic of Turkey, [72] and *O. floribundum* endemic of Algeria.
354 [73]

355 2.1.3 Rosemary

356 Among the five Lamiaceae subject of this review, rosemary is the one that has the least
357 chemical variability in terms of the composition of essential oils as confirmed by the

358 results of the bibliographic research of the last 10 years. A list of some of the most
359 significant recent works is shown in **Table 3**, whereas **Figure 7** shows a Sicilian
360 cultivation of this plant.

361 Probably the unique species of rosemary with a great economic interest is *R.*
362 *officinalis* L. for its use as spice and for the biological properties of its extracts and
363 essential oils with putative health benefits. [86] This is confirmed by the recent work of
364 Borges et al., in which they reviewed studies of *R. officinalis* and its essential oil, giving
365 prominence to its ethnopharmacological importance, phytochemistry, and main
366 biological activities underlying mechanisms of action of its major molecules. [87] Other
367 relevant and recent investigations confirm that EO composition of *R. officinalis* is
368 characterized by the presence of 1,8-cineole, camphor and α -pinene as main compounds
369 although with some differences in terms of relative percentages of each. For the other
370 species of rosemary, the analyses of the essential oils of stems, leaves and flowers of *R.*
371 *ericalix*, showed that camphor is the main component in all three parts of the plant
372 examined. [88]

373 **INSERT TABLE 3**

374 **FIGURE 7**

375 *2.1.4 Sage*

376 Sage is one of the largest genera of the Lamiaceae family. Some species such as *S.*
377 *officinalis*, *S. sclarea*, *S. lavandulifolia*, *S. miltiorrhiza* and *S. hispanica* are more
378 important under a commercial point of view for their use as food/spices and because of
379 the interest in their essential oils. Through the study of the existing literature, [101] sage
380 shows a high chemical diversity in its secondary metabolites. This diversity and richness
381 has allowed this plant to remain until today one of the most studied plants in search of
382 new bioactive metabolites or to confirm with more current scientific data the evidence of
383 traditional therapeutic use. As for the other medicinal plants, the quantitative yield and
384 the quality of the essential oil, also in the case of sage, is strongly conditioned by the
385 species, the geographical area of origin, the time of harvest and the agronomic techniques
386 used for the its cultivation. A non-exhaustive list of sage species studied in the last 5 years
387 is shown in **Table 4**. From the data examined the confirmation of the thujone/camphor
388 chemotype is obtained for *S. officinalis* from different geographical origin. [102-104]

389 Some species show high values of caryophyllene and caryophyllene oxide such
390 as *S. hispanica*,^[105] Turkish *S. trichoclada* and *S. multicaulis*,^[106] *S. montbretii*,^[107]
391 Iranian *S. virgate*, *S. reuteriana* and *S. multicaulis*,^[108] Iranian *S. nemorosa*^[109] and
392 Turkish cultivated *S. forskahlei*.^[110] A lot of species have 1,8-cineole as main component
393 such as *S. lavandulifolia*, *S. bucharanii*, *S. bucharica*, *S. palaestina*, *S. ringens* and *S.*
394 *veneris* from Italy.^[111-117] **Figure 7** shows a cultivation of sage in Sicily, and a particular
395 of its inflorescence.

396 **INSERT TABLE 4**

397 *2.1.5 Thyme*

398 *Thymus* is probably the most taxonomically complex genus of the Lamiaceae family. For
399 this reason giving a succinct account of endemisms and chemical variability of its
400 essential oils is particularly difficult. Recently, Salehi et al.,^[141] provided an overview of
401 the phytochemistry of the EOs of this genus from different locations worldwide
402 confirming the presence of a significant intraspecific chemical diversity in which the two
403 most common chemotypes, thymol and carvacrol, are preponderant. Similar results have
404 been presented within the work of Tohidi et al.,^[142] in which main *Thymus* species from
405 the Iranian region were studied. Trindade and collaborators^[143] addressed the study of
406 terpene synthase genes in the genus *Thymus* by correlating these data to the observed
407 chemotypes, stressing the importance of integrating molecular and biochemical data on
408 chemotype determination. Experimental study of wild samples of *T. capitatus* was made
409 by Napoli et al. and Saija et al.,^[144,145] giving a broad picture of the chemical composition
410 of thyme populations in Sicily (South Italy) with a predominance of the carvacrol
411 chemotype, as per as Tunisian^[146] and Spanish.^[147] EOs from Spanish *T. caespitius*, *T.*
412 *zygis*, *T. mastichina* and *T. capitatus* flowers and fruits were evaluated by Delgado et al.,
413^[148] not showing significant compositional differences between the two parts of the plant.
414 Other examples of studies recently published on *Thymus* endemisms are listed in **Table**
415 **5**.

416 **INSERT TABLE 5**

417 *2.2 Crops management and agronomic techniques*

418 In recent years, much attention has been paid on the effects that some cultivation
419 techniques may have on the growth of officinal plants and on the quality of their essential
420 oils. Many studies are focused on the growth of these plants in conditions of water and

421 salt stress in order to enhance areas with unfavorable pedoclimatic conditions. Nowadays
422 due the increasing of world demand of high quality essential oil the optimization of the
423 harvesting period is becoming crucial to obtain better yields and better products. New
424 data have been released on the effects of phenological stages or harvesting time on the
425 quality of the essential oils. Collection of *O. vulgare* at pre-flowering time seems to
426 increase the essential oil percentages, ^[153] while the flowering stage seems to be the best
427 period to increase some biological activities of *O. compactum*. ^[154] A study conducted on
428 *O. majorana* shows that the harvest time can influence the chemical composition of the
429 essential oil. In fact, the harvest of the herb at full flowering stage allows obtaining an oil
430 containing more *cis*-sabinene hydrate and sabinene than that obtained from the second
431 harvest done at a later time. ^[155] Moreover, the essential oil of *Origanum vulgare* ssp
432 *glandulosum* from North Africa showed a characteristic and different chemical profile
433 from year to another. ^[156] Also quantity and quality of *L. angustifolia* EO is positively
434 regulated by temperature and flowering stage development while it was negatively
435 affected by rainfall during flowering period because the remarkable decline of linalool
436 production. ^[157] Recent studies show that early-summer or summer is the best harvest
437 period for *S. fruticosa* ^[158] and *S. officinalis*, ^[159] for which flowering period as best
438 phenological phase is confirmed, ^[160] as per as for *S. verbenaca*. ^[161] The seasonal
439 variation showed significant effects in the composition of the essential oils of *R.*
440 *officinalis* with a greater amount of the oil and the main components in the summer. ^[162]
441 Very interesting is a recent work evaluating the effect of diurnal variations on chemical
442 composition and biological activities of *S. officinalis* EOs. ^[163] What is interesting in the
443 discussion on the correct interpretation and assignment of chemotypes is that the results
444 of this study indicate that the EO of plants harvested at 7 am and 12 pm were of α -thujone
445 chemotype while those collected at 5 pm were of camphor chemotype with an important
446 impact on biological activities. The effect of low-light conditions on *S. officinalis* growing
447 in a greenhouse over the winter was studied by Mapes and Xu. ^[164] Authors concluded
448 that with reduced light intensity, plant height increased whereas leaf size and number
449 decreased. Similar results was obtained for *O. vulgare*. ^[165] However, growing herbs in
450 northern climates could be a challenge to consider as a new opportunity for our 'old guys'.
451 And in this perspective fits the promising study by Shiwakoti et al., ^[166] who have grown
452 thyme, oregano and rosemary using plastic tunnels. The ornamental potential and freezing

453 tolerance of six *Thymus* species was studied by Lajayer and collaborators ^[167] in order to
454 use thyme as ground-covering plants in landscaping, with the results that all the examined
455 species showed valuable ornamental characteristics and freezing stress tolerance.

456 As already said EOs yields are influenced by various factors, so it is important
457 to keep finding new growing procedures that increase the quantitative content of EOs.
458 The nutrient level in the soil is one of the most investigated aspects of research into
459 aromatic plants with contrasting results, especially when the goal of their cultivation is
460 different from herbage yield. ^[168] The foliar application of nutrients and abiotic elicitors
461 seems to be a promising technique. Foliar application of 1.5 mg/L of 24-epibrassinolide
462 (a brassinosteroid analogue) on a 8 years old plants of *Lavandula x intermedia* var. Super
463 increase the EO yields from 6.2% to 8.7% (v/w) and also the percentage of linalyl acetate
464 from 27.3% to 30.5% . ^[169] Negative impact on the quantity and quality of the *O.*
465 *majorana* EO has been expressed by the application of hydroalcoholic extract of *Zingiber*
466 *officinale*, ^[170] while a considerable increment of EO quantity and thymol stabilization of
467 percentage range within analyzed population of *T. vulgaris* was obtained by Pavela et al.,
468 ^[171] with the foliar application of mixed N, P, K and salicylic acid. Again, a recent study
469 on *T. vulgaris* showed that treatment with abiotic elicitors lead to differential up-
470 regulation of genes involved in thymol/carvacrol biosynthesis with the probable
471 consequent increment of the corresponding metabolites. ^[172] Rosemary tolerance to the
472 salinity stress after foliar treatments with zinc-oxide in common and nano-form was
473 studied by Mehrabani et al.: ^[173] it seems that zinc foliar spray improved flavonoids, total
474 soluble solids and essential oil contents. Salinity as abiotic stress is a permanent major
475 threat to the agriculture industry worldwide and usually associated with reduced growth
476 and productivity of cultivations. Several approaches had been adopted to control salinity
477 adverse effects on plants and one of the more interesting is the use of salicylic acid. The
478 effects of the use of salicylic acid on *R. officinalis* were studied by El-Esawi and
479 collaborators. ^[174] Salinity stress caused the reductions in α -pinene, β -pinene, and cineole
480 along with increases in linalool, camphor, borneol, and verbenone. However, salicylic
481 acid applications at 100–300 ppm largely reversed such effects of salinity. Another recent
482 work shows the relationship between EO yields of *R. officinalis* and salinity conditions
483 either alone or in presence of plant growth-promoting rhizobacteria (PGPR). ^[175] EO yield
484 increases with increasing salinity levels up to 10 g/L NaCl, but decreased with further

485 increases in salinity levels in treatments without using PGPR and being constant in
486 treatment with PGPR. Moderate NaCl salinity was shown to improve the yields of volatile
487 compounds and to modify the relative percentages of main compounds in *S. mirzayanii*
488 and in this process cineole synthase gene is deeply involved. ^[176,177] *L. angustifolia* grown
489 hydroponically and subjected to salt stress have been studied by Chrysargyris et al., ^[178]
490 finding that high salinity decreased essential oil yield, while low-moderate salinity levels
491 maintained the volatile oil profile in lavender. The integrated foliar application of K and
492 Zn lighten the presumable detrimental effects of salinity in terms of fresh biomass,
493 antioxidant capacity, and EO yield. Also Mehrabani et al. reached similar conclusions on
494 *L. stoechas*. ^[179] Additionally increasing salinity of the applied water caused significant
495 decreases in total fresh, total dry and dry leaf yield, total essential oil yield and antioxidant
496 activity values of oregano (*O. onites*). ^[180]

497 Sustainable water management has become a global emergency and studies
498 have also multiplied on the cultivation of medicinal plants in conditions of water stress
499 or drought. Oregano can be significantly affected by water stress conditions in terms of
500 herbal yield, essential oil content and composition. *O. vulgare* ssp. *gracile* seems to suffer
501 less than *O. vulgare* ssp. *virens* in terms of EO composition, ^[181] while the beneficial
502 effects of sprayed chitosan on *T. daenensis* ^[182] and on the expression increments of
503 stress-related genes in *O. majorana* was demonstrated. ^[183] Anyway drought stress
504 triggered different responses in the Lamiaceae species as demonstrated by García-
505 Caparrós et al. in their study on six different species. ^[184] Authors recommend the
506 cultivation of *S. lavandulifolia*, *T. capitatus*, and *T. mastichina* because there are no
507 differences in essential content but if there is a water-saving rate of 30%, while the
508 essential oil content decreased in *L. latifolia* and *S. sclarea* plants under water deficit
509 conditions. EO content and quality of *L. angustifolia* and *S. fruticosa* seemed to increase
510 under water stress conditions in a study of Chrysargyris et al. ^[185] This behavior is
511 confirmed also for *S. dolomitica* ^[186], *S. officinalis* ^[187] and *S. sinaloensis* ^[188] and some
512 Greek Oregano populations. ^[189] Foliar application of salicylic acid under water stress
513 conditions increase EO content on *T. kotschyanus*. ^[190] The importance of water
514 management on this type of cultivations is confirmed by numerous studies on combined
515 variables such as drought/manure or nitrogen application, ^[191] water stress/arbuscular
516 mycorrhizal fungi, ^[192,193] water stress/different soil fertility systems, ^[194] irrigation

517 frequency/manure, ^[195] irrigation frequency /chitosan application, ^[196] irrigation
518 frequency/mineral fertilization. ^[197] An interesting study on the effect of irrigation with
519 secondary-treated effluent, as compared with potable water irrigation, on the
520 composition, biological activities and yield of EO from *O. syriacum* L. var. *syriacum* was
521 conducted by Ali-Shtayeh et al. ^[198] and their data demonstrate that this kind of water can
522 be successfully utilized for irrigation for essential oil production from *O. syriacum*.

523 In order to improve plant productivity performance the mycorrhizal inoculation
524 is a promising technology in sustainable agricultural system already investigated also in
525 aromatic herbs and recently some interesting data were published on normal soils ^[199,200]
526 and industrially polluted soil. ^[201] Heavy metal contaminated soils due to industrial
527 activities are a major environmental problem that can reduce both the productivity of
528 plants and the safety of plant products. Phytoremediation promoted by aromatic plants
529 could be a promising method for removal of heavy metals from soil due their capability
530 in terms of phytoextraction of metals with a negligible transfer of them to EOs. In this
531 light is very interesting the work of Pistelli and collaborators ^[202] on some spontaneous
532 species growing in an abandoned mining of Elba island and the work of Stancheva et al.,
533 ^[203] on *Origanum vulgare* L. grown on industrially polluted soil.

534 2.3 Herbicidal activity, phytotoxic potential and pest control

535 Interference of weeds with agricultural crops causes huge economic losses to farmers
536 reducing crop quality and quantity, and increasing cost in terms of work and herbicides
537 used to control them. Furthermore, the increasing resistance of weeds has resulted in a
538 dramatic increase in the use of herbicides. Nowadays, the negative effects of synthetic
539 herbicides to human health and environment is regarded as a real problem and trying to
540 solve or mitigate it is a novel priority. Essential oils have been extensively studied in the
541 past as valuable eco-friendly compounds with herbicidal activity and the interest is still
542 high on the possible applications of them in the open field. Phytotoxic potential of *O.*
543 *vulgare* ssp. *hyrtum* EO has been evaluated on plant model *Arabidopsis thaliana* through
544 a physiological and metabolomic approach by Araniti et al., ^[204] with the evidence that
545 EOs firstly caused growth reduction and leaf chlorosis, together with a series of
546 interconnected metabolic alterations. Interesting is the work of Atak et al., ^[205] on the
547 herbicidal effect of *O. onites* and *R. officinalis* EOs on germination and seedling growth
548 of bread wheat and weeds (*Avena sterilis* and *Sinapis avensis*). Authors state that wheat

549 cultivars were less affected compared to weed species suggesting proper dose of essential
550 oils could be used as a bio herbicide for weeds control. These results could be useful for
551 wheat breeders to improve varieties resistant to specific allelochemicals, which kill other
552 weeds causing yield losses. EOs of commercial oregano, marjoram and *T. mastichina*
553 have been compared for their phytotoxicity against *Portulaca oleacea* L., *Lolium*
554 *multiflorum* Lam. and *Echinochloa crus-galli* (L.) Beauv. in the work of Ibanez and
555 Blazquez. [206] Their results show that Oregano essential oil completely inhibited seed
556 germination and seedling growth at all concentrations assayed, whereas marjoram and *T.*
557 *mastichina* essential oils only showed significant effects in hypocotyl and/or hypocotyl +
558 radicle length depending on the weed and dose. Among other EOs, *O. vulgare* L. was
559 experimented for their herbicidal activity against *Sinapis avensis* weed at different
560 concentration then compared with commercial herbicides. [207] Although the results are
561 promising about the EOs use as alternative herbicides, the authors propose further studies
562 are required to determine the cost, applicability, safety and phytotoxicity against the
563 cultivated plants. *O. vulgare* EO shows a significant inhibitory effect on *Solidago*
564 *canadensis* seeds germination in a study where other EOs are ineffective and significant
565 differences were noted by using different concentrations of single components. [208] EOs
566 chemical composition is affected by plant phenological stages so their biological activity,
567 including phytotoxicity, could be influenced by this factor as stated by Alipour &
568 Saharkhiz in their study on *R. officinalis*. [209]

569 The encapsulation on a starch matrix of *R. officinalis* EO for phytotoxic
570 purposes has been reported for the first time very recently by Alipour et al., [210] in a study
571 on inhibition effects against *Amaranthus retroflexus* and *Rhaphanus sativus* under
572 greenhouse conditions. A larger study involving 12 EOs (*L. angustifolia*, *S. officinalis*
573 and *T. vulgaris* among the others) and their inhibitory effect on four weeds (*Amaranthus*
574 *retroflexus*, *Avena fatua*, *Bromus secalinus*, *Centaurea cyanus*) and three crops (*Avena*
575 *sativa*, *Brassica napus* and *Zea mays*) was conducted on 2017 [211] and it is worth
576 mentioning that three tested crops were significantly more tolerant than the weeds tested.
577 Synergistic allelopathic potential of *S. officinalis* and *T. vulgaris* EOs was studied by
578 Alexa et al., [212] and the results indicated that the tested EOs alone as well as in
579 combination have allelopathic effect against investigated seeds. Their results revealed

580 significant differences in the inhibitory effect of the investigated EOs on germination
581 capacity of wheat and tomato versus weed species.

582 The biological activity of essential oils on insects is one of the most studied
583 aspects (**Table 6**). Not only was the toxicity of these substances investigated, but also
584 their potential repellency activity due to volatility and their ability to behave as signal
585 molecules. Despite the good results obtained in many *in vitro* studies, the use of essential
586 oils on a large scale and in the open field for the control of pests has not yet wide diffusion.
587 This is mainly due to outdoor persistence problems and to the difficult dissolution in water
588 with the need to use surfactants to facilitate and homogenize the distribution over large
589 areas. Moreover, much published literature demonstrates a not high selectivity of the
590 insecticidal or repellent action and is often lacking in data on non-target insects and more
591 generally on their impact on the ecosystem.

592 **INSERT TABLE 6.**

593 These findings could improve the industrial formulation (also again with the aid
594 of nano-techniques) in order to overcome the obstacles that have slowed down the use of
595 essential oils in this field.

596 *2.4 Diet supplement for animals feeding and application for animal health*

597 The use of EOs as diet supplements for farm animals is an interesting application of these
598 phytocomplexes. The aim is to exploit mainly their antimicrobial potential and their
599 interaction with some physiological parameters to improve growth performances, meat
600 quality and animal wellness under stress conditions such as transportation and heat-stress.
601 Another emergent aspect is the potential role of these natural compounds on ruminant
602 fermentation due the increasing concern over their methane emissions and their
603 consequent contribution to global warming. The latest estimates calculate the contribution
604 of ruminants equal to 16-25% to the total greenhouse gases emissions and EOs are
605 recognised as safe a rumen modifier feed additives. ^[237] Three works of Cobellis and
606 collaborators ^[238-240] draw a very clear picture of the advantages and disadvantages of
607 using secondary metabolites such as rumen modulators evaluating scientific production
608 in recent years including essential oils. Authors concluded that if on one hand EOs may
609 be promising natural substances to mitigate rumen methane production, improve rumen
610 fermentations, and reduce environmental impact of ruminant production from the other
611 side further research will be required to determine their active compounds/effective

612 doses/mode of action, effect on organoleptic characteristics of animal products in order
613 to evaluate a clear cost–benefit ratio prior to use them as additives at farm level. In
614 addition, a study on *T. vulgaris* and other EOs by Gunal et al. [241] comes to similar
615 conclusions stating that the addition of thyme EO at the high doses resulted in lower
616 methane production but it had negatively affected rumen microbial fermentation.
617 Ruminants are not the only animal category under experimentation in the last years with
618 EOs as food supplement as showed in **Table 7**.

619 For example a lot of studies were focused on broiler chicken. *L. angustifolia* EO
620 at the dose of 0.4 mL in 1L of drinking water significantly affected some production
621 results as body weight, feed and water conversion ratios. [242] While a supplementation of
622 Oregano EO at the dose of 25 mg/kg to a normal diet promote a better average body
623 weight, feed conversion ratio decreasing mortality. [243] In the same study the composition
624 of the microflora was determined at the day of slaughter. For the oregano supplemented
625 groups significantly higher values of lactic acid and bifidobacteria in ileum and caecum
626 were noted, while, in caecum, reduced counts of coliform bacteria ($P<0.05$) were also
627 recorded compared to control group. A concern about the use of aromatic plants and EOs
628 as food additive in milk-producing farm animals is that their flavors and aromas may
629 make their way into the milk, altering its sensory properties. Two levels of essential oils
630 of caraway and oregano (0.2 and 1.0 g of oil/kg of dry matter) were added to the feed of
631 lactating cows for 24 d in a study of 2016. [244] With this treatment the amount and
632 composition of volatile terpenes were altered in the produced milk, and the sensory
633 properties of the produced milk were altered as well, and milk samples from animals
634 receiving essential oil treatment were perceived as having a fresher aroma and lower
635 stored aroma and flavor. Interesting are the studies on the effects of EOs dietary
636 supplementation on animals under stress conditions. Heat stress, for example, causes dire
637 economic losses in poultry. The *O. syriacum* EO showed some protective effects against
638 heat stress in broilers, [245] while dietary supplementation with *O. vulgare* EO may be
639 superior in alleviating the negative effects of transportation on pigs by improving the
640 pigs' antioxidant status in a comparison study with dietary supplementation of vitamin E.
641 [246] The administration of EO in the diet is becoming an interesting tool also in
642 aquaculture, with the aim of exploiting above all the antimicrobial activity of these
643 phytocomplexes. *O. majorana* EO and its nanocapsules were evaluated recently for their

644 antibacterial activity in silver catfish, *Rhamdia quelen*, infected with *Aeromonas*
645 *hydrophila*.^[247] All treatments improved the survival of infected fish, but authors suggest
646 daily baths containing 20 µL/L EO or 5 µL/L nanoencapsulated EO for five consecutive
647 days because these are the lowest effective concentrations tested which did not interfere
648 with the metabolic parameters of the animals. The *in-vitro* antimicrobial potency against
649 *Aeromonas hydrophila* and other two bacterial fish pathogens *Streptococcus iniae* and
650 *Photobacterium damsela* subspecies *damsela* of four EOs (*Origanum vulgare*,
651 *Eucalyptus globulus*, *Melaleuca alternifolia* and *Lavandula angustifolia*) was evaluated
652 by Gholipourkanani et al.^[248] All treatments showed antibacterial activity, and in almost
653 all cases the activity of the nano-emulsions was superior to their essential oil counterparts.
654 Origanum essential oil had a pronounced influence on the innate immunity and increased
655 the *Tilapia zillii* resistance to *Vibrio anguillarum*^[249] and dietary supplementation with
656 0.02% oregano essential oil is a practical prevention strategy for *Ichthyobodo salmonis*
657 and *Trichodina truttae* infection in *Oncorhynchus keta*.^[250] Finally, a relevant and
658 promising study on the efficacy of several EOs against *Pseudomonas* spp. isolated from
659 fish was conducted by Kacániová et al.,^[251] suggesting that the EO may be used as natural
660 compounds with antipseudomonal activity to improve the microbiological quality of
661 freshly caught freshwater fish.

662 A new vapour phase assay for evaluating the antimicrobial activities of EOs
663 against bovine respiratory pathogens (BRPs) was proposed by Amat and collaborators
664^[252] which have highlighted for *T. zigys* (among the others) vapour phase antimicrobial
665 activity against *M. haemolytica* S1, *H. somni* and *P. multocida*. On the basis of these first
666 results authors declare that future studies are needed to determine whether the application
667 of these EOs as a nasal spray can mitigate BRPs, as alternatives to antibiotics.
668 Antimicrobial activity of *L. berlandieri* and *T. vulgaris* EOs in gaseous phase was studied
669 also by Reyes-Jurado and collaborators^[253] with promising results suggesting a potential
670 use also to protect packed food against the growth of microorganisms.

671 De Aguiar and collaborators^[254,255] recently published two works on the
672 antimicrobial potency of several essential oils against one of the major swine pathogens
673 *Streptococcus suis*. In the first study eight EOs were tested against 19 *S. suis* strains
674 isolated from diseased and healthy carrier pigs concluding that the essential oils of
675 oregano, red thyme and common thyme showed a notable *in vitro* bactericidal activity,

676 by vapor and/or direct contact. In the second study, authors changed the approach
677 evaluating the combined interaction of four essential oils or their main components with
678 conventional antimicrobials against 53 resistant *S. suis* strains. The positive interaction
679 observed in their study between conventional antimicrobials and essential oils suggest
680 that combination therapy is a promising alternative for the control of diseases caused by
681 *S. suis* in pig farms. Mastitis is a mammary inflammation and is one of the most
682 economically impacting health events in the dairy cattle industry with high losses due to
683 veterinary and treatment costs and reduced milk production. ^[256] As alternative to
684 conventional antibiotics, EOs have been evaluated also against mastitis pathogenic agents
685 such as bacteria, fungi and microalgae. EOs from *O. floribundum*, *R. officinalis* and *T.*
686 *ciliatus* were tested *in-vitro* against *Candida albicans* isolated from bovine clinical
687 mastitis ^[257] and all essential oils revealed highly effective anticandidal activity. Several
688 essential oils (*Thymus vulgaris* L., *Origanum vulgare* L. and *Origanum majorana* L.
689 among others) were tested also against ten *Prototheca zopfii* strains that cause
690 inflammation of the mastitis in cows ^[258]. Results show that the studied essential oils can
691 effectively reduce the growth of *P. zopfii* strains (MIC between 0.25-1.0 µl/ml), including
692 those resistant to antifungal chemotherapeutics used in the treatment of protothecosis.
693 The antimicrobial activity of *L. stoechas* subsp. *luisieri*, *R. officinalis* and *T. mastichina*
694 EOs among others was investigated towards *Staphylococcus aureus* and *Staphylococcus*
695 *epidermidis* isolates from ovine mastitic milk origin. ^[259] Results of disk diffusion assay
696 revealed that *L. luisieri* and *T. mastichina* EOs are highly active against both *S. aureus*
697 and *S. epidermidis* strains, whereas *R. officinalis* EO is highly active against *S. aureus*
698 strains but inactive against several *S. epidermidis* isolates. Finally the effects of
699 intramammary infusion of sage (*Salvia officinalis*) essential oil on milk somatic cell
700 count, milk composition parameters and selected hematology and serum biochemical
701 parameters in 20 ewes affected with subclinical mastitis were studied by Alekish and
702 collaborators. ^[260] In this study, the intramammary infusion of sage EO to ewes affected
703 with subclinical mastitis resulted in a significant decrease in milk somatic cells count 24
704 h and 48 h post treatment. Furthermore, milk fat and lactose were increased in animals
705 that received the EO as well as in those treated with the amoxicillin as reference antibiotic.

INSERT TABLE 7

707 *2.5 Antimicrobial activity*

708 The antimicrobial activity of essential oils is one of the oldest among those attributed to
709 these phytocomplexes and consequently the most studied. It is therefore difficult to
710 extrapolate from such a diverse context, which are actually the most current trends and
711 the most promising prospects. In this section, we will therefore try to give an account of
712 some of the most interesting studies of the last few years, trying to draw at the end of the
713 considerations on the road we are taking. Since the crisis of antibiotics emerged as a
714 global emergency, mainly due to the emergence of bacterial multiresistance, essential oils
715 have represented more than just hope as broad-spectrum antimicrobial agents.

716 The chemical diversity, the undeniable efficacy *in vitro* and the ability to
717 perform antimicrobial activity not thanks to the presence of only one active ingredient,
718 but to the synergy of several structurally very different molecules have left hope for many
719 years to find the right way to fight less serious infections with natural, less toxic and even
720 inexpensive substances such as EOs. But this hope had to come up against the
721 hydrophobia, the aforementioned chemical variability that limits its standardization, the
722 extreme volatility of these phytocomplexes, the lack of information on mechanism of
723 action, factors that have turned out to be obstacles that have not yet been overcome and
724 that turn into a lack of *in vivo* and clinical data on their effectiveness. Nevertheless, the
725 research continues to produce interesting studies especially with regard to multi-resistant
726 strains and the use of nanotechnologies gives a glimpse of a new era for essential oils in
727 microbiology with an inevitable impact on human and animal health and on other fields
728 of application such as the food industry. All our five Lamiaceae species are largely studied
729 for the antimicrobial activity of their EOs. *In vitro* antibacterial activity of essential oils
730 from *O. vulgare*, *T. vulgaris*, and *L. angustifolia*, against 32 erythromycin-resistant and
731 cell-invasive streptococci isolated from children with pharyngotonsillitis was studied by
732 Magi et al. [279] Thyme and Origanum essential oils demonstrated the highest
733 antimicrobial activity with minimal inhibitory concentration (MIC) ranging from 256 to
734 512 µg/mL. A synergistic action of the EO main compound carvacrol and erythromycin
735 was detected in 17/23 strains using 24-h time-kill curves. Antimicrobial activity against
736 multiresistant clinical isolates of 11 pathogenic bacteria of a carvacrol chemotype of
737 *Lavandula coronopifolia* was assessed. [280] In a recent study Thyme EO is able to reduce
738 of 59.7-85.0% biofilm biomass formed by methicillin resistant *Staphylococcus aureus*
739 (MRSA) strains, [281] while *L. multifida* L. EO showed anti-MRSA activity since the

740 inhibition diameters reached 27 mm and MICs were lower than 0.1 µg/mL in a study
741 against 14 strains. [282] Synergistic effects of sub-inhibitory concentrations of *S. officinalis*
742 EO and different antibiotics against MRSA were also evaluated by Milenković et al. [283]
743 Authors highlighted the presence of different interactions between tested oil and
744 antibiotics from synergistic to antagonistic on different clinical isolates of MRSA
745 suggesting the necessity for systematic *in vitro* studies of interactions of EOs and
746 antibiotics to reveal any undesirable combinations. The antimicrobial activity against
747 multi-drug resistant *Escherichia coli* J53 R1 of *L. angustifolia* EO in combination with
748 piperacillin was also evaluated. [284] Furthermore, their stability against sodium dodecyl
749 sulphate, the scanning electron microscopy analysis and zeta potential measurement
750 revealed that the EO played a role in disrupting the bacterial cell membrane while
751 reduction in light production expression of *E. coli* showed the presence of potential
752 quorum sensing (QS) inhibitors. The crucial role of the membrane disruption in the
753 antimicrobial action of the EO is confirmed by other studies such as the one on *O. vulgare*
754 in which the mode of action of the oil is investigated by using a simple approach which
755 is the estimation of the total proteins in the culture media before and after the addition of
756 increasing concentrations of EO. [285] The conclusion of this study state that oregano EO
757 is effective against clinical pathogens as *E. coli* and *S. aureus* with a mode of action
758 targeting the bacterial membrane permeability or integrity. However, bacteria resistant to
759 chemical antibiotics seem to be capable of overcoming the action of EOs and this is a
760 problem that needs further studies for its resolution.

761 Can EOs also have any intracellular mechanisms of antibacterial action? It
762 seems so by evaluating results of a study on antimicrobial activity of *O. vulgare*, *T.*
763 *vulgaris* and *Eugenia caryophyllata* EOs against *Burkholderia cepacia* a complex,
764 opportunistic human pathogens highly resistant to antibiotics. [286] Evaluating the MIC of
765 the EOs alone, with antibiotics or with efflux pump inhibitors, authors conclude that the
766 antimicrobial activity of the six EOs versus *B. cenocepacia* strains might rely on the
767 inactivation of different molecular targets located in the cell cytoplasm rather than at the
768 cell membrane level, even though the latter possibility could be detected at higher
769 concentrations. An intracellular mode of action is confirmed also in a study on the
770 antimicrobial activity of *T. daenensis* and *O. vulgare* EOs against fluoroquinolone-
771 resistant *Streptococcus pneumoniae* clinical isolates [287] in which the tested EOs have a

772 total or partial synergistic effects with ciprofloxacin and ethidium bromide in all strains.
773 Furthermore, MIC/2 concentration of *T. daenensis* and *O. vulgare* EOs caused a
774 significant downregulation of efflux pump gene (*pmrA*) in seven of eight strains. A
775 further confirmation of the potential plural mode of action of the EO come from a study
776 on *S. sclarea* EO on seven pathogens bacterial strains: it seems that the antimicrobial way
777 of action involves a series of events both on the cell surface and within the cytoplasm.
778 ^[288] Many concerns are directed to bacterial species capable of forming biofilms, an
779 aggravating factor for the difficult eradication of the bacterial colonization in surfaces
780 (especially in the hospitals and food industries) and human organs (lung for example).
781 Prevention of biofilm formation and anti pre-formed biofilm activity of oregano essential
782 oils was evaluated mainly against *S. aureus* and *P. aeruginosa*, ^[289,290] with the result that
783 the tested EOs are active in preventing biofilm formation and are effective agents to
784 remove young and mature *S. aureus* biofilms on stainless steel surfaces. Both *O. vulgare*
785 and *S. officinalis* EOs acts as a potent anti-biofilm agent with dual actions, preventing and
786 eradicating the biofilm of *Streptococcus pyogenes* suggesting that these oils can be used
787 in developing potential plant-derived antimicrobial agents in the management of
788 streptococcal pharyngitis. ^[291] *T. vulgaris* EO is effective against planktonic of *Bacillus*
789 *cereus* with a significant inhibitory effect on biofilm formation ^[292] and was found to have
790 a high biofilm eradication ability, causing eradication that ranged from 80.1 to 98.0% at
791 10 $\mu\text{L}/\text{mL}$ against pathogenic *Klebsiella pneumoniae*. ^[293] *O. majorana* and *T. vulgaris*
792 EOs were assessed for their antibiofilm activity against immature and mature biofilms of
793 *E. coli* and *L. monocytogenes* formed on polypropylene surfaces, using the Response
794 Surface Box-Behnken Design to optimize concentration of essential oils, disinfection
795 time and level of pH in the EO-based disinfection solutions. ^[294] In this study the
796 disinfectant effect of the EO-based natural solutions was in most cases equivalent or better
797 compared to the peracetic acid-based chemical sanitizer used in food industry or to
798 sodium hypochlorite.

799 The interest on antimicrobial activity of EOs is still high not only for
800 humans/animals pathogens but also for plants pathogens. An *in-vitro* and *in-planta*
801 pathogenic study has been conducted on *O. compactum* and *T. vulgaris* EOs against
802 *Allorhizobium vitis*. The EOs show an interesting MIC, while the *in-planta* experiment
803 conducted with a 1:1 mixture of the two oils at the MIC concentration inoculated into the

804 injured stem of a tomato plant and a grapevine reduced both the number of plants
805 developing gall symptoms and the size of the tumors. ^[295] *O. rotundifolium* EO was tested
806 against 20 plant pathogenic strains showing a MIC in the range of 7.81 - 62.5 $\mu\text{L}/\text{mL}$. ^[296]
807 *O. vulgare* and *T. vulgaris* EOs were tested against *Pseudomonas syringae* showing a
808 MIC from 5.8 to 11.6 mg/mL and 1.43 to 11.5 mg/mL respectively with inhibition of
809 biofilm formation in several strains. ^[297] Finally the potential use of essential oils in the
810 field of the control of oral pathogenic bacteria as active principles of toothpastes and
811 mouthwashes is still of interest. The essential oils of four *Thymus* species in vitro
812 exhibited an antibacterial activity against *S. mutans* and the effects were also confirmed
813 by an interesting in situ method. ^[298] A higher antibacterial activity was showed by herbal
814 toothpastes after the addition of *O. dubium* EO against *S. mutans*. ^[299] A similar
815 enhancement of activity against oral pathogens was observed after the addition of a
816 mixture of EOs (*O. vulgare* L. ssp. *hirtum*, *L. nobilis* L., *R. officinalis* L., *S. fruticosa*
817 Mill.) in a percentage between 4.5-9.0% on a mouthwash formulation. ^[300] As per as
818 antibacterial activity of the EOs, their antifungal activity is largely studied still now. A
819 lot of them are *in-vitro* studies with *Candida albicans* as target strain. *C. albicans* is an
820 important opportunistic pathogen, responsible for the majority of yeast infections in
821 humans. In a study of 2016, 12 EOs (oregano and rosemary among the others) were tested
822 against 30 different vaginal isolated strains of *C. albicans* and the results compared with
823 those of three main used drugs (clotrimazole, fluconazole and itraconazole). ^[301] Results
824 show that oregano essential oils inhibited both the growth and the activity of *C. albicans*
825 more efficiently than clotrimazole and damages induced by essential oils at the cellular
826 level were stronger than those caused by the reference drug. *Lavandula binadulensis* EO
827 alone or in combination with *Cuminum cyminum* EO shows an interesting antifungal
828 activity against 20 clinical isolates from patients with recurrent vulvovaginal candidiasis,
829 ^[302] while in another study *L. angustifolia* EO show a similar behavior against 80 clinical
830 isolates with an additional antibiofilm activity against 1 of them. ^[303] In a huge work of
831 Gucwa and collaborators several EOs including *T. vulgaris* were tested against 183
832 clinical isolates of *C. albicans* and 76 of *C. glabrata*. The authors also tried to evaluate
833 the mode of action of the EOs concluding that three of the tested EOs (thyme, lemon, and
834 clove) affected cell membranes and all of the tested oils demonstrated the ability to inhibit
835 the transition of yeast to mycelium form. ^[304] An interesting study was recently conducted

836 by mean of electron paramagnetic resonance (EPR) spectroscopy to evaluate the effect of
837 clary sage oil and its main components, linalool and linalyl acetate, on the plasma
838 membrane of *Candida albicans* [305] confirming that clary sage oil causes membrane
839 perturbations which leads to cell apoptosis process. Recently two studies have been
840 published on the antifungal activity of oregano and thyme EOs [306] and their main
841 compounds thymol and carvacrol [307] against *Cryptococcus neoformans* an emerging and
842 recalcitrant systemic infection occurring in immunocompromised patients and are
843 occasionally being a problem for immunocompetent patients. Their findings highlight the
844 potential EOs effectiveness as natural and cost-effective adjuvants to counteract
845 *Cryptococcus infections* and inhibition of *Cryptococcus* biofilms.

846 Use of EOs as natural and safer antifungal agents for fruits, [308,309] vegetables
847 and mushrooms [310] also during their postharvest operations [311] it is a very current option
848 and of great interest. According to the study of Bill and collaborators [312] thyme essential
849 oil vapors reduced the anthracnose disease incidence by up-regulation of phenylalanine
850 ammonia-lyase (PAL) gene expression and at the same time led to down-regulation of
851 lipoxygenase (LOX) gene expression of two Avocado fruits variety prior to cold storage.
852 Because all these components are involved in the response to fungal infection, the
853 exposure to thyme oil vapors prior to cold storage is a promising alternative to the use of
854 commercial fungicide. The inhibitory effect of vapor phase of eight EOs from lamiaceae
855 species against seven strains of *Penicillium commune* isolated from moldy milk products
856 was assessed by Tančinová and collaborators [313] founding that essential oils have
857 different effects on individual strains of *P. commune*, and thyme was most effective
858 among the tested essential oil. The potentiality of rosemary EO against two strains of
859 *Fusarium graminearum* isolated from infected cereals was tested founding that the tested
860 one, among others oils, reduce the rate of mycelial growth and showed fungistatic
861 activity. [314] Finally, EOs can be used as alternative to chemical fungicides also in
862 floriculture as showed in a study on the effectiveness of oregano and clove essential oils
863 against *Podosphaera pannosa* and *Botrytis cinerea* affecting two cut rose cultivars in
864 greenhouse. [315]

865 2.6 Uses of EOs in the food industries

866 The modern food industry is increasingly oriented towards the production of minimally
867 processed and ready-to-eat products with a particular focus on practices that can

868 guarantee extended refrigerated storage periods. In this logic the major problems are
869 related to lipid oxidation (especially of polyunsaturated fatty acids) and microbial
870 contamination especially for those highly perishable meat and fish products. Nowadays
871 synthetic food additives have been developed to carry out the antioxidant or antimicrobial
872 action remaining safe for public health, cheap and with minimal impact on the sensory
873 qualities of food. Furthermore, improved modern packaging methods have been
874 developed with the use of edible and active coatings or with the use of modified
875 atmosphere packaging techniques. In this context, essential oils can play an important
876 role thanks to their antioxidant ^[316] and antimicrobial properties, replacing all or part of
877 the synthetic additives to limit both oxidative phenomena and microbial contamination,
878 prolonging, in some cases, the shelf-life of the more perishable products. Up to now some
879 limitations to the use of EOs with this purposes are emerged. First of all the interactions
880 with other food ingredients, which can limit biological activity of the EO. Furthermore
881 compared with synthetic antioxidants EOs are sometimes less effective and more
882 expensive. ^[317] Finally, the EOs can negatively affect the sensory quality of food and for
883 this reason many studies have been aimed at the possibility of resorting to
884 nanoformulations capable of mitigating the organoleptic impact of EOs. Despite these
885 limitations, studies on the application and effectiveness of EOs in the food industry are
886 continuing and the most studied oils are those of oregano, rosemary and thyme. ^[318] The
887 main methods through which the biological properties of EOs are transferred to food
888 products are the addition to animal feed (which we have already discussed in paragraph
889 2.4), direct addition to the food product during production and incorporation into food
890 edible films or coatings.

891 Oregano essential oil significantly reduced ($p < 0.05$) lipid and protein oxidation,
892 and improved color stability of raw and cooked ground chicken breast meat, showing the
893 strongest effect for all parameters at 400 ppm. ^[319] A similar study has been conducted
894 with *O. syriacum* L. EO showing 150 ppm as effective dose to delay lipid and protein
895 oxidation. ^[320] The effects of homogenization of oregano, sage and rosemary EOs with
896 minced beef (2% w/w) with respect to lipid oxidation and color properties during
897 refrigerated storage were evaluated by Unal et al. ^[321] Thyme and especially oregano EOs
898 show capacity of suppress biogenic amines formation in refrigerated vacuum packed
899 fillets of carp prolonging their shelf-life up to five times than suggested by the

900 manufacturers.^[322] In order to extend the shelf-life of food products, the microbial growth
901 is another aspect that must be strongly taken into consideration. EOs are recently
902 considered as powerful antimicrobial agent for food applications as evidenced by the
903 many studies carried out in recent years on the use of oils directly applied on food, on
904 their use as part of active packaging/coating and also in combination with modified
905 atmosphere packaging techniques. The addition of oregano essential oil (0.4% v/w)
906 increased lag phase and decreased growth rates of lactic acid bacteria natural microbiota
907 under isothermal conditions extending shelf-life of vacuum-packed cooked sliced ham.
908 ^[323] Rosemary EO showed interesting antibacterial activity in several food model
909 experiments conducted on thermally processed chicken meat,^[324] ready to eat vegetables
910 ^[325] and fish.^[326] Antimicrobial edible coatings or films acting as a protective barrier can
911 be used to retard food spoilage, thus extending food shelf life. EOs can be active part of
912 the coating as demonstrated by several studies conducted with alginate-based coating on
913 fish or fruits^[327,328] and on cooked beef with the confirmation of a high consumers'
914 acceptability. Caseinate-based coating with rosemary oil (0.5%) together with gamma
915 irradiation was evaluated on fish with the result of reduced counts of Enterobacteriaceae,
916 *Staphylococcus aureus*, and *Bacillus cereus*, *Vibrio* spp. and *Salmonella* spp with no
917 adverse effects on the sensory properties.^[329] Another way to maintain food quality,
918 freshness and safety of minimally processed food or ready-to-eat fresh food is the use of
919 active packaging. These materials can be made by using polymers with antimicrobial
920 activity themselves or by means direct incorporation of the antimicrobial active
921 ingredients into the polymer. EOs have demonstrated the ability to be incorporated into
922 these materials, becoming useful tools for reducing microbial counts and increasing the
923 shelf-life of food products. Several studies have been conducted in this field sometimes
924 taking into account also the final acceptance of consumers, which remains a fundamental
925 variable for the large-scale application of these materials. Incorporation of oregano EO
926 into polypropylene film gave a material with similar thermal and mechanical properties
927 of polypropylene with a higher antimicrobial activity in a food model of ready-to-eat salad
928 but with 'negative impact on consumers' purchase intentions.^[330] Rosemary oil was
929 incorporated on a packaging film of bilayer structure based on low-density polyethylene
930 for maintaining the freshness and extending the shelf life of packaged shrimps up to 4
931 days.^[331] A polyvinyl alcohol film containing oregano essential oil as an active packaging

932 application for fresh tomatoes without significant changes in the hardness, weight, and
933 color of the product during storage was developed. ^[332] In addition, biodegradable
934 packaging materials with different percentages of *Origanum* EOs content, which show
935 antimicrobial activity maintaining their biodegradability, can be developed as
936 demonstrated by Ketkaew and collaborators. ^[333]

937 EOs can be used in the food industry also in synergy with irradiation and as
938 reinforcement of modified atmosphere packaging techniques. The direct effect of gamma
939 irradiation on EOs is up to now subject of discussion and some data on the impact of this
940 practice on chemical composition and organoleptic descriptors was already published
941 ^[334,335] and it seems that the irradiation-induced changes in the quality depend on
942 atmosphere during the treatment. However, the combination of irradiation and coating
943 containing EOs is an effective strategy to reduce microbial count in food with minimal
944 impact on sensory properties which is worth investigating in the coming years.

945 The use of EOs as antimicrobial reinforcement of modified atmosphere
946 techniques is an innovative approach to manage microbial contamination of food.
947 Exploiting the synergy between the antimicrobial activity of the essential oil in a
948 packaging atmosphere suitably modified to reduce microbial growth is an interesting
949 approach because it is relatively simple but does not completely resolve the limits already
950 highlighted for the previous methodologies. The modified atmosphere packaging with
951 EOs system could be efficiently used to control the growth of the pathogen when strict
952 temperature control is difficult during the whole shelf life of the product as demonstrated
953 by Mahgoub and collaborators on a ready-to eat product inoculated with *Listeria*
954 *monocytogenes*. ^[336] Boskovic and collaborators ^[337] with the limitation of the negative
955 effect on sensory properties of the treated food successfully reported the synergistic effect
956 of modified atmosphere and thyme EO against four serovars of *Salmonella* inoculated in
957 minced pork meat. In fact, despite an optimal concentration of 0.9% of EO to obtain the
958 higher antimicrobial activity in combination with the modified atmosphere conditions
959 adopted in the study, only the 0.3 % of EO was considered acceptable by panelists.

960 *2.7 EOs and human health*

961 Since the dawn of modern medicine, essential oils have always been considered as
962 phytocomplexes useful for improving the health of human beings and for maintaining or
963 restoring a state of well-being. Over the centuries, it has gone from magic and superstition

964 to a consolidated use in traditional and popular medicine up to the present day in which
965 the greater awareness about the composition and biological properties of essential oils
966 make them increasingly part of modern alternative and complementary medicine. Surely,
967 this is due to their broad-spectrum antimicrobial activity that in a new era of bacterial
968 antibiotic resistance to synthetic antibiotics puts them at the center of an intense scientific
969 research in order to be able to use them alone or in combination with commercial drugs
970 for a tighter fight to this new health emergency. But the interest in essential oils has also
971 shifted over the years towards their potential use as antiviral and antitumoral agents, not
972 forgetting the growing interest in their interaction with the central nervous system and
973 more generally with the controversial aromatherapy. Unfortunately, this has led to the
974 production of a disproportionate number of *in-vitro* studies that although confirm the
975 potential applicability of these complexes in the human health field, have not translated
976 into adequate *in-vivo* studies able to generate clinical data that confirm the efficacy and
977 selectivity of these substances against human diseases. There are technological limits that
978 have not yet been overcome, even if they have held back the development of these studies.
979 First and foremost, the difficulty of conveying these substances of such a lipophilic nature
980 in aqueous media, a problem that is being faced by resorting to nanoformulations as
981 previously mentioned. However, the main constituents of these essential oils are terpenes
982 and terpenoids, which are small, lipid soluble organic molecules that can be absorbed,
983 through the skin, across nasal mucosa or after ingestion, into the systemic blood
984 circulation producing a systemic effect. Many of them can also cross the blood-brain
985 barrier.

986 Secondly, the chemical variability of these phytocomplexes that strongly
987 depends on both anthropic and non-anthropic factors. The compositional variability of
988 EOs that often makes it difficult to rationalize the biological activity and discourage from
989 a deepening about their pharmaceutical development. So, the real challenge for these ‘old
990 guys’ is to overcome the above mentioned limitations starting to keep what we have
991 largely promised. All this being stated and considered below we will try to trace a brief
992 state of the art and the most interesting perspectives on the use of essential oils for human
993 health.

994 Literature data on *in-vitro* cytotoxic activity of EOs are huge and an exhaustive
995 discussion is out of the objectives of this review. A selection of recent publications is
996 summarized in **Table 8**.

997 What we find most interesting to point out is rather the most recent data on the
998 mechanisms of cytotoxic action of essential oils. Up to now several different mechanisms
999 were hypothesized for their cytotoxic effects. These include mainly the induction of cell
1000 death by apoptosis and/or necrosis and cell cycle arrest. Similarly, to what has been
1001 demonstrated regarding the mechanism of antimicrobial action of essential oils, the
1002 lipophilic nature and low molecular weight of many of their components allows the
1003 passage through the cell membrane. Once destabilized, it is no longer able to play its role
1004 as a barrier and some of the cytoplasmic content can come out. The physiological
1005 consequences of a profoundly altered cellular membrane are many and among these the
1006 reduced production of ATP, the alteration of the pH and the loss of the mitochondrial
1007 potential can be mentioned. ^[338] An *in-vitro* and *in-vivo* study on the effect of *L.*
1008 *angustifolia* EO on human prostate cancer cells PC-3 and DU145 demonstrated that this
1009 oil is effective in inhibiting tumor growth of human prostate cancer xenografts in nude
1010 mice. The use, during the study, of the two main components of the oil linalool and linalyl
1011 acetate allow to address the main contribution to the anticancer activity to the former one.
1012 ^[339] A less common EO of lavender (*L. dentata*) was assayed on lung cancer cells Calu-
1013 3 in an *in-vitro* model which include both liquid and vapor phases. The oil showed
1014 cytotoxic activity in the vapor phase acting by necrotic and apoptotic processes not
1015 involving inhibition of P-glycoprotein. ^[340]

1016 Looking at published data several EOs show antiviral activity against many
1017 enveloped RNA and DNA viruses. Among our five EOs, thyme and oregano seems to
1018 have more consideration in this field. ^[341] Only few essential oils, e.g. oregano oil, were
1019 also tested against non-enveloped RNA and DNA viruses. ^[342] Oregano oil and one of its
1020 main active component, carvacrol, was assayed against the non enveloped murine
1021 norovirus (MNV), a human norovirus surrogate, demonstrating that carvacrol is effective
1022 in inactivating MNV within 1 h of exposure by acting directly on the viral capsid and
1023 subsequently the RNA. ^[343]

1024 In a recent screening *in-vitro* study, majoram and clary sage EOs among others
1025 exhibited strong anti-influenza A/WS/33 virus activity. Due the relevant presence of

1026 linalool in their chemical composition, the antiviral activity appeared to be associated
1027 with this compound. [344]

1028 *Thymus vulgaris* and *Rosmarinus officinalis* EOs were studied for the first time
1029 for its possible interference with the Tat/TAR-RNA interaction and with Tat-induced
1030 HIV-1 LTR transcription with the result that EOs interfered with Tat function, a feature
1031 that was never described before, [345] while *Salvia desoleana* EO and its less polar fraction
1032 were here shown to be active against herpes simplex virus HSV-2 inhibiting a step of its
1033 replicative cycle that occurs after virus attachment and entry. [346]

1034 Among natural products, essential oils present promising results in the main
1035 mechanisms involved in the pathology of inflammation and recently de Lavor and
1036 collaborators [347] have published an excellent review of their potential use in this field.
1037 In their conclusion authors declare that the main action targets of the EOs for the therapy
1038 of chronic inflammations were the reduction in reactive oxygen and nitrogen species and
1039 the reduction in NF- κ B (nuclear factor kappa-light-enhancer of activated B cells). The
1040 anti-inflammatory and immune-modulating activities of essential oils and their bioactive
1041 compounds was also linked to cytokine expression showing their potential therapeutic
1042 activity for the treatment of asthma. [348] Specifically, for our five essential oils, the
1043 antinflammatory activity has been recently reported for oregano essential oil on
1044 lipopolysaccharide (LPS)-treated murine macrophage cells. Oregano EO protects against
1045 the LPS-induced cell inflammatory response through the NADPH oxidase/ROS pathway.
1046 [349] Similarly the results of the anti-inflammatory tests on sage essential oils using murine
1047 macrophages indicate that these oils significantly ($p < 0.05$) reduced nitric oxide (NO),
1048 and NF- κ B production in murine machophages cells. [350] The antinflammatory activity
1049 of rosemary EO is mainly attributed to its main compounds 1,8-cineole, α -pinene, and
1050 camphor. The anti-inflammatory properties traditionally ascribed to *Thymus carnosus* and
1051 *Thymus camphoratus* was recently demonstrated in a study showing inhibitory effects
1052 towards nitric oxide production and concomitantly inhibiting the expression of two
1053 crucial pro-inflammatory proteins, iNOS and COX-2 in lipopolysaccharide (LPS)-
1054 stimulated macrophages. [351] As per as the antinociceptive activity of *Thymus capitatus*
1055 essential oil, which acts via peripheral nervous excitability blockade. [352]

1056 In the last few years, essential oil components have been evaluated for their
1057 mechanism of action and also to obtain leading compounds active in central nervous

1058 system (CNS). Linalool alone as well as the *L. angustifolia* essential oils affect, in a
1059 concentration-dependent manner, neural firing activity. [353] *L. angustifolia* EO exerts
1060 receptor binding affinities with a relevant activity on the N-methyl-D-aspartate receptor
1061 and inhibition of the serotonin transporter. [354] Inhalation and the external application of
1062 these oils for the treatment of mental and physical balance are the very basics of
1063 aromatherapy in which olfactory nerves from nose to the brain are the site of action for
1064 these essential oils. [355] The sense of smell plays an important role in the physiological
1065 effects of mood, stress, and working capacity. Electrophysiological studies have revealed
1066 that various fragrances affected spontaneous brain activities and cognitive functions. In
1067 addition, the electroencephalograph studies clearly revealed that fragrances significantly
1068 modulate the activities of different brain waves and are responsible for various states of
1069 the brain. [356] Aromatherapy massage with lavender oil increased the sleep quality and
1070 reduced the level of anxiety in patients with colorectal surgery in the preoperative period,
1071 [357] may reduce the incidence of activities of daily living disability in patients with
1072 osteoarthritis of the knee, [358] inhalation aromatherapy can decrease the level of fatigue
1073 in the patients undergoing hemodialysis [359] and reduced anxiety and pain suffered during
1074 gynecological examination. [360] Rosemary EO inhalation has therapeutic potential against
1075 stress-related psychiatric disorders [361] and may improve cognitive function in a model
1076 of Alzheimer's type dementia. [362]

1077 **INSERT TABLE 8**

1078 *2.8 Cultural heritage preservation*

1079 Cultural heritage is daily attacked by biodeteriogens (fungi and bacteria) which can cause
1080 several damages to the materials. All types of materials are subject to biodeterioration
1081 from paper, stone materials to paintings. These deteriorations can be chromatic
1082 (browning, loss of color, alteration of color due to cellular exudates) or structural
1083 (leaching of salts due to rain for example). In some cases, such as molds in historic homes,
1084 they can also pose a danger to human health. The use of synthetic products with antifungal
1085 and antibacterial activity is today an established practice, but scientific research is
1086 increasingly interested in the evaluation and use in this field of natural substances that are
1087 effective, ecologically sustainable and safe for cultural heritage and for operators.
1088 Essential oils (and hydrolates) seem to have all the requirements to be used in this field.
1089 Their broad-spectrum of antimicrobial activity, their safety and their sustainability make

1090 them ideal candidates for the replacement of synthetic products in some bioremediation
1091 applications. The antifungal activity of *O. vulgare*, *R. officinalis* and *L. angustifolia*
1092 essential oils was investigated against fungi isolated from stone (*Bipolaris spicifera* and
1093 *Epicoccum nigrum*) and wooden substrata (*Aspergillus niger*, *Aspergillus ochraceus*,
1094 *Penicillium sp.* and *Trichoderma viride*) of cultural heritage objects. Antifungal potential
1095 of *O. vulgare* was higher or nearly the same as benzalkonium chloride. ^[376] Thyme EO
1096 was the most effective, among others, against 16 fungal species isolated from three tested
1097 archaeological objects (wall painting stone, wooden statue, and pottery coffin) from
1098 Saqqara in Egypt. ^[377] Antimicrobial activity of *T. vulgaris* and *O. vulgare* EOs, have
1099 been evaluated in-vitro, ex-situ and in-situ for their antimicrobial activity for the mosaic
1100 tesserae of “Casa di Leda” in the Greco - Roman site of Solunto in Sicily. ^[378] *T.*
1101 *pulegioides* among others EOs was evaluated as antifungal agent in water-damaged
1102 buildings of Vilnius Old City, ^[379] as per as *T. vulgaris* and *T. serpyllum* as antifungal
1103 against four mould types isolated from building surfaces. ^[380] Antifungal activity of *O.*
1104 *vulgare* essential oil as a was tested against fungi causing alterations on the frescoes and
1105 facade of the Holy Virgin Church of Gradac Monastery in Serbia highly colonised with
1106 cyanobacteria, algae, fungi, lichens, mosses and even higher plants in a long-term
1107 ecological succession. ^[381]

1108 Oregano oil not only was effective against fungi but also prevented their
1109 sporulation of them showing in addition a stronger antibacterial activity in a study against
1110 4 strains of fungi and 6 bacterial strains isolated from documents of museal archives. ^[382]
1111 *O. vulgare* L. and *T. vulgaris* L. had antifungal activity against *Scopulariopsis sp.* and
1112 *Fusarium sp.* isolated from documentary heritage. ^[383] The effectiveness of disinfection
1113 method with thyme essential oil microatmosphere was compared with silver nanoparticles
1114 misting (AgNPs), and low temperature plasma (LTP) by means of culture-dependent
1115 method and RNA analysis on paper from historical books with different levels of
1116 microbial contamination. ^[384] Finally, the combined use of gellan and hydrolates seems
1117 to be a very promising and eco-friendly system to clean paper artworks. ^[385]

1118 2.9 Nanotechnologies at the service of essential oils

1119 In the preceding paragraphs we have repeatedly referred to how the biological activities
1120 of essential oils suffer the negative effects of hydrophobia and the excessive volatility of
1121 these substances. These obstacles can be partly overcome by the use of nanotechnologies,

1122 a generic term that includes, in the case of essential oils, nanoemulsions and
1123 nanocapsules. These techniques allowing a greater and more uniform distribution of the
1124 essential oil in the aqueous medium creates the best conditions for the increase of the
1125 contact surface with the target substrates indispensable for the performance of the
1126 biological activity. Nano-encapsulations also have the ability to reduce the volatility of
1127 EOs by increasing their persistence and reducing their odor, which is an important factor
1128 for application in the food sector. Nanoemulsion of EOs is cheap, readily scalable and
1129 environment-friendly and comprises mainly three constituents oil phase, aqueous and a
1130 surfactant. The size of nanoemulsion droplets ranges from 10 to 100 nm. The surfactant
1131 chosen must be able to lower interfacial tension to a very small value to assist dispersion
1132 process during the preparation of the nanoemulsion. ^[386] Its concentration affect
1133 emulsions' separation, viscosity, electrical charge (Z-potential) of droplets, and droplet
1134 size of the dispersed phase. Several studies demonstrate that low temperatures and long
1135 times of homogenization produced systems with small droplet sizes, which promoted the
1136 stability of emulsions. ^[387] Recent comparative studies on antimicrobial efficacy against
1137 food-borne bacteria between sage EOs and their nanoformulation with nonionic
1138 surfactants demonstrate that the nanoemulsion was up to four-times higher than the bulk
1139 oil, more rapid in its action causing extensive bacterial cell membrane damage. ^[388,389]
1140 Droplet growth can be retarded by incorporating water-insoluble oils, known as ripening
1141 inhibitors, into the oil phase prior to nanoemulsion formation. The effect of some of these
1142 ripening inhibitors on antimicrobial activity of thyme oil nanoemulsion was evaluated by
1143 Ryu and collaborators ^[390] establishing the optimum amount of ripening inhibitor (around
1144 40% of the oil phase) required to maintain antimicrobial activity.

1145 As food consumers are highly interested in products containing more natural
1146 ingredients, also surfactant could be chosen among natural compounds as highlighted by
1147 Doost and collaborators. ^[391] In their study Quillaja Saponin was used as a natural
1148 surfactant and sucrose monopalmitate (SMP) and octyl modified starch (OMS) as non-
1149 petroleum-based ingredients to create a nanoemulsion with *O. compactum* EO. An
1150 emulsifier derived from wheat waste (alkyl polyepentoside) was successfully used with
1151 rosemary EO obtaining monodispersed concentrated emulsions with high physical
1152 stability and nanometric sizes ^[392] and with thyme EO. ^[393] Whey protein isolate was used
1153 by Rashed et al., ^[394] as a natural-biodegradable polymer wall material to formulate and

1154 stabilize Oil-in-Water nanoemulsion of *L. angustifolia* EO with a higher thermal stability.
1155 Another example of the use of ecological friendly materials is the nanoencapsulation of
1156 oregano EO into ultrathin fibers derived from fruit waste using solution electrospinning
1157 in order to obtain films for food packaging with enhanced antimicrobial activity. [395]

1158 The biological activities of EOs nanoemulsion could be further increased through
1159 a co-formulation with synthetic active principles. This approach could be useful mainly
1160 in pharmaceutical fields in order to reduce the quantity of the active principle through a
1161 synergistic action with the EO reducing side-effects. [396] Mainly in the pharmaceutical
1162 field the complexation of EOs with cyclodextrins is another way of solving the problems
1163 by improving the solubility and stability in water and avoiding the degradation and
1164 volatilization of bioactive compounds in the EOs. An excellent review of the *in vitro* and
1165 preclinical studies of EOs complexed with different types of cyclodextrins was recently
1166 published by de Oliveira-Filho and collaborators. [397] They conclude that both *in vitro*
1167 studies and preclinical studies gave evidence that cyclodextrin complexation is able to
1168 improve the pharmacological activity of EOs in various animal models with improved
1169 efficacy, reducing therapeutic doses and side effects without showing toxicity. Lavender
1170 EO encapsulated in hydroxypropyl-beta-cyclodextrin increase its antimicrobial activity
1171 up-to three times. [398]

1172 Biological activities of EOs (I.e. antifungal) could be significantly enhanced
1173 loading them in chitosan nanocapsules also with the aid of innovative approaches as
1174 electrospinning technique, [399] via ionic gelification reaction [400] or in chitosan nanofibers
1175 by means of coaxial electrospinning. [401] As per as loading EOs on nanocapsules with
1176 biologically active and recognized as safe for human health metal elements (such as
1177 copper). [402].

1178 The delivery of EOs could be optimized also by their encapsulation on
1179 nanostructured lipid carriers prepared by phase inversion temperature and high pressure
1180 homogenization methods, [403] on liposomes prepared using the film hydration method
1181 [404] or on nanocochleates based on phosphatidylcholine, cholesterol and calcium ions [405]
1182 with interesting increase of their effectiveness and stability. Nanoarchaeosomes made of
1183 soybean phosphatidylcholine, polysorbate 80 and total polar archaeolipids extracted from
1184 archaeobacterial *Halorubrum tebenquichense* and loaded with thyme EO were

1185 successfully tested for their anti MRSA biofilm activity opening new ways for the
1186 exploitation of these kind of nanovesicles loaded with EOs. [406]

1187 Biodegradable packaging materials has been introduced as a promising solution
1188 for the problem of huge amounts of synthetic materials, which cause serious
1189 environmental problems. The incorporation of EOs within these materials is a new
1190 frontier on the field active packaging. Recently a lot of studies were published with poly
1191 (ϵ -caprolactone), [407] sodium alginate-montmorillonite, [408] soybean polysaccharides [409]
1192 and alginate/soy protein. [410]

1193 The sol-gel method to prepare hybrid organic–inorganic silica materials able to
1194 encapsulate EOs with a better-controlled release seems to be a promising way for the
1195 design of innovative and cheap antibacterial materials. [411,412]

1196 Can EOs based magnetic nanofluid be used in microwave assisted imaging of
1197 tissues? A promising study of Dzimitrowicz and collaborators [413] was conducted with *S.*
1198 *hispanica* EO as ‘size-definer’ for the synthesis of magnetic nanorods which might be
1199 used as a contrast agent for microwave-imaging of tissues. In the authors’ opinion
1200 terpenes-like compounds were likely involved in the green synthesis and surface
1201 functionalization of the magnetite nanorods which were susceptible to microwave-
1202 excitation.

1203 An innovative approach to exploit the antimicrobial activity of nanoformulated
1204 EOs could be the synthesis of antimicrobial textiles able to resist the growth of
1205 microorganisms and pathogens causing bad odours and hygienic problems. Thyme
1206 essential oil in the presence of second generation of polypropylene Imine dendrimer as a
1207 control release agent was investigated. [414] The odour release was measured by electronic
1208 nose and the antibacterial activity of the resulting materials was evaluated with
1209 encouraging results opening the door to a new generation of composite textile materials.

1210

1211 **3. Extracts from selected Lamiaceae herbs: biological activities** 1212 **discovered/assessed during years 2015-2019**

1213 Even if not as popular as essential oils, extracts from Lamiaceae herbs (mainly polar,
1214 obtained by maceration in alcoholic or hydro-alcoholic mixtures) have always raised
1215 interest at some extent, and recent literature confirms this trend. In this paragraph a small
1216 survey of the biological activities ascribable to lavender, oregano, rosemary, sage and

1217 thyme will be given, as examples from a simple research on one of the most used scientific
1218 databases (Scopus).

1219 3.1 Lavender

1220 Lavender (*Lavandula* spp.) aerial parts, either in the form of mostly essential oil or
1221 different forms of extracts have been traditionally used as therapeutics since ancient
1222 times. Nevertheless, data showing that the effects of lavender polar extracts are limited.
1223 In a study conducted over 10 years, Soheili and Salami ^[415] demonstrated that the aqueous
1224 extract of lavender improves the impaired learning and memory, positively affect the
1225 synaptic transmission in an animal model of Alzheimer's disease (AD), clears the amyloid
1226 beta (A β) plaques from the hippocampus of these animals, and dose dependently inhibits
1227 the formation of A β aggregate, caffeic acid (1) and luteolin (5) were characterized as
1228 components of the aqueous extract. Alexa and coworkers ^[416] investigated the *in vitro*
1229 antimicrobial effect against *Staphylococcus aureus* and the antiproliferative activity
1230 against two cancerous cell lines (A375 human melanoma and MDA-MB-231 breast
1231 carcinoma) of *Mentha piperita* L. and *Lavandula angustifolia* Mill. extracts prepared
1232 using "aromatic" water (obtained after hydrodistillation and separation of the
1233 corresponding essential oil) as extraction medium and apple vinegar to acidify the
1234 solution at pH 4. A mixture of hydroxycinnamic acids (HCAs) have been detected in both
1235 aromatic plants and in particular caffeic (1), *p*-coumaric (2), ferulic (3), and rosmarinic
1236 (4) acids. Unfortunately, the results obtained were only moderately positive. Regarding
1237 antioxidant activity, a lavender (*L. latifolia*) extract obtained by sequential cold
1238 maceration with dichloromethane, ethyl acetate and methanol was tested *in vitro* (DPPH,
1239 ABTS, FRAP, TPC assays) and *in vivo* (*Caenorhabditis elegans* model) together with
1240 extracts from *Melissa officinalis* and *Origanum vulgare* as potential antioxidants for
1241 functional food formulations, giving promising results. The novelty in this case is
1242 represented by the *in vivo* model used: the nematode *Caenorhabditis elegans*, which in
1243 contrast to cellular models allows to study whole organisms with many different organs
1244 and tissues and increases the chance of identifying synergistic and/or off-target effects.
1245 ^[417] As in the previous cases the main components were coumaric (2), ferulic (3) and
1246 rosmarinic (4) acids together with luteolin (5). Finally, methanolic extracts from several
1247 Portuguese lavenders (*Lavandula stoechas* ssp. *luisieri* and *L. pedunculata*) with well
1248 known antioxidant activity, being rich in polyphenols and in particular of flavonoids and

1249 rosmarinic acid (4), were selected for encapsulation in polymeric poly (lactic-co-glycolic)
1250 acid (PLGA) nanoparticles in order to improve their stability and provide a better
1251 efficiency for treatment of cutaneous diseases. The authors obtained promising results
1252 especially in term of low risks of toxicity of the produced nanoparticles. [418]

1253 3.2 *Oregano*

1254 The most recent findings on the biological activities ascribable to *O. vulgare* mainly deal
1255 with its anticancer, antibacterial and cytotoxic potential. As example Rubin and
1256 coworkers [419] analysed for the first time a crude extract of oregano in 2 tumour cell lines,
1257 SW13 and H295R cells, the only available for adrenocortical tumors, finding out that this
1258 extract showed promising effects. The anti-mycobacterial and anti-inflammatory
1259 activities of *Origanum vulgare* L. ssp. *hirtum* in innate immune cells were assessed by
1260 De Santis et al. [420] The authors run the experiments using eight different fractions of a
1261 hydroalcoholic extract (50% aqueous ethanol) from this plant; the results obtained
1262 showed that the capability to activate antimicrobial and anti-inflammatory response is
1263 shared by different fractions, suggesting that different molecules take part in the process
1264 and that oregano may be exploitable as phytocomplex for novel therapeutic approaches.
1265 Still on anti-mycobacterial and anti-mycotoxin properties of oregano, Ponzilacqua and
1266 co-authors [421] evaluate the ability of an aqueous extract from this herb for its ability to
1267 degrade aflatoxin B1 *in vitro*, together with extracts from sweet passion fruit (*Passiflora*
1268 *alata*), araçá (*Psidium cattleianum*) and rosemary. A significant decrease in AFB1 levels
1269 was observed in the aqueous extracts of all plants, with rosemary extract having the
1270 highest percentage of AFB1 reduction, followed by oregano. With these data the authors
1271 demonstrated that these plant extracts are interesting alternatives for the control of
1272 aflatoxins in food. Finally, recent applications of oregano involve veterinary research,
1273 where extracts/infusions of this herb were used as food supplement in the diet of cows in
1274 the transition period [422] and to fight avian infectious bronchitis virus with very promising
1275 results *in vitro*. [423]

1276 An exhaustive study of the chemical composition of a large sampling of Sicilian
1277 *O. vulgare* ssp. *hirtum* carried out some years ago allowed to establish the chemical
1278 composition of ethyl acetate and ethanol extracts: four flavanones [eriodictyol 7-*O*
1279 rutinoside (6), aromadendrin (7), eriodictyol (8) and naringenin (9)], seven flavones
1280 [luteolin (5), sorbifolin (10), cirsiol (11), apigenin (12), cirsilin (13), cirsimaritin (14)

1281 and xanthomycol (**15**), and two organic acids [(caffeic (**1**) and rosmarinic (**4**)] have been
1282 detected and quantified. The antioxidant effectiveness of the extracts, obtained with
1283 different methods (ABTS, DPPH, FRAP, ORAC, β -carotene bleaching test to quote the
1284 most applied), was quite significant and of course associate with the polyphenols content
1285 previously described. ^[59]

1286 *O. vulgare* is one of the main species of this genus and then the most studied,
1287 however, other minor species show interesting chemical composition associated with
1288 biological activities, for example a study of the chemical characterization and the
1289 evaluation of antimicrobial, enzyme inhibitory, and antioxidant activities of methanol and
1290 water extracts of Turkish *O. onites* has been recently reported. ^[424] In total 28 components
1291 belonging to polyphenols, flavonoids, terpenoids, organic acids and esters have been
1292 identified: 3-*O*-(4-coumaroyl)-quinic acid (**16**), acacetin (**17**), cirsimaritin (**14**),
1293 naringenin (**9**), tuberonic acid glucoside (**18**), ferulic acid (**3**), apigenin-7,4'-dimethyl
1294 ether (**19**), azelaic acid (**20**), baicalin (**21**), casticin (**22**), cirsiol (**11**), dihydrorobinetin
1295 (**23**), gardenin B (**24**), genkwanin (**25**), isoferulic acid (**26**), kaempferol (**27**), nevadensin
1296 (**28**), oroxylin A 7-*O*-glucuronide (**29**), quercetin (**30**), rosmarinic acid methyl ester (**31**),
1297 sakuranetin (**32**) and salvigenin (**33**).

1298 *O. dictamnus* collected in Greece showed a simpler polyphenol profile, ^[425] with
1299 vicenin-2 (**34**), apigenin 7-*O*-glucuronide (**35**), luteolin (**5**), chlorogenic (**36**) and
1300 rosmarinic acids (**4**) among the main components.

1301 3.3 Rosemary

1302 Among the herbs belonging to the Lamiaceae family, rosemary (*Rosmarinus officinalis*
1303 L.) is by far the one to which the highest number of biological activities have been
1304 ascribed. In the latest five years, the *in-vivo* hepato- and hepato-renal protective effects
1305 of rosemary have been extensively studied; as example Hegazy and coworkers ^[426]
1306 investigated the preventive effect of the aqueous extract of this herb on the gentamicin-
1307 induced hepatotoxicity in rats with promising result connecting these activities with the
1308 presence in the extract of catechin (**37**), coumarin (**38**), cinnamic acid (**39**), rutin (**40**),
1309 sinapic acid (**41**) and oleuropein (**42**), whilst Mohamed et al. ^[427] studied the protection
1310 of rosemary ethanolic extract on lead acetate hepato- and nephro-toxicity in male albino
1311 rabbits. The authors found that pre-treatment with rosemary extract significantly
1312 suppressed levels of hepatic and renal damage products as well as lipid peroxidation.

1313 Furthermore, the extract preserved blood cells and their structure and renal and hepatic
1314 architecture. An ethanolic extract from rosemary is also capable of attenuate pulmonary
1315 fibrosis induced by the use of the chemotherapeutic bleomycin, as reported by Hosseini
1316 et al. ^[428] in a recent review. Wen et al. ^[429] reported the synergic action of carnosic acid
1317 in combination with cisplatin in the treatment of mouse Lewis lung cancer. Carnosic acid,
1318 together with carnosol (43) and rosmanol (44), are diterpenoids belonging to the
1319 metabolic pool of rosemary, responsible for the majority of its biological activities. In a
1320 recent survey of Sicilian *R. officinalis* the ethyl acetate and ethanol extracts showed the
1321 presence of several polyphenols subdivided in organic acids: caffeic (1) and rosmarinic
1322 (4), flavonoids: luteolin (5), luteolin glucuronide (45), 6-hydroxy luteolin glucoside (46),
1323 scutellarin (47) and scutellarin 7-O-glucoside (48), cirsimaritin (14), genkwanin (25),
1324 salvigenin (33), and diterpenes: carnosol (43), carnosic acid (49) and methyl carnosate
1325 (50). Rosmarinic and carnosic acids were largely the main components contributing to
1326 the antioxidant properties of the extracts. ^[95] Another compound with recognized
1327 biological activities is rosmarinic acid (4), which can be considered as a sort of “family
1328 marker” for Lamiaceae. Rahbardar and co-workers ^[430] investigated the anti-
1329 inflammatory effects of an ethanolic extract from *R. officinalis* and rosmarinic acid (4) in
1330 a rat model of sciatic nerve chronic constriction injury (CCI)-induced neuropathic pain to
1331 verify usage of rosemary in folk medicine. Their data supported the traditional use of this
1332 herb as an effective remedy for pain relief and inflammatory disorders, and also suggested
1333 that the ethanolic extract of *R. officinalis* and rosmarinic acid might be potential
1334 candidates in treating different neurological disorders associated with inflammation.

1335 3.4 Sage

1336 *Salvia officinalis* L. (common sage) is a very popular herb frequently used in many
1337 countries as spice and as a folk remedy. Nevertheless, the molecular mechanism beyond
1338 sage extracts' biological activity has not yet been fully understood. Horváthová and
1339 coworkers ^[431] investigated the antioxidant potential of sage in rat liver cells in search for
1340 evidences of a hepatoprotective potential of this herb. The authors found out that, after
1341 only two weeks of drinking extracts, not only the level of DNA damage induced by
1342 oxidants was decreased, but sage extract was able to start up the antioxidant protection
1343 expressed by increased content of glutathione. More recently, El Gabbas et al. ^[432]
1344 evaluated the anxiolytic and antidepressant-like effects of a methanolic extract from

1345 *Salvia officinalis*; the same extract was also studied for its potential to enhance rat's
1346 memory using conditioned learning test. The administration of sage extract showed a
1347 significant anxiolytic effect in marble burying test. In the case of conditioned learning
1348 paradigm, memory enhancement was observed in sage treated group, which indicates a
1349 cognition improvement. These are to be ascribed to the phytochemicals characterized in
1350 the extract of *S. officinalis*, namely caffeic acid (1), *p*-coumaric acid (2), rutin (40),
1351 rosmarinic acid (4), quercetin (30), luteolin (5), apigenin (12), carnosol (43) and carnosic
1352 acid (49).

1353 3.5 Thyme

1354 The antibacterial potential of thyme is well recognized and documented; recently,
1355 Cheurfa and Allem ^[433] tested the aqueous and hydro-alcoholic extracts prepared from
1356 *Thymus vulgaris* and other vegetable matrices (*Aloysia triphylla*, *Pistacia lentiscus*, *Olea*
1357 *europaea* leaves and *Trigonella foenum-graecum* seeds) on a collection of pathogenic
1358 bacteria responsible for gastroenteritis, where some of them isolated from clinical
1359 samples (*Escherichia coli*, *Staphylococcus aureus*, *Citrobacter freundii*, *Pseudomonas*
1360 *aeruginosa*, *Proteus mirabilis*, *Proteus vulgaris* and *Salmonella typhimurium*), thus
1361 obtaining encouraging results against all the strains. The antidiabetic effect, measured as
1362 *in vitro* antiglycation activity (ability of the extracts to inhibit the methyl glyoxal mediated
1363 development of fluorescence of bovine serum albumin BSA) of a 80% acetone extract
1364 from *Thymus vulgaris* (thyme) was studied by Kazeem and others ^[434] obtaining excellent
1365 results as the extract of this herb had a high antiglycation effect with 50% inhibitory
1366 concentration (IC50) of 0.02 mg/mL. As already stated for essential oils (see previous
1367 paragraphs), the use of nano-constructs can modulate and even ameliorate the
1368 bioactivities of vegetable matrices; Proks and coworkers ^[435] conducted an experiment
1369 on 4 different cancer cell lines: non-melanoma skin cancer (A431 cell line), melanoma
1370 skin cancer (A375 cell line), hormone dependent breast cancer (MCF-7 cell line), and non-
1371 hormone dependent breast cancer (MDA-MB-231 cell line). The authors compared
1372 anticancer activities of thyme aqueous extract alone and silver nanoparticles prepared
1373 with thyme aqueous extract. The extract tested at various concentrations (10-1000
1374 mg/mL) exhibited a pronounced effect only on MCF-7 cells whilst the biosynthesized
1375 silver nanoparticles the effect can be noted even at 1 M, thus indicating a pronounced
1376 effect of silver nanoparticles, in a dose-dependent manner, on the viability of hormone

1377 dependent breast cancer cells. The composition of the tested *T. vulgaris* aqueous extracts
1378 revealed the presence of several polyphenols: *p*-hydroxybenzoic (51), caffeic (1)
1379 rosmarinic (4) acids, catechin (37), eriocitrin (6), luteolin 7-*O*-rutinoside (52), luteolin 7-
1380 *O*-glucuronide (45), luteolin (5), apigenin 7-*O*-glucoside (53), apigenin (12), quercetin
1381 (30) and phloridzin (54).

1382 As regarding uncommon thyme varieties, Khouya and others [436] evaluated the
1383 anti-inflammatory, anticoagulant and antioxidant effects of aqueous extracts from
1384 *Thymus atlanticus*, *T. satureioides* and *T. zygis*. All varieties were found to possess
1385 considerable antioxidant activity and potent anti-inflammatory activity. Administration
1386 of aqueous extracts of two varieties (*T. zygis* and *T. atlanticus*) reduced significantly the
1387 carrageenan-induced paw edema similar to non-steroidal anti-inflammatory drug
1388 (indomethacin). In partial thromboplastin time and prothrombin time tests, *T. atlanticus*
1389 and *T. zygis* extracts showed the strongest anticoagulant activity, while *T. satureioides*
1390 did not show the anticoagulant activity in these tests. The observed activities of the
1391 aforesaid thyme species can be ascribed to the presence (in the aqueous extracts) of
1392 caffeic (1) and rosmarinic acids (4), quercetin (30), hesperitin (55) and luteolin-7-*O*-
1393 glycoside (56).

1394 In south Italy and in particular in Sicily *Thymus capitatus* (syn. *Coridothymus*
1395 *capitatus* and *Thymbra capitata*) is the main wild species belonging to this genus. Also
1396 in this case the ethyl acetate and ethanol extracts of several samples collected in the
1397 Sicilian territory showed the presence of several polyphenols: caffeic (1) and rosmarinic
1398 (4) acids were the main organic acids, all others components were flavonoid derivatives:
1399 vicenin-2 (34), luteolin rutinoside (53), 6-hydroxy luteolin (57), 6-hydroxy luteolin
1400 glucoside (46), taxifolin di-*O*-glucoside (58), apigenin (12), apigenin 7-*O*-rutinoside (59),
1401 apigenin 7-*O*-glucuronide (35), gardenin B glucoside (60), eriodictyol (8), genkwanin
1402 (25), thymusin (61), naringenin (9), cirsilineol (13), and chrysoeriol (62). [145]

1403 **Figure 8** shows the molecular formulas of the currently known Lamiaceae
1404 components, whereas **Figure 9** reports some typical HPLC profiles of the extracts of
1405 some Lamiaceae

1406

INSERT FIGURE 8

1407

INSERT FIGURE 9

1408 **4. Extracts from *Lamiaceae* in the food preservation sector: food** 1409 **additives, edible coatings & films, active and intelligent packaging**

1410 *4.1 Background*

1411 The quality and acceptability of food products is seriously limited by oxidation
1412 phenomena, usually initiated by free radicals; oxidation affects attributes such as taste,
1413 color, texture and nutritional value as oxidative chain reactions are associated with
1414 oxygen reaction with lipids, pigments, proteins and vitamins. ^[437] Synthetic and natural
1415 food additives with antioxidant properties have been developed to limit and control lipid
1416 oxidation and formation of the unpleasant byproducts in foods. In this context, spices and
1417 herbs have been always used in foods for their flavor; they have also been proven to
1418 contain high concentrations of components able to counteract oxidative reactions:
1419 phenolic compounds, carotenoids, and tocopherols just to cite a few. ^[438] As previously
1420 stated, nowadays EOs are widely used in the food industry for their ability to inhibit
1421 pathogen growth and retarding the food spoilage, thus improving food organoleptic
1422 quality. ^[439] Plant polyphenols can be definitely offered as alternatives to EOs; the plant
1423 extracts obtained from different sources such as fruits, vegetables, herbs, and spices have
1424 been in fact proven to possess strong antioxidant activity because of their high content of
1425 phenolic compounds. ^[440]

1426 The extracts have been prepared from the plant materials by using different
1427 solvents and extraction methods, both conventional or green. ^[441] In order to decrease or
1428 inhibit oxidative reactions in foods, antioxidants are usually used at an average level.
1429 Higher concentration may cause various side effects through pro-oxidative action. ^[442]

1430 *4.2 Food additives*

1431 The simplest way to employ an antioxidant compound (synthetic or natural) in food to
1432 preserve its nutritional and quality features is to add it to the food matrix before storage.
1433 One class of additives that could be replaced with natural extracts (spices, fruits and
1434 vegetable residues) are the antioxidants used in meat products. These compounds increase
1435 the shelf life of foods to which they are added by inhibiting the oxidation of lipids,
1436 proteins and pigments, preserving attributes such as color, texture, aroma, taste and the
1437 overall quality of the product. ^[443] As regarding the object of this review, the herbs from
1438 the *Lamiaceae* family, numerous papers present in literature report the use of sage,
1439 rosemary and oregano in the quality preservation of meat, fish and bread. Examples

1440 include the use of a sage extract to reduce the lipid oxidation in pork preparations ^[444] and
1441 oregano extracts employed to inhibit the spoilage of tilapia meat; anyhow the most
1442 popular herb in the food preservation sector seems to be rosemary, which was
1443 successfully tested as additive in the extension of the shelf life of carp, ^[445] pork meat ^[446]
1444 and chicken, ^[447] or in combination with thyme to preserve the quality of the already
1445 mentioned Nile tilapia. ^[448] The antioxidant properties of these extracts are due to the
1446 presence of phenolic acids caffeic and rosmarinic (4) and phenolic diterpenes, such as
1447 carnosic acid and carnosol. ^[60,95] Another serious problem related with food shelf life and
1448 storage is the spoilage caused by bacteria, yeast and molds as in fruits, vegetables and
1449 bakery products, mainly bread. The most critical factors controlling the growth of fungi
1450 on foodstuffs are again oxygen, but also temperature, pH, and water activity. Mold
1451 spoilage is particularly dangerous for consumer health as fungi are also responsible for
1452 the generation of mycotoxins and off-flavors, which are usually produced before fungal
1453 growth is visible. ^[449] In order to counteract these kind of contaminants in foodstuff
1454 physical methods like ultraviolet light, infrared radiation, microwave heating or ultra-
1455 high pressure treatments have been developed; however, the consumer-preferred solution
1456 is to replace traditionally used chemical preservatives with environmental friendly,
1457 “green” alternatives such as natural extracts bearing antifungal activity for the so called
1458 “biopreservation”. ^[450] As example, methanolic extract from oregano was employed in
1459 shelf life prolongation of “Rocha” pears via reduction of mold spoilage. ^[451] Finally,
1460 extract from oregano has been proven to act as antibacterial agent against *Listeria*
1461 *monocytogenes*, *Staphylococcus aureus*, and *Salmonella enterica* in cheese at room
1462 temperature; treatments with this extract also increased the stability of cheese against lipid
1463 peroxidation. ^[452]

1464 4.3 Edible coatings and films

1465 Rather than adding the additive directly to the food matrix, the use of edible coatings is
1466 one of the most important methods applied for preserving food quality. Edible films and
1467 coatings are promising systems to be used in the prolongation of food shelf life; as they
1468 can be consider both a packaging and a food component, they should have good sensory
1469 attributes, peculiar mechanical properties, biochemical, physicochemical, and microbial
1470 stability, safety, non-polluting nature, simple technology and low raw material and
1471 processing cost. These coatings are useful to improve food appearance and to delay

1472 transmission of moisture and oxygen (which cause oxidative degradation) but also of
1473 aroma and solutes during storage, thus enhancing product shelf life; furthermore, they can
1474 retard food spoilage and maintain food security by inhibiting the growth of
1475 microorganisms, usually due to the incorporation of antimicrobial compounds such as
1476 plant-derived matrices (single molecules, extracts or essential oils). The most famous
1477 edible coatings are made of proteins or polysaccharides, so they are considered
1478 environmentally friendly. In the “antimicrobial version”, the phytochemicals are
1479 gradually released on the food surface over time, maintaining a proper concentration of
1480 active components during the storage thus allowing the use of smaller amounts of active
1481 compounds compared with direct application (see previous paragraph). The controlled
1482 release of phytochemicals into the food matrix to preserve *via* edible coating depends on
1483 several factors: interaction between the antimicrobial agent and the polymer, osmosis,
1484 and environmental conditions (temperature, humidity, pH).^[453] As already stated
1485 aromatic herbs from Lamiaceae have been used for centuries to improve the sensory
1486 characteristics and to prolong the shelf life of food due to the antimicrobial and
1487 antioxidants properties of their essential oils and extracts.^[454] Examples reported in
1488 literature on edible films incorporating antioxidant/antimicrobial extract from Lamiaceae
1489 herbs include edible cassava starch film containing rosemary extracts having antioxidant
1490 activity,^[455] rosemary and oregano extracts-enriched gelatin films,^[456,457] again rosemary
1491 as additive in chitosan -based edible biopolymers for the preservation of beef burgers^[458]
1492 and the reduction of lipid oxidation and microbial spoilage in fish.^[459] Finally, rosemary
1493 in association with olive oleoresin in chitosan edible polymers has been proven to
1494 improve antioxidant protection and to prevent browning reaction in butternut squash
1495 (*Cucurbita moschata* Duch) and Romaine lettuce.^[460]

1496 *4.4 Active and intelligent packaging*

1497 Edible films and coatings are just a subset in the broader scenario of the use of technology
1498 to increase food shelf life and food security in the design, creation and employment of the
1499 so called active and intelligent (smart) packaging. An active package was firstly defined
1500 by Rooney^[461] as a material that “performs a role other than an inert barrier to the outside
1501 environment”. Nowadays an “active” packaging is defined as a “packaging in which
1502 subsidiary constituents have been deliberately included in or on either the packaging
1503 material or the package headspace to enhance the performance of the package system”,

1504 while the current definition of “intelligent packaging is “packaging that contains an
1505 external or internal indicator to provide information about aspects of the history of the
1506 package and/or the quality of the food”, respectively. In contrast to traditional packaging,
1507 active and intelligent packaging may change the composition and organoleptic
1508 characteristics of food, provided that the changes are consistent with the provisions for
1509 food. ^[462] The principles behind active packaging are based either on the intrinsic
1510 properties of the polymer used as packaging material itself or on the introduction of
1511 specific substances inside the polymer, such as plant extracts. Packaging may be called
1512 “active” when it performs some desired role in food preservation other than providing an
1513 inert barrier to external conditions. ^[463] Intelligent packaging gives useful information to
1514 the consumer based on its ability to sense, detect, or record external or internal changes
1515 in the food products’ environment. There are two types of intelligent packaging: one
1516 based on measuring the condition of the package on the outside, the other measuring
1517 directly the quality of the food product inside the packaging. ^[464] On the other hand, active
1518 packaging includes additives capable of scavenging, absorbing or releasing gasses such
1519 as oxygen, carbon dioxide and ethylene but also moisture, ethanol, antioxidants and
1520 antimicrobials. The main advantages of using active packaging for the application of
1521 natural antimicrobials in foods are the controlled release of the bioactive compounds into
1522 the product during storage time and the lower possibility of development of undesirable
1523 flavors than the direct addition into food could cause. ^[465] Plant or fruit extracts having
1524 antioxidant and/or antimicrobial activities can be used in the packaging with the aim of i)
1525 having an action in the packaging or ii) with the intention of being released into the food
1526 to avoid its oxidation and spoilage. Rosemary extract is one of the plant extracts that has
1527 already been successfully incorporated into food packaging. Rosemary contains several
1528 flavones (apigenin, genkwanin, hesperetin and cirsimaritin), phenolic diterpenes
1529 (carnosic acid, carnosol, rosmadial, epirosmanol, rosmanol) and phenolic acids (caffeic
1530 acid and rosmarinic acid, the latest being the main compound in polar extract from this
1531 herb). ^[95] Carnosic acid and rosmarinic acid are considered as the compounds responsible
1532 for rosemary’s activities. According to numerous papers found in literature, rosemary
1533 extracts are effective in delaying the lipid oxidation in meat when immobilized on
1534 polypropylene films ^[466] or in contrasting oxidation and browning in fresh-cut pears. ^[467]
1535 An ethanolic extract from rosemary was also successfully tested as antimicrobial agent

1536 on cooked ready-to-eat vacuum-packaged shrimps. Another herb from the Lamiaceae
1537 family that has been incorporated into food packaging is oregano, which again contains
1538 rosmarinic acid as one of principal compounds in its polar extracts. ^[60] Camo and others
1539 ^[468] studied the prolongation of the shelf life of beef when packaged with a film containing
1540 oregano extract in various concentrations. Bentayeb and co-authors ^[469] have confirmed
1541 the effectiveness of the film with oregano extract when comparing the antioxidant
1542 capacity of various natural extracts incorporated in active packaging.

1543 **5. A novel key to understanding bioactivity: interactions between** 1544 **Lamiaceae herbs and human microbiota**

1545 *5.1 Symbiotic (commensal) bacteria: the human microbiota*

1546 We are all inhabited by complex communities of microorganisms known as the
1547 microbiota, which reside in most surfaces of the human body. It is estimated that there is
1548 up to 100 trillion bacteria in the same individual, which is around 10-fold greater than the
1549 number of human cells. ^[470] The majority of the human commensal bacteria (10–100
1550 trillion) can be found in the gastrointestinal tract and particularly in the distal intestine,
1551 where they have many beneficial functions such as fermentation of indigestible dietary
1552 residues, production of vitamins, control of intestinal epithelial cell proliferation and
1553 differentiation, and the creation of a protective barrier against pathogens; ^[471] the
1554 gastrointestinal tract microbiota also contributes to the development and differentiation
1555 of the mammalian immune system. The composition of the microbiota affects immune
1556 responses, susceptibility to infection and development of allergic and inflammatory
1557 diseases. Antibiotic administration usually perturbs microbial communities and decreases
1558 host resistance to antibiotic resistant microbes. Following perturbations by antibiotics,
1559 diet, immune deficiency, or infection, this ecosystem can shift to a state of dysbiosis, a
1560 state that may be deleterious to the host. Recent evidence suggests that there is a link
1561 between the human gut microbiome and the development various illnesses such as
1562 obesity, cardiovascular disease, and metabolic syndromes. Similarly, there is increasing
1563 evidence for dysbiotic microbiota in carcinogenesis. Also a panel of gastrointestinal
1564 complaints, including functional and inflammatory gastrointestinal diseases, are
1565 obviously associated with altered gut microbiota. ^[472] A deeper understanding of the
1566 mechanisms correlating changes in microbiota composition (dysbiosis) with disease
1567 states is needed to improve our knowledge. ^[473]

1568 5.2 Interactions between vegetable dietary components, phytochemicals and human gut
1569 microbiota: still a lot to investigate

1570 Numerous papers in literature suggests that dietary components, including
1571 phytochemicals, can interact with microbial populations in the human host and have the
1572 potential to modify them. Phytochemicals are compounds present in our diet having a
1573 wide range of recognized effects including anti-inflammatory, anti- cancer, anti-oxidant,
1574 and other beneficial properties both *in vivo* and *in vitro*. Understanding how diet can
1575 interact with microbiota will contribute to developing personalized nutrition to manage
1576 diseases. Interactions between the gut microbiome and the diet are complex and dynamic,
1577 as diet is the primary determinant in the development of the microbiota colonization
1578 pattern from the first stages of life. ^[474] Not all the phytochemicals orally ingested via
1579 plant-based food or herbal medicinal products get into contact with colonic
1580 microorganisms, but only those that remain unaltered in the upper intestinal tract. For
1581 these molecules, interactions with gut microbiota are dual and reciprocal: plant
1582 constituents are metabolized by gut microbial populations, leading to the formation of
1583 bacterial catabolites with altered bioactivity profile, and on the other hand, plant
1584 constituents can initiate changes in microbial community composition and function.
1585 Thumann et al. ^[475] recently surveyed nearly 50 plants traditionally used for
1586 gastrointestinal diseases, concluding that only a very limited portion of them has been
1587 investigated so far with regard to potential interactions with gut microbiota. As example,
1588 constituents present in some of these plants like curcumin (curcuma), shogaol (ginger),
1589 and rosmarinic acid (Lamiaceae herbs) have been shown to be metabolized by human gut
1590 microbiota, and preliminary data also indicate potential gut microbiome modulatory
1591 effects. Rosmarinic acid can be considered a “family marker” for Lamiaceae herbs, being
1592 present in polar extracts from the aerials of all family members; it has been shown to be
1593 readily degraded by gut microbiota *in vitro*, as well as after oral intake in experimental
1594 animals and in humans ^[476] producing caffeic acid and other several hydroxypropionic
1595 acid as catabolites. In a recent paper, Madureira et al. demonstrated that in turn rosmarinic
1596 acid affected microbiome composition. ^[477] The interaction of herbs and their
1597 components, phytochemicals, with gut microbiota is still therefore not fully investigated
1598 offering an open field for future research.

1599

1600 **6. Conclusions**

1601 Plants have ‘accompanied’ the evolution of the human beings supplying an essential
1602 portion of foods, and a series of ‘tools’ able to fight several pathologies or generically
1603 speaking many health problems.

1604 The species belonging to the Lamiaceae family represents a small portion of this
1605 large resource that the humanity would do well to protect and safeguard, through
1606 concerted and targeted efforts of public and private institutions, and the research sectors.

1607 To-day, alongside the classic and still effective methods of analyses aimed to the
1608 discover of new compounds and phytocomplexes, are available new tools, for example in
1609 the health sector but not only, to fight the medical emergencies of this millennium.

1610 With this in mind, the here reported limited survey on the Lamiaceae family shows
1611 as the phytochemical studies of these species (as well as of other plants) are still essential
1612 being the main phytochemical components, represented by the essential oils and not
1613 volatile extracts, an invaluable and fundamental help for all of us.

1614 Their biological activities are continuously confirmed by new studies, carried out
1615 with more sophisticated models and more suited to new applications and technological
1616 needs. The potential use of medicinal plants in arid and with difficult climatic conditions
1617 areas has continued to promote the identification and study of new endemisms.

1618 In particular, concerning the essential oils one of the most interesting development
1619 regards their antimicrobial studies, while still in the health field, they seem to slow down
1620 the progress in their application as anticancer and antiviral. Moreover, the best prospects
1621 are currently concentrated in the food industry thanks to the synergy with
1622 nanotechnologies. Very early and interesting are the first results of application of essential
1623 oils in the field of cultural heritage preservation.

1624 Regarding the extracts from the plants of this family they are not so popular and
1625 known as the essential oils, however, from the studies performed in these last years they
1626 have showed a large plethora of bioactivities, thank also to the ethnopharmacology and
1627 ethnobotanical surveys of their uses in many countries. The phytochemical studies, as
1628 reported in this review, showed that the main components characterized in the various
1629 types of extracts belong to the polyphenols family, and these components are considered
1630 the main actors for the anticancer, antibacterial, anti-inflammatory, and cytotoxic and
1631 others activities of several phytocomplexes.

1632 In conclusion taking into account all the aspects reported in this review it is
1633 possible to affirm that the species of this family seem to behave like those great classics
1634 of literature, for which even if time passes they always remain current. The next big
1635 challenges probably concern an increase of their production possibly correlated to a
1636 sustainable management, the development of alternative and green technological
1637 processes requiring high energy consumption, and an ecological and rationale treatment
1638 of the waste products.

1639

1640 **Acknowledgements**

1641 Special thanks are due to Dr. Carmelo Rinoldo [Assessorato Regionale dell'Agricoltura,
1642 Regione Siciliana – Aragona (AG)] for the kind gift of the original photos, and to Mr.
1643 Antonio Greco (ICB-CNR, Catania) for his skilful technical assistance.

1644

1645 **Author Contribution Statement**

1646 G. R. conceived and designed the structure of the work. E. N. organized and wrote the
1647 section of the essential oils, L. S. carried out the organization and the writing of the section
1648 dealing with the extracts. G. R. assembled all sections and wrote the final version with
1649 the contribution from all co-authors. All authors participated in literature analysis,
1650 reviewed and approved the final version of the manuscript.

1651

1652 **References**

- 1653
- 1654
- 1655 [1] www.theplantlist.org; www.kew.org; www.mpns.science.kew.org. (Accessed November 2019).
- 1656
- 1657 [2] A. Lubbe, R. Verpoorte, ‘Cultivation of medicinal and aromatic plants for speciality industrial
- 1658 material’, *Ind. Crops Prod.* **2011**, *34*, 785-801.
- 1659
- 1660 [3] L. E. Craker, ‘Medicinal and aromatic plants – new opportunities, n: J. Janick, A. Whipkey (Eds.)
- 1661 New crops and new uses: Creating markets for economic development. Alexandria, VA, ASHS,
- 1662 2007, pp. 248-257.
- 1663
- 1664 [4] WHO, Global Health Estimates, The top ten causes of death, 2016: Deaths by Cause, Age, Sex,
- 1665 by Country and by Region, 2000-2016, World Health Organization, 2018, Geneva.
- 1666
- 1667 [5] WHO global report on traditional and complementary medicine 2019. Geneva: World Health
- 1668 Organization; 2019. Licence: CC BY-NC-SA 3.0 IGO.
- 1669
- 1670 [6] E. M. Napoli, G. Ruberto, Sicilian aromatic plants: from traditional heritage to a new agro-
- 1671 industrial exploitation, in: J.F. Kralis (Ed.) Spices: types, uses and health benefits, Nova Science
- 1672 Publ. Inc. NY, USA, 2012, pp. 1-56. ISBN: 978-1-61470-820-9.
- 1673
- 1674 [7] B. Allkin, Useful plants–medicines use, in: K.J. Willis (Ed.) State of the World’s Plants- 2017,
- 1675 Royal Botanic Gardens, Kew, London, UK, 2017, pp. 22-29.
- 1676
- 1677 [8] F. Jamshidi-Kia, Z. Lorigooini, H. Amini-Khoei, ‘Medicinal plants: past history and future
- 1678 perspective’, *J. Herbmed Pharmacol.* **2018**, *7(1)*, 1-7.
- 1679
- 1680 [9] M. Inoue, S. Hayashi, L. E. Craker, Role of medicinal and aromatic plants: Past, present, and
- 1681 future, IntechOpen, 2019, pp. 1-13, [<http://creativecommons.org/licenses/by/3.0>, DOI:
- 1682 [dx.doi.org/10.5772/intechopen.82497](https://doi.org/10.5772/intechopen.82497)].
- 1683
- 1684 [10] F. Barbero, M. Maffei, Chapter 3 - Biodiversity and chemotaxonomic significance of specialized
- 1685 metabolites, in: G. Arimura, M.E. Maffei (Eds.) Plant specialized metabolism: Genomics,
- 1686 biochemistry, and biological functions, CRC Press, 2016, pp. 24-63.
- 1687
- 1688 [11] J. D. Ross, C. Sombroero, Environmental control of essential oil production in Mediterranean
- 1689 plants, in: J.B. Harborne, F.A.T. Barberin (Eds) Ecological chemistry and biochemistry of plant
- 1690 terpenoids. Clarendon Press, Oxford, 1991, pp. 83–94.
- 1691
- 1692 [12] W. Thuiller, S. Lavorel, M.B. Araujo, M.T. Sykes, I.C. Prentice, ‘Climate change threats to plant
- 1693 diversity in Europe’, *PNAS*, **2005**, *102(23)*, 8245–8250.
- 1694
- 1695
- 1696 [13] B. Khameneh, M. Iranshahy, N. Vahdati-Mashhadian, A. Sahebkar, B. S. F. Bazzaz, ‘Non-
- 1697 antibiotic adjunctive therapy: A promising approach to fight tuberculosis’, *Pharmacol. Res.* **2019**,
- 1698 *146*, #104289.
- 1699
- 1700 [14] M. Ayaza, F. Ullah, A. Sadiq, F. Ullah, M. Ovais, J. Ahmed, H.P. Devkota, ‘Synergistic
- 1701 interactions of phytochemicals with antimicrobial agents: Potential strategy to counteract drug
- 1702 resistance’, *Chem.-Biol. Inter.* **2019**, *308*, 294-303.
- 1703
- 1704 [15] A. L. Demain, B. Gómez-Ortiz, B. Ruiz-Villafán, R. Rodríguez-Sanoja, S. Sánchez, ‘Recent
- 1705 findings of molecules with anti-infective activity: screening of non-conventional sources’, *Curr.*
- 1706 *Opin. Pharm.* **2019**, *48*, 40-47.
- 1707
- 1708 [16] V. Giannakis, S. Miyaki, ‘Novel antimicrobial agents against multi-drug resistant gram-positive
- 1709 bacteria: an overview’. *Recent Pat. Antiinfect. Drug Discov.* **2012**, *7(3)*, 182-188.

- 1710 [17] I. Nicolas, V. Bordeau, A. Bondon, M. Baudy-Floc'h, B. Felden, 'Novel antibiotics effective
1711 against gram-positive and -negative multi-resistant bacteria with limited resistance. *PLoS Biol*
1712 **2019**, *17*(7), e3000337, pp. 1-23.
1713
- 1714 [18] G. N. S. Hema Sree, G. R. Saraswathy, M. Muraharia, M. Krishnamurth, 'An update on drug
1715 repurposing: Re-written saga of the drug's fate', *Biomed. Pharmacot.* **2019**, *110*, 700-716.
1716
- 1717 [19] M. R. Byuna, C. H. Kim, H. S. Lee, J. W. Choia, S. K. Lee, 'Repurposing of ginseng extract as
1718 topoisomerase I inhibitor based on the comparative analysis of gene expression patterns',
1719 *Phytochemistry*, **2019**, *164*, 223-227.
1720
- 1721 [20] M. R. Byuna, D. H. Lee, Y. P. Jang, H. S. Lee, J. W. Choi, S. K. Lee, 'Repurposing natural products
1722 as novel HDAC inhibitors by comparative analysis of gene expression profiles', *Phytomedicine*
1723 **2019**, *59*, #152900.
1724
- 1725 [21] J. Lamb, E. D. Crawford, D. Peck, J. W. Modell, I. C. Blat, M. J. Wrobel, J. Lerner, J. -P. Brunet,
1726 A. Subramanian, K. N. Ross, 'The Connectivity Map: using gene expression signatures to connect
1727 small molecules, genes, and disease', *Science* **2006**, *313*, 1929-1935.
1728
- 1729 [22] J. L. Smalley, T. W. Gant, S. -D. Zhang, 'Application of connectivity mapping in predictive
1730 toxicology based on gene-expression similarity', *Toxicology* **2010**, *268*, 143-146.
1731
- 1732 [23] Y. Nishimura, H. Hara, 'Editorial: Drug repositioning: current advances and future perspectives',
1733 *Front. Pharm.* **2018**, #1068
1734
- 1735 [24] K. N. De Abrew, Y. K. Shan, X. Wang, J. M. Krailler, R. M. Kainkaryama, C. C. Lester, R. S.
1736 Settivari, M. J. LeBaron, J. M. Naciff, G. P. Daston, 'Use of connectivity mapping to support read
1737 across: A deeper dive using data from 186 chemicals, 19 cell lines and 2 case studies', *Toxicology*,
1738 **2019**, 84-94.
1739
- 1740 [25] H. Zhang, J. Pan, X. Wu, A. -R. Zuo, Y. Wei, Z. -L. Ji, 'Large-scale target identification of herbal
1741 medicine using a reverse docking approach', *ACS OMEGA*, **2019**, *4*, 9710-9719.
1742
- 1743 [26] E. Danila, D. A. Kaya, M. Patrascu, M. A. Kaya, S. Kumbakisaka, 'Comparative study of
1744 *Lavandula angustifolia* essential oils obtained by microwave and classic hydrodistillation', *Rev.*
1745 *Chim.* **2018**, *69*(8), 2240-2244.
1746
- 1747 [27] M. Elyemmi, B. Louaste, I. Nechad, T. Elkamli, A. Bouia, M. Taleb, M. Chaouch, N. Eloutassi,
1748 'Extraction of essential oils of *Rosmarinus officinalis* L. by two different methods:
1749 hydrodistillation and microwave assisted hydrodistillation', *The Scientific World Journal* **2019**,
1750 Article ID 3659432.
1751
- 1752 [28] M. Mohammadhosseini, A. Akbarzadeh, G. Flamini, 'Profiling of compositions of essential oils
1753 and volatiles of *Salvia limbata* using traditional and advanced techniques and evaluation for
1754 biological activities of their extracts', *Chem. Biodiversity* **2017**, *14*, e1600361.
1755
- 1756 [29] C. Lilia, A. Abdelkader, A. A. Karima, B. Tarek, 'The effect of ultrasound pre-treatment on the
1757 yield, chemical composition and antioxidant activity of essential oil from wild *Lavandula stoechas*
1758 L.', *J. Essent. Oil Bearing Plants* **2018**, *21*(1), 253-263.
1759
- 1760 [30] G. Khalili, A. Mazloomifar, K. Larijani, M. S. Tehrani, P. A. Azar, 'Solvent-free microwave
1761 extraction of essential oils from *Thymus vulgaris* L. and *Melissa officinalis* L.', *Ind. Crops Prod.*
1762 **2018**, *119*, 214-217.
1763
- 1764 [31] M. M. A. Rashed, Q. Tong, A. Nagi, J. Li, N. U. Khan, L. Chen, A. Rotail, A. M. Bakry, "Isolation
1765 of essential oil from *Lavandula angustifolia* by using ultrasonic-microwave assisted method
1766 preceded by enzymolysis treatment, and assessment of its biological activities", *Ind. Crops Prod.*
1767 **2017**, *100*, 236-245.

- 1768 [32] F. H. Giray, 'An analysis of world Lavender oil markets and lessons for Turkey', *J. Essent. Oil*
1769 *Bearing Plants* **2018**, 21(6), 1612-1623.
1770
- 1771 [33] A. C. Aprotosoae, E. Gille, A. Trifan, V. S. Luca, A. Miron, 'Essential oils of *Lavandula* genus:
1772 a systematic review of their chemistry", *Phytochem. Rev.* **2017**, 16, 761-799.
1773
- 1774 [34] K. Smigielski, R. Prusinowska, A. Stobiecka, A. Kunicka-Styczyńska, R. Gruska, 'Biological
1775 Properties and Chemical Composition of Essential Oils from Flowers and Aerial Parts of Lavender
1776 (*Lavandula angustifolia*)', *J. Essent. Oil Bearing Plants* **2018**, 21(5), 1303-1314.
1777
- 1778 [35] L. Pistelli, B. Najara, S. Giovanelli, L. Lorenzini, S. Tavarini, L. G. Angelini, 'Agronomic and
1779 phytochemical evaluation of lavandin and lavender cultivars cultivated in the Tyrrhenian area of
1780 Tuscany (Italy)', *Ind. Crops Prod.* **2017**, 109, 37-44.
1781
- 1782 [36] B. Blažeković, W. Yang, Y. Wang, C. Li, M. Kindl, S. Pepeljnjak, S. Vladimir-Knežević,
1783 'Chemical composition, antimicrobial and antioxidant activities of essential oils of
1784 *Lavandula × intermedia* 'Budrovka' and *L. angustifolia* cultivated in Croatia', *Ind. Crops Prod.*
1785 **2018**, 123, 173-182.
1786
- 1787 [37] S. Küçük, E. Çetintaş, M. Kürkçüoğlu, 'Volatile compounds of the *Lavandula angustifolia* Mill.
1788 (Lamiaceae) Species Cultured in Turkey', *J. Turk. Chem. Soc., Sect. A: Chem.* **2018**, 5(3), 1303-
1789 1308.
1790
- 1791 [38] A. Carrasco, R. Martinez-Gutierrez, V. Tomas, J. Tudela, '*Lavandula angustifolia* and *Lavandula*
1792 *latifolia* essential oils from Spain: aromatic profile and bioactivities', *Planta Med.* **2016**, 82, 163-
1793 170.
1794
- 1795 [39] S. Kivrak, 'Essential oil composition and antioxidant activities of eight cultivars of Lavender and
1796 Lavandin from western Anatolia', *Ind. Crops Prod.* **2018**, 117, 88-96.
1797
- 1798 [40] P. Singh, H. Andola, M. S. M. Rawat, G. J. N. Panta, J. S. Jangwan, 'GC-MS Analysis of Essential
1799 Oil from *Lavandula angustifolia* Cultivated in Garhwal Himalaya", *The Natural Products Journal*
1800 **2015**, 5, 268-272.
1801
- 1802 [41] M. Belhadj Mostefa, A. Kabouche, I. Abaza, T. Aburjai, R. Touzani, Z. Kabouche, 'Chemotypes
1803 investigation of *Lavandula* essential oils growing at different North African soils', *J. Mater.*
1804 *Environ. Sci.* **2014**, 5(6), 1896-1901.
1805
- 1806 [42] A. Carrasco, V. Ortiz-Ruiz, R. Martinez-Gutierrez, V. Tomas, J. Tudela, '*Lavandula stoechas*
1807 essential oil from Spain: Aromatic profile determined by gas chromatography-mass
1808 spectrometry, antioxidant and lipoxygenase inhibitory bioactivities', *Ind. Crops Prod.* **2015**, 73,
1809 16-27.
1810
- 1811 [43] T. Tuttolomondo, G. Dugo, G. Ruberto, C. Leto, E. M. Napoli, A. G. Potorti, M. R. Fede, G. Virga,
1812 R. Leone, E. D' Anna, M. Licata, S. La Bella, 'Agronomical evaluation of Sicilian biotypes of
1813 *Lavandula stoechas* L. spp. *stoechas* and analysis of the essential oils', *J. Essent. Oil Res.* **2015**,
1814 27(2), 115-124.
1815
- 1816 [44] S. La Bella, T. Tuttolomondo, G. Dugo, G. Ruberto, C. Leto, E. M. Napoli, A. G. Potorti, M. R.
1817 Fede, G. Virga, R. Leone, E. D'Anna, M. Licata, "Composition and Variability of the Essential
1818 Oil of the Flowers of *Lavandula stoechas* from Various Geographical Sources, *Nat. Prod. Comm.*
1819 **2015**, 10(11), 2001-2004.
1820
- 1821 [45] S. Robu, B. I. Chesaru, C. Diaconu, O. Dumitribuzia, D. Tutunaru, U. Stanescu, E. L. Lisa,
1822 'Lavandula hybrida: microscopic characterization and the evaluation of essential oil', *Farmacia*
1823 **2016**, 64(6), 914-917.
1824

- 1825 [46] I. Bajalan, R. Rouzbahani, A. G. Pirbalouti, F. Maggi, 'Chemical composition and antibacterial
1826 activity of Iranian *Lavandula x hybrida*', *Chem. Biodiversity* **2017**, e1700064.
1827
- 1828 [47] A. El Hamdaoui, F. Msanda, H. Boubaker, D. Leach, I. Bombarda, P. Vanloot, N. El Aouad, A.
1829 Abbad, E. H. Boudyach, F. Achemchem, A. Elmoslih, A. Ait Ben Aoumar, A. El Mousadik,
1830 'Essential oil composition, antioxidant and antibacterial activities of wild and cultivated
1831 *Lavandula mairei* Humbert', *Biochem. Syst. Ecol.* **2018**, *76*, 1-7.
1832
- 1833 [48] M. P. Argentieri, B. De Lucia, G. Cristiano, P. Avato, 'Compositional Analysis of *Lavandula*
1834 *pinnata* Essential Oils', *Nat. Prod. Comm.* **2016**, *11(3)*, 287-290.
1835
- 1836 [49] R. N. Al-Badani, J. K. R. Da Silva, W. N. Setzer, N. A. Awadh Ali, B. A. Muharam, A. J. A. Al-
1837 Fahad, 'Variations in Essential Oil Compositions of *Lavandula pubescens* (Lamiaceae) Aerial
1838 Parts Growing Wild in Yemen', *Chem. Biodiversity* **2017**, *14*, e1600286.
1839
- 1840 [50] A. Alizadeh, Z. Aghaee, 'Essential oil constituents, phenolic content and antioxidant activity of
1841 *Lavandula stricta* Delile growing wild in southern Iran', *Nat. Prod. Res.* **2016**, *30(19)*, 2253-2257.
1842
- 1843 [51] F. Baali, S. Boumerfeg, E. Napoli, A. Boudjelal, N. Righi, A. Deghima, A. Baghiani, G. Ruberto,
1844 Chemical composition and biological activities of essential oils from two wild Algerian medicinal
1845 plants: *Mentha pulegium* L. and *Lavandula stoechas* L. *J. Essent. Oil Bear, Pl.* **2019**, *22(3)*, 821-
1846 837.
- 1847 [52] N. Leyva-López, E. P. Gutiérrez-Grijalva, G. Vazquez-Olivo, J. Basilio Heredia, 'Essential Oils
1848 of Oregano: Biological Activity beyond Their Antimicrobial Properties', *Molecules* **2017**, *22*, 989.
1849
- 1850 [53] B. Lukas, C. Schmiderer, J. Novak, 'Essential oil diversity of European *Origanum vulgare* L.
1851 (Lamiaceae)', *Phytochemistry* **2015**, *119*, 32-40.
1852
- 1853 [54] G. De Mastro, W. Tarraf, L. Verdini, G. Brunetti, C. Ruta, 'Essential oil diversity of *Origanum*
1854 *vulgare* L. populations from Southern Italy', *Food Chem.* **2017**, *235*, 1-6.
1855
- 1856 [55] E. M. Napoli, G. Curcuruto, G. Ruberto, 'Screening the essential oil composition of wild Sicilian
1857 oregano', *Biochem. Syst. Ecol.* **2009**, *37*, 484-493.
1858
- 1859 [56] A. Carrasco, E. Perez, A. Cutillas, R. Martinez-Gutierrez, V. Tomas, J. Tudela, '*Origanum vulgare*
1860 and *Thymbra capitata* essential oils from Spain: determination of aromatic profile and
1861 bioactivities', *Nat. Prod. Comm.* **2016**, *11(1)*, 113-120.
1862
- 1863 [57] M. R. Morshedloo, S. A. Salami, V. Nazeri, F. Maggi, L. Craker, 'Essential oil profile of oregano
1864 (*Origanum vulgare* L.) populations grown under similar soil and climate conditions', *Ind. Crops*
1865 *Prod.* **2018**, *119*, 183-190.
1866
- 1867 [58] M. Hatipi, V. Papajani, S. Cavar, R. Koliqi, 'Analysis of volatile compounds of *Origanum vulgare*
1868 L. growing wild in Kosovo', *J. Essent. Oil Bear. Pl.* **2014**, *17(1)*, 148-157.
1869
- 1870 [59] C. M. Asensio, N. R. Grosso, H. R. Juliani, 'Quality characters, chemical composition and
1871 biological activities of oregano (*Origanum* spp.) essential oils from Central and Southern
1872 Argentina', *Ind. Crops Prod.* **2015**, *63*, 203-213.
1873
- 1874 [60] T. Tuttolomondo, S. La Bella, M. Licata, G. Virga, C. Leto, A. Saija, D. Trombetta, A. Tomaino,
1875 A. Speciale, E. M. Napoli, L. Siracusa, A. Pasquale, G. Curcuruto, G. Ruberto, 'Biomolecular
1876 characterization of wild Sicilian oregano: phytochemical screening of essential oils and extracts,
1877 and evaluation of their antioxidant activities', *Chem. Biodiversity* **2013**, *10*, 411-433.
1878
- 1879 [61] T. Tuttolomondo, C. Leto, R. Leone, M. Licata, G. Virga, G. Ruberto, E. M. Napoli, S. La Bella,
1880 'Essential oil characteristics of wild Sicilian oregano populations in relation to environmental
1881 conditions', *J. Essent. Oil Res.* **2014**, *26(3)*, 210-220.

- 1882 [62] M. Licata, T. Tuttolomondo, G. Dugo, G. Ruberto, C. Leto, E. M. Napoli, R. Rando, M.R. Fede,
1883 G. Virga, R. Leone, S. La Bella, 'Study of quantitative and qualitative variations in essential oils
1884 of Sicilian oregano biotypes', *J. Essent. Oil Res.* **2015**, *27(4)*, 293-306.
1885
- 1886 [63] E. Mancini, I. Camele, H. S. Elshafie, L. De Martino, C. Pellegrino, D. Grulova, V. De Feo,
1887 'Chemical composition and biological activity of the essential oil of *Origanum vulgare* ssp. *hirtum*
1888 from different areas in the Southern Apennines (Italy)', *Chem. Biodiversity* **2014**, *11*, 639-651.
1889
- 1890 [64] D. Stešević, Ž. Jaćimović, Z. Šatović, A. Šapčanin, G. Jančan, M. Kosović, B. Damjanović-
1891 Vratnica, 'Chemical characterization of wild growing *Origanum vulgare* populations in
1892 Montenegro', *Nat. Prod. Comm.* **2018**, *13(10)*, 1357-1362.
1893
- 1894 [65] E. Napoli, A. Mazzaglia, C. Restuccia, P. Ragni, C.M. Lanza, G. Ruberto, 'The effect of γ -
1895 irradiation on chemical composition, microbial load and sensory properties of Sicilian oregano',
1896 *LWT Food Sci. Tech.* **2016**, *72*, 566-572.
1897
- 1898 [66] A. Bouyahya, F. Guaouguaou, N. Dakka, Y. Bakri, 'Pharmacological activities and medicinal
1899 properties of endemic Moroccan medicinal plant *Origanum compactum* (Benth) and their main
1900 compounds', *Asian Pac. J. Trop. Dis.* **2017**, *7(10)*, 628-640.
1901
- 1902 [67] Y. Laghmouchi, O. Belmehdi, N. S. Senhaji, J. Abrini, 'Chemical composition and antibacterial
1903 activity of *Origanum compactum* Benth. essential oils from different areas at northern Morocco',
1904 *S. Afr. J. Bot.* **2018**, *115*, 120-125.
1905
- 1906 [68] K. Aboukhalid, A. Lamiri, M. Agacka-Mołdoch, T. Doroszevska, A. Douaik, M. Bakha, J.
1907 Casanova, F. Tomi, N. Machon, C. Al Faiza, 'Chemical polymorphism of *Origanum compactum*
1908 grown in all natural habitats in Morocco', *Chem. Biodiversity* **2016**, *13*, 1126-1139.
1909
- 1910 [69] R. Zgheib, M. El-Beyrouthy, S. Chaillou, N. Ouaini, D. N. Rutledge, D. Stien, A. Kassouf, M.
1911 Leonti, M. Iriti, 'Chemical variability of the essential oil of *Origanum ehrenbergii* Boiss. from
1912 Lebanon, assessed by independent component analysis (ICA) and common component and
1913 specific weight analysis (CCSWA)', *Int. J. Mol. Sci.* **2019**, *20*, 1026.
1914
- 1915 [70] M. Marrelli, F. Conforti, C. Formisano, D. Rigano, N. A. Arnold, F. Menichini, F. Senatore,
1916 'Composition, antibacterial, antioxidant and antiproliferative activities of essential oils from three
1917 *Origanum* species growing wild in Lebanon and Greece', *Nat. Prod. Res.* **2016**, *30(6)*, 735-739.
1918
- 1919 [71] M. Al Hafi, M. El Beyrouthy, N. Ouaini, D. Stien, D. Rutledge, S. Chaillou, 'Chemical
1920 composition and antimicrobial activity of *Origanum libanoticum*, *Origanum ehrenbergii*, and
1921 *Origanum syriacum* growing wild in Lebanon', *Chem. Biodiversity* **2016**, *13*, 555-560.
1922
- 1923 [72] A. K. Gulec, P. Erecevit, E. Yuce, A. Arslan, E. Bagci, S Kirbag, 'Antimicrobial activity of the
1924 methanol extracts and essential oil with the composition of endemic *Origanum acutidens*
1925 (Lamiaceae)', *J. Essent. Oil Bear. Pl.* **2014**, *17(2)*, 353-358.
1926
- 1927 [73] F. Daoudi-Merbah, M. Hazzit, M. Dahmani-Megrerouche, 'Influence of morphological variability
1928 and habitat on the chemical composition of essential oils of an Algerian endemic *Origanum* species
1929 (*Origanum floribundum* MUNBY)', *Chem. Biodiversity* **2016**, *13*, 1088-1094.
1930
- 1931 [74] O. Kosakowska, W. Czupa, 'Morphological and chemical variability of common oregano
1932 (*Origanum vulgare* L. subsp. *vulgare*) occurring in eastern Poland', *Herba Pol.* **2018**, *64(1)*, 11-
1933 21.
1934
- 1935 [75] A. P. Raina, K. S. Negi, 'Chemical diversity among different accessions of *Origanum vulgare* L.
1936 ssp. *vulgare* collected from Central Himalayan region of Uttarakhand, India', *J. Essent. Oil Res.*
1937 **2014**, *26(6)*, 420-426.
1938

- 1939 [76] Ö. Kilic, F. A. ÖÖ, ‘Variability of essential oil composition of *Origanum vulgare* L. subsp. *gracile*
1940 populations from Turkey’, *J. Essent. Oil Bear. Pl.* **2016**, *19(8)*, 2083-2090.
- 1941
- 1942 [77] M. Moradi, A. Hassani, F. Sefidkon, H. Maroofi, ‘Chemical composition of leaves and flowers
1943 essential oil of *Origanum vulgare* ssp. *gracile* growing wild in Iran’, *J. Essent. Oil Bear. Pl.* **2015**,
1944 *18(1)*, 242-247.
- 1945
- 1946 [78] K. Turgut, Y. Özyigit, B. Tutuncu, E. Ucar Sozmen, ‘Agronomic and chemical performance of
1947 selected *Origanum dubium* Boiss. clones for industrial use’, *Turk. J. Agric. For.* **2017**, *41*, 272-
1948 277
- 1949
- 1950 [79] A. N. El Gendy, M. Leonardi, L. Mugnaini, F. Bertelloni, V. V. Ebani, S. Nardoni, F. Mancianti,
1951 S. Hendawy, E. Omer, L. Pistelli, ‘Chemical composition and antimicrobial activity of essential
1952 oil of wild and cultivated *Origanum syriacum* plants grown in Sinai, Egypt’, *Ind. Crops Prod.*
1953 **2015**, *67*, 201-207.
- 1954
- 1955 [80] G. Semiz, A. Semiz, N. Mercan-Doğan, ‘Essential oil composition, total phenolic content,
1956 antioxidant and antibiofilm activities of four *Origanum* species from southeastern Turkey’, *Int. J.*
1957 *Food Prop.* **2018**, *1*, 194-204.
- 1958
- 1959 [81] H. Hajlaoui, H. Mighri, M. Aouni, N. Gharsallah, A. Kadri, ‘Chemical composition and in vitro
1960 evaluation of antioxidant, antimicrobial, cytotoxicity and anti-acetylcholinesterase properties of
1961 Tunisian *Origanum majorana* L. essential oil’, *Microb. Pathog.* **2016**, *95*, 86-94.
- 1962
- 1963 [82] H. O. Elansary, ‘Chemical diversity and antioxidant capacity of essential oils of marjoram in
1964 Northwest Egypt’, *J. Essent. Oil Bear. Pl.* **2015**, *18(4)*, 917-924.
- 1965
- 1966 [83] A. Stefanaki, C. M. Cook, T. Lanaras, S. Kokkini, ‘The Oregano plants of Chios Island (Greece):
1967 Essential oils of *Origanum onites* L. growing wild in different habitats’, *Ind. Crops Prod.* **2016**,
1968 *82*, 107-113.
- 1969
- 1970 [84] S. Albayrak, A. Aksoy, ‘Phenolic contents and biological activity of endemic *Origanum*
1971 *minutiflorum* grown in Turkey’, *Indian J. Pharm. Educ.* **2019**, *53(1)*, 160-170.
- 1972
- 1973 [85] H. Özbek, Z. Güvenalp, T. Özek, H. G. Sevindik, H. Yuca, K. Ö. Yerdelen, L. Ö. Demirezer,
1974 ‘Chemical composition, antioxidant and anticholinesterase activities of the essential oil of
1975 *Origanum rotundifolium* Boiss. from Turkey’, *Rec. Nat. Prod.* **2017**, *11(5)*, 485-490.
- 1976
- 1977 [86] R. Ribeiro-Santos, D. Carvalho-Costa, C. Cavaleiro, H. S. Costa, T. Gonçalves Albuquerque, M.
1978 Conceição Castilho, F. Ramos, N. R. Melo, A. Sanches-Silva, ‘A novel insight on an ancient
1979 aromatic plant: The rosemary (*Rosmarinus officinalis* L.)’, *Trends Food Sci. Technol.* **2015**, *45*,
1980 355-368.
- 1981
- 1982 [87] R. S. Borges, B. L. Sánchez Ortiz, A. C. Matias Pereira, H. Keita, J. C. Tavares Carvalho,
1983 ‘*Rosmarinus officinalis* essential oil: A review of its phytochemistry, anti-inflammatory activity,
1984 and mechanisms of action involved’, *J. Ethnopharmacol.* **2019**, *229*, 29-45.
- 1985
- 1986 [88] H. Bendif, M. Boudjeniba, M. Djamel Miara, L. Biqiku, M. Bramucci, G. Caprioli, G. Lupidi, L.
1987 Quassinti, G. Sagratini, L. A. Vitali, S. Vittori, F. Maggi, ‘*Rosmarinus eriocalyx*: An alternative
1988 to *Rosmarinus officinalis* as a source of antioxidant compounds’, *Food Chem.* **2017**, *218*, 78-88.
- 1989
- 1990 [89] E. M. Napoli, G. Curcuruto, G. Ruberto, ‘Screening of the essential oil composition of wild
1991 Sicilian rosemary’, *Biochem. Sys. Ecol.* **2010**, *38*, 659-670,
- 1992
- 1993 [90] M. M. Hudaib, K. A. Tawaha, H. S. Hudaib, A. H. Battah, ‘Chemical composition of volatile oil
1994 from the aerial parts of *Rosmarinus officinalis* L. grown in Jordan’, *J. Essent. Oil Bear. Pl.* **2015**,
1995 *18(5)*, 1282-1286.
- 1996

- 1997 [91] S. Grandi, S. Biffi, A. Vecchi, L. Barbanti, 'Assessing essential oil composition of various
1998 Lamiaceae accessions in view of most suitable uses', *J. Essent. Oil Bear. Pl.* **2016**, *19(6)*, 1351-
1999 1367.
- 2000
- 2001 [92] M. B Jemia, R. Tundis, A. Pugliese F. Menichini, F. Senatore, M. Bruno, M. E. Kchouk, M. R.
2002 Loizzo, 'Effect of bioclimatic area on the composition and bioactivity of Tunisian *Rosmarinus*
2003 *officinalis* essential oils', *Nat. Prod. Res.* **2015**, *29(3)*, 213-222.
- 2004
- 2005 [93] W. Yeddes, W. A. Wannas, M. Hammami, M. Smida, A. Chebbi, B. Marzouk, M. S. Tounsi,
2006 'Effect of environmental conditions on the chemical composition and antioxidant activity of
2007 essential oils from *Rosmarinus officinalis* L. growing wild in Tunisia', *J. Essent. Oil Bear. Pl.*
2008 **2018**, *21(4)*, 972-986.
- 2009
- 2010 [94] N. Hendel, E. Napoli, M. Sarri, A. Saija, M. Cristani, A. Nostro, G. Ginestra, G. Ruberto, 'Essential
2011 oil from aerial parts of wild Algerian rosemary: screening of chemical composition, antimicrobial
2012 and antioxidant activities', *J. Essent. Oil Bear. Pl.* **2019**, *22(1)*, 1-17.
- 2013
- 2014 [95] E. M. Napoli, L. Siracusa, A. Saija, A. Speciale, D. Trombetta, T. Tuttolomondo, S. La Bella, M.
2015 Licata, G. Virga, R. Leone, C. Leto, L. Rubino, G. Ruberto, 'Wild Sicilian rosemary:
2016 phytochemical and morphological screening and antioxidant activity evaluation of extracts and
2017 essential oils', *Chem. Biodiversity* **2015**, *12*, 1075-1094.
- 2018
- 2019 [96] I. Bajalan, R. Rouzbahani, A. G. Pirbalouti, F. Maggi, 'Quali-quantitative variation of essential oil
2020 from Iranian rosemary (*Rosmarinus officinalis* L.) accessions according to environmental factors',
2021 *J. Essent. Oil Res.* **2018**, *30(1)*, 16-24.
- 2022
- 2023 [97] I. Bajalan, R. Rouzbahani, A. G. Pirbalouti, F. Maggi, 'Antioxidant and antibacterial activities of
2024 the essential oils obtained from seven Iranian populations of *Rosmarinus officinalis*', *Ind. Crops*
2025 *Prod.* **2017**, *107*, 305-311.
- 2026
- 2027 [98] T. Tuttolomondo, G. Dugo, G. Ruberto, C. Leto, E. M. Napoli, N. Cicero, T. Gervasi, G. Virga,
2028 R. Leone, M. Licata, S. La Bella, 'Study of quantitative and qualitative variations in essential oils
2029 of Sicilian *Rosmarinus officinalis* L.', *Nat. Prod. Res.* **2015**, *29(20)*, 1928-1934.
- 2030
- 2031 [99] M. Jardak, J. Elloumi-Mseddi, S. Aifa, Sami Mnif, 'Chemical composition, anti-biofilm activity
2032 and potential cytotoxic effect on cancer cells of *Rosmarinus officinalis* L. essential oil from
2033 Tunisia', *Lipids Health Dis.* **2017**, *16*, 190.
- 2034
- 2035 [100] G. Li, C. Cervelli, B. Ruffoni, A. Shachter, N. Dudai, 'Volatile diversity in wild populations of
2036 rosemary (*Rosmarinus officinalis* L.) from the Tyrrhenian sea vicinity cultivated under
2037 homogeneous environmental conditions', *Ind. Crops Prod.* **2016**, *84*, 381-390.
- 2038
- 2039 [101] M. Sharifi-Rad, B. Ozcelik, G. Altın, C. Daşkaya-Dikmen, M. Martorell, K. Ramírez-Alarcón, P.
2040 Alarcón-Zapata, M. F. B. Morais-Braga, J. N. P. Carneiro, A. L. A. Borges Leal, H. D. Melo
2041 Coutinho, R. Gyawali, R. Tahergorabi, S. A. Ibrahim, R. Sahrifi-Rad, F. Sharopov, B. Salehi, M.
2042 del Mar Contreras, A. Segura-Carretero, S. Sen, K. Acharya, J. Sharifi-Rad, 'Salvia spp. plants-
2043 from farm to food applications and phytopharmacotherapy', *Trends Food Sci. Tech.* **2018**, *80*, 242-
2044 263.
- 2045
- 2046 [102] G. Asamenew, K. Asres, D. Bisrat, A. Mazumder, P. Lindemann, 'Studies on chemical
2047 compositions, antimicrobial and antioxidant activities of essential oils of *Salvia officinalis* Linn.
2048 grown in two locations of Ethiopia', *J. Essent. Oil Bear. Pl.* **2017**, *20(2)*, 426-437.
- 2049
- 2050 [103] D. Stesevic, M. Ristic, V. Nikolic, M. Nedovic, D. Cakovic, Z. Satovic, 'Chemotype diversity of
2051 indigenous Dalmatian sage (*Salvia officinalis* L.) populations in Montenegro', *Chem. Biodiversity*
2052 **2014**, *11*, 101-114.
- 2053

- 2054 [104] A. Cutillas, A. Carrasco, R. Martinez-Gutierrez, V. Tomas, J. Tudela, ‘*Salvia officinalis* L.
2055 essential oils from Spain: determination of composition, antioxidant capacity, antienzymatic, and
2056 antimicrobial bioactivities’, *Chem. Biodiversity* **2017**, e1700102.
2057
- 2058 [105] H. S. Elshafie, L. Aliberti, M. Amato, V. De Feo, I. Camele, ‘Chemical composition and
2059 antimicrobial activity of chia (*Salvia hispanica* L.) essential oil’, *Eur. Food Res. Technol.* **2018**,
2060 244, 1675-1682.
2061
- 2062 [106] O. Kilic, ‘Chemical composition of four *Salvia* L. species from Turkey: A chemotaxonomic
2063 approach’, *J. Essent. Oil Bear. Pl.* **2016**, 19(1), 229-235.
2064
- 2065 [107] F. Abak, G. Yildiz, V. Atamov, M. Kurkcuoglu, ‘Composition of the essential oil of *Salvia*
2066 *montbretii* Benth. from Turkey’, *Rec. Nat. Prod.* **2018**, 12(5), 426-431.
2067
- 2068 [108] B. Saadatjoo, K. Saeidi, A. Mohammadkhani, H. A. Shirmadi, ‘Assessment of Variation in
2069 Essential Oil Content and Composition within and Among *Salvia* sp. From Southwest Iran’, *J.*
2070 *Essent. Oil. Bear. Pl.* **2018**, 21(1), 237-245.
2071
- 2072 [109] M. Mahdieh, S. M. Talebi, M. Akhiani, ‘Infraspecific essential oil and anatomical variations of
2073 *Salvia nemorosa* L. (Labiatae) populations in Iran’, *Ind. Crops Prod.* **2018**, 123, 35-45.
2074
- 2075 [110] A. Ipek, B. Gurbuz, U. Bingol, F. Geven, M. Uyanik, G. Akgul, K. Rezaeieh, B. C. Senkal,
2076 ‘Comparison of essential oil components of *Salvia forskahlei* L. collected from Nature and
2077 cultivated’, *J. Essent. Oil Bear. Pl.* **2014**, 17(5), 1012-1016.
2078
- 2079 [111] R. Tundis, M. R. Loizzo, M. Bonesi, M. Leporini, F. Menichini, N. G. Passalacqua, ‘Study of
2080 *Salvia fruticosa* Mill subsp. *thomasi* (Lacaita) Brullo, Guglielmo, Pavone & Terrasi, an endemic
2081 Sage of Southern Italy’, *Plant Biosyst.* **2018**, 152(1), 130-141.
2082
- 2083 [112] I. Méndez-Tovar, J. Novak, S. Sponza, B. Herrero, M. C. Asensio-S-Manzanera, ‘Variability in
2084 essential oil composition of wild populations of Labiatae species collected in Spain’, *Ind. Crops*
2085 *Prod.* **2016**, 79, 18-28.
2086
- 2087 [113] K. A. Beladjila, D. Berrehal, A. Al-Aboudi, Z. Semra, H. Al-Jaber, K. Bachari, Z. Kabouche,
2088 ‘Composition and antioxidant, anticholinesterase and antibacterial activities of the essential oil of
2089 *Salvia buchananii* from Algeria’, *Chem. Nat. Compd.* **2018**, 54(3), 581-583.
2090
- 2091 [114] M. Nadir, M. Rasheed, A. Ahmed, V. U. Ahmad, R. B. Tareen, ‘First GC-MS study of essential
2092 oil from *Salvia bucharica*’, *Chem. Nat. Compd.* **2014**, 50(1), 144-146.
2093
- 2094 [115] H. Hejaz, R. Sabbobeh, H. Al-Jaas, A. Jahajha, S. Abu-Lafi, ‘Essential oil secondary metabolites
2095 variation of *Salvia palaestina* leaves growing wild from different locations in Palestine’, *J. Appl.*
2096 *Pharm. Sci.* **2015**, 5(11), 84-89.
2097
- 2098 [116] A. Alimpic, D. Pljevljakusic, K. Savikin, A. Knezevic, M. Curcic, D. Velickovic, T. Stevic, G.
2099 Petrovic, V. Matevski, J. Vukojevic, S. Markovic, P. D. Marin, S. Duletic-Lausevic, ‘Composition
2100 and biological effects of *Salvia ringens* (Lamiaceae) essential oil and extracts’, *Ind. Crops Prod.*
2101 **2015**, 76, 702-709.
2102
- 2103 [117] G. G. Toplan, M. Kurkcuoglu, F. Goger, G. Iscan, H. G. Agalar, A. Mata, K. H. C. Baser, M.
2104 Koyuncu, G. Sariyard, ‘Composition and biological activities of *Salvia veneris* Hedge growing in
2105 Cyprus’, *Ind. Crops Prod.* **2017**, 97, 41-48.
2106
- 2107 [118] F. Medjahed, A. Merouane, A. Saadi, A. Bader, P. L. Cioni, G. Flamini, ‘Chemical profile and
2108 antifungal potential of essential oils from leaves and flowers of *Salvia algeriensis* (Desf.): A
2109 comparative study’, *Chil. J. Agr. Res.* **2016**, 76(2), 195-200.
2110

- 2111 [119] L. Fahed, D. Stien, N. Ouaini, V. Eparvier, M. El Beyrouthy, ‘Chemical diversity and
2112 antimicrobial activity of *Salvia multicaulis* VAHL essential oils’, *Chem. Biodiversity* **2016**, *13*,
2113 591-595.
2114
- 2115 [120] B. Fattahi, V. Nazeri, S. Kalantari, M. Bonfill, M. Fattahi, ‘Essential oil variation in wild-growing
2116 populations of *Salvia reuterana* Boiss. collected from Iran: Using GC–MS and multivariate
2117 analysis’, *Ind. Crops Prod.* **2016**, *81*, 180-190.
2118
- 2119 [121] J. Safaei-Ghomi, R Masoom, F. Jookar Kashi, H. Batooli, ‘Bioactivity of the Essential Oil and
2120 Methanol Extracts of Flowers and Leaves of *Salvia sclarea* L. from Central Iran, *J. Essent. Oil*
2121 *Bear. Pl.* **2016**, *19(4)*, 885-896.
2122
- 2123 [122] E. E. Hanlidou, R. Karousou, D. Lazari, ‘Essential-oil diversity of *Salvia tomentosa* Mill. in
2124 Greece”, *Chem. Biodiversity* **2014**, *11*, 1205-1215.
2125
- 2126 [123] N. Tan, S. Yazıcı-Tütüniş, Y. Yeşil, B. Demirci, E.Tan, ‘Antibacterial activities and composition
2127 of the essential oils of *Salvia sericeo-tomentosa* varieties”, *Rec. Nat. Prod.* **2017**, *11(5)*, 456-461.
2128
- 2129 [124] A. Kaya, M. Dinç, S. Doğu, B. Demirci, ‘Compositions of essential oils of *Salvia adenophylla*,
2130 *Salvia pilifera*, and *Salvia viscosa* in Turkey’, *J. Ess. Oil Res.* **2017**, *29(3)*, 233-239.
2131
- 2132 [125] L. Riccobono, A. Maggio, S. Rosselli, V. Iardi, F. Senatore, M. Bruno, ‘Chemical composition of
2133 volatile and fixed oils from of *Salvia argentea* L. (Lamiaceae) growing wild in Sicily’, *Nat. Prod.*
2134 *Res.* **2016**, *30(1)*, 25-34.
2135
- 2136 [126] E. Emadipoor, M. Jamzad, K. Ghaffari, B. Ghadam, Z. Jamzad, ‘Essential oil composition, total
2137 phenolic and flavonoid contents, and biological activities of *Salvia aristata* Aucher ex Benth.
2138 extracts’, *J. Essent. Oil Bear. Pl.* **2016**, *19(6)*, 1426-1434.
2139
- 2140 [127] M. E. Khiyari, A. Kasrati, C. A. Jamali, S. Zeroual, M. Markouk, K. Bekkouche, H. Wohlmuth,
2141 D. Leach, A. Abbad, ‘Chemical composition, antioxidant and insecticidal properties of essential
2142 oils from wild and cultivated *Salvia aucheri* subsp. *Blancoana* (Webb. & Helder)), an endemic,
2143 threatened medicinal plant in Morocco’, *Ind. Crops Prod.* **2014**, *57*, 106-109.
2144
- 2145 [128] T. A. Ibrahim, ‘Chemical composition and antimicrobial activity of essential oil of *Salvia bicolor*
2146 Desf. growing in Egypt”, *J. Essent. Oil Bear. Pl.* **2014**, *17(1)*, 104-111.
2147
- 2148 [129] B. Najar, L. Pistelli, C. Cervelli, G. Fico, C. Giuliani, ‘*Salvia broussonetii* Benth.: aroma profile
2149 and micromorphological analysis’, *Nat. Prod. Res.* **2018**, *32(14)*, 1660-1668.
2150
- 2151 [130] S. Krimat, T. Dob, M. Toumi, L. Lamari, D. Dahmane, ‘Chemical composition, antimicrobial and
2152 antioxidant activities of essential oil of *Salvia chudaei* Batt. et Trab. endemic plant from Algeria’,
2153 *J. Essent. Oil. Res.* **2015**, *27(5)*, 447-453.
2154
- 2155 [131] E. Rapposelli, S. Melito, G. G. Barmina, M. Foddai, E. Azara, G. M. Scarpa, ‘AFLP fingerprinting
2156 and essential oil profiling of cultivated and wild populations of Sardinian *Salvia desoleana*’, *Genet.*
2157 *Resour. Crop Evol.* **2015**, *62*, 959-970.
2158
- 2159 [132] A. Sonboli, P. Salehi, S. Ghareh-naghadeh, ‘Chemical variability in the essential oil composition of
2160 *Salvia hypoleuca*, an endemic species from Iran’, *J. Essent. Oil Res.* **2016**, *28(5)*, 421-427.
2161
- 2162 [133] S. Fang, X. Xing, P. Lai, J. Huang, ‘Chemical composition and antioxidant activity of the essential
2163 oil from *Salvia kiangsiensis*’, *Chem. Nat. Compd.* **2018**, *54(3)*, 591-592.
2164
- 2165 [134] S. N. Moadeli, V. Rowshan, A. Aboutalebi, ‘Comparison of *Salvia lachnocalyx* Hedge. essential
2166 oil components in wild and field population’, *Res. J. Pharm. Biol. Chem. Sci.* **2014**, *5(2)*, 903-906.
2167

- 2168 [135] M. Nekoei, M. Mohammadhosseini, 'Chemical composition of the essential oils and volatiles of
2169 *Salvia leriifolia* by three different extraction methods prior to gas chromatographic-mass
2170 spectrometric determination: comparison of HD with SFME and HS-SPME", *J. Essent. Oil Bear.*
2171 *Pl.* **2017**, *20(2)*, 410-425.
2172
- 2173 [136] K. Morteza-Semnani, M. Saedi, M. Akbarzadeh, 'Chemical composition of the essential oil of
2174 *Salvia limbata* C. A. Mey", *J. Essent. Oil Bear. Pl.* **2014**, *17(4)*, 623-628.
2175
- 2176 [137] G. Dehghan, S. Torbati, R. Mohammadian, A. Movafeghi, A. H. Talebpour, 'Essential oil
2177 composition, total phenol and flavonoid contents and antioxidant activity of *Salvia sahendica* at
2178 different developmental stages", *J. Essent. Oil Bear. Pl.* **2018**, *21(4)*, 1030-1040.
2179
- 2180 [138] M. B. Bahadori, H. Valizadeh, M. M. Farimani, 'Chemical composition and antimicrobial activity
2181 of the volatile oil of *Salvia santolinifolia* Boiss. from Southeast of Iran", *Pharmaceutical Sciences*
2182 **2016**, *22*, 42-48.
2183
- 2184 [139] P. H. H. Gavyar, H. Amiri, 'Chemical composition of essential oil and antioxidant activity of
2185 *Salvia sclareopsis* an endemic species from Iran', *J. Essent. Oil Bear. Pl.* **2018**, *21(4)*, 1138-1145.
2186
- 2187 [140] J. Asgarpanah, E. Oveyli, S. Alidoust, 'Volatile components of the endemic species *Salvia sharifii*
2188 Rech. f. & Esfand", *J. Essent. Oil Bear. Pl.* **2017**, *20(2)*, 578-582.
2189
- 2190 [141] B. Salehi, M. S. Abu-Darwish, A. H. Tarawneh, C. Cabral, A. V. Gadetskaya, L. Salgueiro, T.
2191 Hosseinabadi, S. Rajabi, W. Chanda, M. Sharifi-Rad, R. B. Mulaudzi, S. A. Ayatollahi, F.
2192 Kobarfard, D. K Arserim-Uçar, J. Sharifi-Rad, A. Ata, N. Baghalpour, M. del Mar Contreras,
2193 'Thymus spp. plants - Food applications and phytopharmacy properties', *Trends Food Sci. Tech.*
2194 **2019**, *85*, 287-306.
2195
- 2196 [142] B. Tohidi, M. Rahimmalek, H. Trindade, 'Review on essential oil, extracts composition, molecular
2197 and phytochemical properties of Thymus species in Iran', *Ind. Crops Prod.* **2019**, *134*, 89-99.
2198
- 2199 [143] H. Trindade, L. G. Pedro, A. C. Figueiredo, J. G. Barroso, 'Chemotypes and terpene synthase
2200 genes in *Thymus* genus: State of the art', *Ind. Crops Prod.* **2018**, *124*, 530-547.
2201
- 2202 [144] E. M. Napoli, G. Curcuruto, G. Ruberto, 'Screening of the essential oil composition of wild
2203 Sicilian thyme', *Biochem. Sys. Ecol.* **2010**, *38*, 816-822.
2204
- 2205 [145] A. Saija, A. Speciale, D. Trombetta, C. Leto, T. Tuttolomondo, S. La Bella, M. Licata, G. Virga,
2206 G. Buonsangue, M. C. Gennaro, E. Napoli, L. Siracusa, G. Ruberto, 'Phytochemical, ecological
2207 and antioxidant evaluation of wild Sicilian Thyme: *Thymbra capitata* (L.) Cav.', *Chem.*
2208 *Biodiversity* **2016**, *13*, 1-15.
2209
- 2210 [146] S. Tammar, N. Salem, I. B. Rebey, J. Sriti, M. Hammami, S. Khammassi, B. Marzouk, R. Ksouri,
2211 K. Msaad, 'Regional effect on essential oil composition and antimicrobial activity of *Thymus*
2212 *capitatus* L.', *J. Essent. Oil Res.* **2019**, *31(2)*, 129-137.
2213
- 2214 [147] A. -B. Cutillas, A. Carrasco, R. Martinez-Gutierrez, V. Tomas, J. Tudela, 'Thyme essential oils
2215 from Spain: Aromatic profile ascertained by GC-MS, and their antioxidant, anti-lipoxygenase and
2216 antimicrobial activities', *J. Food Drug Anal.* **2018**, *26(2)*, 529-544.
2217
- 2218 [148] J. Delgado-Adamez, M. Garrido, M. E. Bote, M. C. Fuentes-Perez, J. Espino, D. Martin-Vertedor,
2219 'Chemical composition and bioactivity of essential oils from flower and fruit of *Thymbra capitata*
2220 and *Thymus* species', *J. Food Sci. Technol.* **2017**, *54(7)*, 1857-1865.
2221
- 2222 [149] A. Stefanaki, C. M. Cook, T. Lanaras, S. Kokkini, 'Essential oil variation of *Thymbra spicata* L.
2223 (Lamiaceae), an East Mediterranean "oregano" herb', *Biochem. Sys. Ecol.* **2018**, *80*, 63-69.
2224

- 2225 [150] K. Bączek, E. Pióro-Jabrucka, O. Kosakowska, Z. Węglarz, 'Intraspecific variability of wild thyme
2226 (*Thymus serpyllum* L.) occurring in Poland', *J. Appl. Res. Med. Aromat. Plants* **2019**, *12*, 30-35.
2227
- 2228 [151] D. Damtie, C. Braunberger, J. Conrad, Y. Mekonnen, U. Beifuss, 'Composition and
2229 hepatoprotective activity of essential oils from Ethiopian thyme species (*Thymus serrulatus* and
2230 *Thymus schimperi*)', *J. Ess. Oil Res.* **2019**, *31(2)*, 120-128.
2231
- 2232 [152] J. Arsenijević, M. Drobac, I. Šoštarić, R. Jevđović, J. Živković, S. Ražić, Đ. Moravčević, Z.
2233 Maksimović, 'Comparison of essential oils and hydromethanol extracts of cultivated and wild
2234 growing *Thymus pannonicus* All.', *Ind. Crops Prod.* **2019**, *130*, 162-169.
2235
- 2236 [153] A. Shokrgoo, M. Madandoust, 'Effect of harvest time on essential oil content and chemical
2237 composition of *Origanum vulgare* (L.) from Iran', *J. Essent. Oil Bear. Pl.* **2018**, *21(6)*, 1682-1686.
2238
- 2239 [154] A. Bouyahya, N. Dakka, A. Talbaoui, A. Et-Touysa, H. El-Boury, J. Abrini, Y. Bakri, 'Correlation
2240 between phenological changes, chemical composition and biological activities of the essential oil
2241 from Moroccan endemic Oregano (*Origanum compactum* Benth)', *Ind. Crops Prod.* **2017**, *108*,
2242 729-737.
2243
- 2244 [155] R. Nurzyńska-Wierdak, G. Zawislak, R. Kowalski, "The content and composition of essential oil
2245 of *Origanum majorana* L. grown in Poland depending on harvest time and method of raw material
2246 preparation", *J. Essent. Oil Bear. Pl.* **2015**, *18(6)*, 1482-1489.
2247
- 2248 [156] K. Mechergui, W. Jaouadi, J. P. Coelho, M. L. Khouja, 'Effect of harvest year on production,
2249 chemical composition and antioxidant activities of essential oil of oregano (*Origanum vulgare*
2250 subsp *glandulosum* (Desf.) Ietswaart) growing in North Africa', *Ind. Crops Prod.* **2016**, *90*, 32-
2251 37.
2252
- 2253 [157] C. N. Hassiotis, F. Ntana, D. M. Lazari, S. Poullos, K. E. Vlachonasios, 'Environmental and
2254 developmental factors affect essential oil production and quality of *Lavandula angustifolia* during
2255 flowering period', *Ind. Crops Prod.* **2014**, *62*, 359-366.
2256
- 2257 [158] E. Sarrou, S. Martens, P. Chatzopoulou, 'Metabolite profiling and antioxidative activity of Sage
2258 (*Salvia fruticosa* Mill.) under the influence of genotype and harvesting period', *Ind. Crops Prod.*
2259 **2016**, *94*, 240-250.
2260
- 2261 [159] R. S. Verma, R. C. Padalia, A. Chauhan, 'Harvesting season and plant part dependent variations in
2262 the essential oil composition of *Salvia officinalis* L. grown in northern India', *J. Herb. Med.* **2015**,
2263 *5*, 165-171.
2264
- 2265 [160] M. B. Farhat, M. J. Jordan, R. Chaouch-Hamada, A. Landoulsi, J. A. Sotomayor, 'Phenophase
2266 effects on sage (*Salvia officinalis* L.) yield and composition of essential oil', *J. Appl. Res. Med.*
2267 *Aromat. Plants* **2016**, *3*, 87-93.
2268
- 2269 [161] M. B. Farhat, J. A. Sotomayor, M. J. Jordan, 'Salvia verbenaca L. essential oil: Variation of yield
2270 and composition according to collection site and phenophase', *Biochem. Sys. Ecol.* **2019**, *82*, 35-
2271 43.
2272
- 2273 [162] M. Fumiere Lemos, M. Fumiere. Lemos, H. Poltronieri Pacheco, D. Coutinho Endringer, R.
2274 Scherer, 'Seasonality modifies rosemary's composition and biological activity', *Ind. Crops Prod.*
2275 **2015**, *70*, 41-47.
2276
- 2277 [163] S. Rguez, K. Msaada, M. Daami-Remadi, I. Chayeb, I. Bettaieb Rebey, M. Hammami, A. Laarif,
2278 I. Hamrouni-Sellami, 'Chemical composition and biological activities of essential oils of *Salvia*
2279 *officinalis* aerial parts as affected by diurnal variations', *Pl. Biosystems* **2019**, *153(2)*, 264-272.
2280
- 2281 [164] C. Mapes, Y. Xu, 'Photosynthesis, vegetative habit and culinary properties of sage (*Salvia*
2282 *officinalis*) in response to low-light conditions', *Can. J. Plant Sci.* **2014**, *94*, 881-889.

- 2283 [165] V. Davidenco, J. A. Arguello, M. B. Piccardi, C. R. C. Vega, 'Day length modulates precocity and
2284 productivity through its effect on developmental rate in *Origanum vulgare* ssp.', *Sci. Hort.* **2017**,
2285 *218*, 164-170.
2286
- 2287 [166] S. Shiwakoti, V. D. Zheljzkov, V. Schlegel, C. L. Cantrell, 'Growing spearmint, thyme, oregano,
2288 and rosemary in Northern Wyoming using plastic tunnels', *Ind. Crops Prod.* **2016**, *94*, 251-258.
2289
- 2290 [167] H. M. Lajayer, H. Zakizadeh, Y. Hamidoghli, M. H. Bigluei, E. Chamani, 'Ornamental potential
2291 and freezing tolerance of six *Thymus* spp. species as ground-covering plants in the landscape',
2292 *Zemdirbyste-Agriculture* **2018**, *105(1)*, 79-88.
2293
- 2294 [168] A. Carrubba, "Sustainable fertilization in medicinal and aromatic plants" in Á. Máthé (Ed.)
2295 Medicinal and aromatic plants of the world, Springer, London, 2015, pp. 187-203.
- 2296 [169] Ö.A. Asci, H. Deveci, A. Erdeger, K. N. Özdemir, T. Demirci, N. G. Baydar, 'Brassinosteroids
2297 Promote Growth and Secondary Metabolite Production in Lavandin (*Lavandula intermedia*
2298 *Emeric ex Loisel.*)', *J. Essent. Oil Bear. Pl.* **2019**, *22(1)*, 254-263.
2299
- 2300 [170] M. H. Shabana, L. K. Balbaa, I. M. Talaat, 'Effect of foliar applications of *Zingiber officinale*
2301 extracts on *Origanum majorana*', *J. Herbs Spices Med. Plants* **2017**, *23(2)*, 89-97.
2302
- 2303 [171] R. Pavela, M. Žabka, N. Vrchotová, J. Tříška, 'Effect of foliar nutrition on the essential oil yield
2304 of Thyme (*Thymus vulgaris* L.)', *Ind. Crops Prod.* **2018**, *112*, 762-765.
2305
- 2306 [172] M. Majdi, A. Malekzadeh-Mashhady, A. Maroufi, C. Crocoll, 'Tissue-specific gene-expression
2307 patterns of genes associated with thymol/carvacrol biosynthesis in thyme (*Thymus vulgaris* L.) and
2308 their differential changes upon treatment with abiotic elicitors', *Plant Physiol. Biochem.* **2017**,
2309 *115*, 152-162.
2310
- 2311 [173] L. V. Mehrabani, M. B. Hassanpouraghdam, T. Shamsi-Khota, 'The effects of common and nano-
2312 zinc foliar application on the alleviation of salinity stress in *Rosmarinus officinalis* L.', *Acta Sci.*
2313 *Pol. Hortorum Cultus* **2018**, *17(6)*, 65-73.
2314
- 2315 [174] M. A. El-ESawi, H. O. Elansary, N. A. El-Shanhorey, A. M. E. Abdel-Hamid, H. M. Ali, M. S.
2316 Elshikh, 'Salicylic acid-regulated antioxidant mechanisms and gene expression enhance rosemary
2317 performance under saline conditions', *Front. Physiol.* **2017**, *8*, 716.
2318
- 2319 [175] R. Dehghani Bidgoli, N. Azarnezhad, M. Akhbari, M. Ghorbani, 'Salinity stress and PGPR effects
2320 on essential oil changes in *Rosmarinus officinalis* L.', *Agric. Food Secur.* **2019**, *8:2*.
2321
- 2322 [176] M. Valifard, S. Mohsenzadeh, B. Kholdebarin, V. Rowshan, 'Effects of salt stress on volatile
2323 compounds, total phenolic content and antioxidant activities of *Salvia mirzayanii*', *S. Afr. J. Bot.*
2324 **2014**, *93*, 92-97.
2325
- 2326 [177] M. Valifard, S. Mohsenzadeh, B. Kholdebarin, V. Rowshan, A. Niazi, A. Moghadam, 'Effect of
2327 salt stress on terpenoid biosynthesis in *Salvia mirzayanii*: from gene to metabolite', *J. Hort. Sci.*
2328 *Biotechnol.* **2019**, *94(3)*, 389-399.
2329
- 2330 [178] A. Chrysargyris, E. Michailidi N. Tzortzakis, 'Physiological and biochemical responses of
2331 *Lavandula angustifolia* to salinity under mineral foliar application', *Front. Plant Sci.* **2018**, *9*, 489.
2332
- 2333 [179] L.V. Mehrabani, R. Valizadeh Kamran, M. Bagher Hassanpouraghdam, M. Pessarakli, 'Zinc
2334 sulfate foliar application effects on some physiological characteristics and phenolic and essential
2335 oil contents of *Lavandula stoechas* L. under sodium chloride (NaCl) salinity conditions', *Commun.*
2336 *Soil Sci. Plant Anal.* **2017**, *48(16)*, 1860-1867.
2337
- 2338 [180] N. E. Hancioglu, A. Kurunc, I. Tontul, A. Topuz, 'Irrigation water salinity effects on oregano
2339 (*Origanum onites* L.) water use, yield and quality parameters', *Sci. Hort.* **2019**, *247*, 327-334.

- 2340 [181] M. R. Morshedloo, L. E. Cracker, A. Salami, V. Nazeri, H. Sang, F. Maggi, 'Effect of prolonged
2341 water stress on essential oil content, compositions and gene expression patterns of mono- and
2342 sesquiterpene synthesis in two oregano (*Origanum vulgare* L.) subspecies', *Plant Physiol.*
2343 *Biochem.* **2017**, *111*, 119-128.
- 2344 [182] Z. E. Bistgani, S. A. Siadat, A. Bakhshandeh, A. G. Pirbalouti, M. Hashemi, 'Morpho-
2345 physiological and phytochemical traits of (*Thymus daenensis* Celak.) in response to deficit
2346 irrigation and chitosan application', *Acta Physiol. Plant.* **2017**, *39*, 231.
- 2347 [183] A. A. Al-Ghamdi, 'Marjoram physiological and molecular performance under water stress and
2348 chitosan treatment', *Acta Physiol. Plant.* **2019**, *41*, 44.
- 2350 [184] P. García-Caparrós, M. J. Romero, A. Llanderal, P. Cermeño, M. T. Lao, M. L. Segura, 'Effects
2351 of drought stress on biomass, essential oil content, nutritional parameters, and costs of production
2352 in six Lamiaceae species', *Water* **2019**, *11*, 573.
- 2353 [185] A. Chrysargyris, C. Panayiotou, N. Tzortzakis, 'Nitrogen and phosphorus levels affected plant
2354 growth, essential oil composition and antioxidant status of lavender plant Mill.', *Ind. Crops Prod.*
2355 **2016**, *83*, 577-586.
- 2356 [186] M. Caser, W. Chitarra, F. D'Angiolillo, I. Perrone, S. Demasi, C. Lovisolò, L. Pistelli, L. Pistelli,
2357 V. Scariot, 'Drought stress adaptation modulates plant secondary metabolite production in *Salvia*
2358 *dolomitica* Codd.', *Ind. Crops Prod.* **2019**, *129*, 85-96.
- 2359 [187] A. Radwan, M. Kleinwächter, D. Selmar, 'Impact of drought stress on specialised metabolism:
2360 Biosynthesis and the expression of monoterpene synthases in sage (*Salvia officinalis*)',
2361 *Phytochemistry* **2017**, *141*, 20-26.
- 2362 [188] M. Caser, F. D'Angiolillo, W. Chitarra, C. Lovisolò, B. Ruffoni, L. Pistelli, L. Pistelli, V. Scariot,
2363 'Ecophysiological and phytochemical responses of *Salvia sinaloensis* Fern. to drought stress',
2364 *Plant Growth Reg.* **2018**, *84*, 383-394.
- 2365 [189] E. Ninou, K. Paschalidis, I. Mylonas, 'Essential oil responses to water stress in Greek oregano
2366 populations', *J. Essent. Oil Bear. Pl.* **2017**, *20(1)*, 12-23.
- 2367 [190] H. Mohammadi, F. Amirikia, M. Ghorbanpour, F. Fatehi, H. Hashempour, 'Salicylic acid induced
2368 changes in physiological traits and essential oil constituents in different ecotypes of *Thymus*
2369 *kotschyanus* and *Thymus vulgaris* under well-watered and water stress conditions', *Ind. Crops*
2370 *Prod.* **2019**, *129*, 561-574.
- 2371 [191] M. Askary, M. Ali Behdani, S. Parsa, S. Mahmoodi, M. Jamialahmadi, 'Water stress and manure
2372 application affect the quantity and quality of essential oil of *Thymus daenensis* and *Thymus*
2373 *vulgaris*', *Ind. Crops Prod.* **2018**, *111*, 336-344.
- 2374 [192] A. Pirzad, S. Mohammadzadeh, 'Water use efficiency of three mycorrhizal Lamiaceae species
2375 (*Lavandula officinalis*, *Rosmarinus officinalis* and *Thymus vulgaris*)', *Agric. Water Manag.* **2018**,
2376 *204*, 1-10.
- 2377 [193] A. Abdollahi Arpanahi, M. Feizian, 'Arbuscular mycorrhizae alleviate mild to moderate water
2378 stress and improve essential oil yield in thyme', *Rhizosphere* **2019**, *9*, 93-96.
- 2379 [194] M. Govahi, A. Ghalavand, F. Nadjafi, A. Sorooshzadeh, 'Comparing different soil fertility systems
2380 in Sage (*Salvia officinalis*) under water deficiency', *Ind. Crops Prod.* **2015**, *74*, 20-27.
- 2381 [195] F. Gerami, P. R. Moghaddam, R. Ghorbani, A. Hassani, 'Effects of irrigation intervals and organic
2382 manure on morphological traits, essential oil content and yield of oregano (*Origanum vulgare* L.)',
2383 *An. Acad. Bras. Ciênc.* **2016**, *88(4)*, 2375-2385.
- 2384
2385
2386
2387
2388
2389
2390
2391
2392
2393
2394
2395
2396
2397

- 2398 [196] N. Vosoughi, M. Gomarian, A. Ghasemi Pirbalouti, S. Khaghani, F. Malekpoor, 'Essential oil
2399 composition and total phenolic, flavonoid contents, and antioxidant activity of sage (*Salvia*
2400 *officinalis* L.) extract under chitosan application and irrigation frequencies', *Ind. Crops Prod.*
2401 **2018**, *117*, 366-374.
- 2402 [197] N. Boke Rioba, F. Musyoka Itulya, M. Saidi, N. Dudai, N. Bernstein, 'Effects of nitrogen,
2403 phosphorus and irrigation frequency on essential oil content and composition of sage (*Salvia*
2404 *officinalis* L.)', *J. Appl. Res. Med. Aromat. Plants* **2015**, *2*, 21-29.
- 2405 [198] M. S. Ali-Shtayeh, R. M. Jamous, S. Y. Abu-Zaitoun, R. J. Akkawi, S. R. Kalbouneh, N. Dudai,
2406 N. Bernstein, 'Secondary treated effluent irrigation did not impact chemical composition, and
2407 enzyme inhibition activities of essential oils from *Origanum syriacum* var. *syriacum*', *Ind. Crops*
2408 *Prod.* **2018**, *111*, 775-786.
- 2409 [199] W. Tarraf, C. Ruta, F. De Cillis, A. Tagarelli, L. Tendone, G. De Mastro, 'Effects of mycorrhiza
2410 on growth and essential oil production in selected aromatic plants', *Ital. J. Agron.* **2015**, *10*, 633.
- 2411 [200] W. Tarraf, C. Ruta, A. Tagarelli, F. De Cillis, G. De Mastro, 'Influence of arbuscular mycorrhizae
2412 on plant growth, essential oil production and phosphorus uptake of *Salvia officinalis* L.', *Ind.*
2413 *Crops Prod.* **2017**, *102*, 144-153.
- 2414 [201] M. Hristozkova, M. Geneva, I. Stancheva, M. Boychinova, E. Djonova, 'Aspects of mycorrhizal
2415 colonization in adaptation of sweet marjoram (*Origanum majorana* L.) grown on industrially
2416 polluted soil', *Turk. J. Biol.* **2015**, *39*, 461-468.
- 2417 [202] L. Pistelli, R. V. Bandera Reidel, F. Parri, E. Morelli, L. Pistelli, 'Chemical composition of
2418 essential oil from plants of abandoned mining site of Elba island', *Nat. Prod. Res.* **2019**, *33(1)*,
2419 143-147.
- 2420 [203] I. Stancheva, M. Geneva, Y. Markovska, N. Tzvetkova, I. Mitova, M. Todorova, P. Petrov, 'A
2421 comparative study on plant morphology, gas exchange parameters, and antioxidant response of
2422 *Ocimum basilicum* L. and *Origanum vulgare* L. grown on industrially polluted soil', *Turk. J. Biol.*
2423 **2014**, *38*, 89-102.
- 2424 [204] F. Araniti, M. Landi, A. Lupini, F. Sunseri, L. Guidi, M. R. Abenavoli, '*Origanum vulgare*
2425 essential oils inhibit glutamate and aspartate metabolism altering the photorespiratory pathway in
2426 *Arabidopsis thaliana* seedlings', *J. Plant Phys.* **2018**, *231*, 297-309.
- 2427 [205] M. Atak, K. Mavi, I. Uremis, 'Bio-Herbicidal effects of oregano and rosemary essential oils on
2428 germination and seedling growth of bread wheat cultivars and weeds', *Rom. Biotech. Lett.* **2016**,
2429 *21(1)*, 11149-11159.
- 2430 [206] M. D. Ibanez, M. A. Blazquez, 'Herbicidal value of essential oils from oregano-like flavor
2431 species', *Food Agric. Immunol.* **2017**, *28(6)*, 1168-1180.
- 2432 [207] R. Fouad, D. Bousta, A. El Ouali Lalami, F. Ouazzani Chahdi, I. Amri, B. Jamoussi, H. Greche,
2433 "Chemical composition and herbicidal effects of essential oils of *Cymbopogon citratus* (DC) Stapf,
2434 *Eucalyptus cladocalyx*, *Origanum vulgare* L and *Artemisia absinthium* L. cultivated in Morocco",
2435 *J. Essent. Oil Bear. Pl.* **2015**, *18(1)*, 112-123.
- 2436 [208] D. Grul'ová, M. Pl'uchtova, J. Fejér, L. De Martina, L. Caputo, V. Sedlák, V. De Feo, 'Influence
2437 of six essential oils on invasive *Solidago canadensis* L. seed germination', *Nat. Prod. Res.* **2019**,
2438 <https://doi.org/10.1080/14786419.2018.1552694>.
- 2439 [209] M. Alipour, M. J. Saharkhiz, 'Phytotoxic activity and variation in essential oil content and
2440 composition of Rosemary (*Rosmarinus officinalis* L.) during different phenological growth
2441 stages', *Biocat. Agric. Biotech.* **2016**, *7*, 271-278.
- 2442
2443
2444
2445
2446
2447
2448
2449
2450
2451
2452
2453
2454
2455

- 2456 [210] M. Alipour, M. J. Saharkhiz, M. Niakousari, M.S. Damyeh, ‘Phytotoxicity of encapsulated
2457 essential oil of rosemary on germination and morphophysiological features of amaranth and radish
2458 seedlings’, *Sci. Hort.* **2019**, *243*, 131-139.
- 2460 [211] A. Synowiec, D. Kalemba, E. Drozdek, J. Bocianowski, ‘Phytotoxic potential of essential oils from
2461 temperate climate plants against the germination of selected weeds and crops’, *J. Pest Sci.* **2017**,
2462 *90*, 407-419.
- 2463 [212] E. Alexa, R. M. Sumalan, C. Danciu, D. Obistoiu, M. Negrea, M. A. Poiana, C. Rus, I. Radulov,
2464 G. Pop, C. Dehelan, ‘Synergistic antifungal, allelopathic and anti-proliferative potential of *Salvia*
2465 *officinalis* L., and *Thymus vulgaris* L. essential oils’, *Molecules* **2018**, *23*, 185.
- 2466 [213] D. Dris, F. Tine-Djebbar, N. Soltani, ‘*Lavandula dentata* essential oils: chemical composition and
2467 larvicidal activity against *Culiseta longiareolata* and *Culex pipiens* (Diptera: Culicidae)’, *Afr.*
2468 *Entomol.* **2017**, *25(2)*, 387-394.
- 2470 [214] L.F. Julio, C. E. Díaz, N. Aissani, F. Valcarcel, J. Burillo, S. Olmeda, A. González-Coloma,
2471 ‘Ixadical compounds from pre-domesticated *Lavandula luisieri*’, *Ind. Crops Prod.* **2017**, *110*,
2472 83-87.
- 2473 [215] S. Bedini, G. Flamini, F. Cosci, R. Ascrizzi, M. C. Echeverria, E. V. Gomez, L. Guidi, M. Landi,
2474 A. Lucchi, B. Conti, ‘Toxicity and oviposition deterrence of essential oils of *Clinopodium*
2475 *nubigenum* and *Lavandula angustifolia* against the myiasis inducing blowfly *Lucilia sericata*’,
2476 *PlosOne* **2019**, *14(2)*, e0212576.
- 2477 [216] E. Nicolás Jesser, J. O. Werdin-González, A. P. Murray, A. A. Ferrero, ‘Efficacy of essential oils
2478 to control the Indian meal moth, *Plodia interpunctella* (Hübner) (Lepidoptera: Pyralidae)’, *J. Asia-*
2479 *Pac. Entomol.* **2017**, *20*, 1122-1129.
- 2480 [217] C. G. Yi, T. T. Hieu, S. H. Lee, B. Choi, M. Kwond, Y. Ahn, ‘Toxicity of *Lavandula angustifolia*
2481 oil constituents and spray formulations to insecticide-susceptible and pyrethroid-resistant *Plutella*
2482 *xylostella* and its endoparasitoid *Cotesia glomerata*’, *Pest Manag. Sci.* **2016**, *72*, 1202–1210.
- 2483 [218] G. S. Germinara, M. G. Di Stefano, L. De Acutis, S. Pati, S. Delfine, A. De Cristofaro, G. Rotundo,
2484 ‘Bioactivities of *Lavandula angustifolia* essential oil against the stored grain pest *Sitophilus*
2485 *granaries*’, *Bull. Insectology* **2017**, *70(1)*, 129-138.
- 2486 [219] B. S. Badreddine, E. Olfa, D. Samir, C. Hnia, B. J. M. Lahbib, ‘Chemical composition of
2487 *Rosmarinus* and *Lavandula* essential oils and their insecticidal effects on *Orgyia trigotephra*
2488 (Lepidoptera, Lymantriidae)’, *Asian Pac. J. Trop. Med.* **2015**, 98-103.
- 2489 [220] G. Ortiz de Elguea-Culebras, R. Sanchez-Vioque, M. I. Berruga, D. Herraiz-Penalver, A.
2490 Gonzalez-Coloma, M. Fe Andres, O. Santana-Merida, ‘Biocidal potential and chemical
2491 composition of industrial essential oils from *Hyssopus officinalis*, *Lavandula x intermedia* var.
2492 SUPER, and *Santolina chamaecyparissus*’, *Chem. Biodiversity* **2018**, *15*, e1700313.
- 2493 [221] S. Djebir, S. Ksouri, M. Trigui, S. Tounsi, A. Boumaaza, Y. Hedef, A. Benakhla, ‘Chemical
2494 composition and acaricidal activity of the essential oils of some plant species of Lamiaceae and
2495 Myrtaceae against the vector of tropical bovine theileriosis: *Hyalomma scupense* (syn. *Hyalomma*
2496 *detrutum*)’, *Biomed. Res. Int.* **2019**, 7805467.
- 2497 [222] A. La Pergola, C. Restuccia, E. Napoli, S. Bella, S. Brighina, A. Russo, P. Suma, ‘Commercial
2498 and wild Sicilian *Origanum vulgare* essential oils: chemical composition, antimicrobial activity
2499 and repellent effects’, *J. Essent. Oil Res.* **2017**, *29(6)*, 451-460.
- 2500 [223] J. F. Carroll, B. Demirci, M. Kramer, U. R. Bernier, N. M. Agramonte, K. H. Can Baser, N.
2501 Tabanca, ‘Repellency of the *Origanum onites* L. essential oil and constituents to the lone star tick
2502 and yellow fever mosquito’, *Nat. Prod. Res.* **2017**, *31(18)*, 2192-2197.

- 2514 [224] A. Giatropoulos, A. Kimbaris, A. Michaelakis, D. P. Papachristos, M. G. Polissiou, N.
2515 Emmanouel, 'Chemical composition and assessment of larvicidal and repellent capacity of 14
2516 Lamiaceae essential oils against *Aedes albopictus*', *Parasitol. Res.* **2018**, *117*, 1953-1964.
2517
- 2518 [225] G. Benelli, R. Pavela, R. Petrelli, L. Cappellacci, F. Bartolucci, A. Canale, F. Maggi, '*Origanum*
2519 *syriacum* subsp. *syriacum*: From an ingredient of Lebanese 'manoushe' to a source of effective
2520 and eco-friendly botanical insecticides', *Ind. Crops Prod.* **2019**, *134*, 26-32.
2521
- 2522 [226] M. Govindarajan, S. Kadaikunnan, N. S. Alharbi, G. Benelli, 'Acute toxicity and repellent activity
2523 of the *Origanum scabrum* Boiss. & Heldr. (Lamiaceae) essential oil against four mosquito vectors
2524 of public health importance and its biosafety on non-target aquatic organisms', *Environ. Sci.*
2525 *Pollut. Res.* **2016**, *23*, 23228–23238.
2526
- 2527 [227] H. Ramzi, M. R. Ismaili, M. Aberchane, S. Zaanoun, 'Chemical characterization and acaricidal
2528 activity of *Thymus satureioides* C. & B. and *Origanum elongatum* E. & M. (Lamiaceae) essential
2529 oils against *Varroa destructor* Anderson & Trueman (Acari: Varroidae)', *Ind. Crops Prod.* **2017**,
2530 *108*, 201-207.
2531
- 2532 [228] M. Khoobdel, S. M. Ahsaei, M. Farzaneh, 'Insecticidal activity of polycaprolactone nanocapsules
2533 loaded with *Rosmarinus officinalis* essential oil in *Tribolium castaneum* (Herbst)', *Entomol. Res.*
2534 **2017**, *47*, 175-184.
2535
- 2536 [229] L. Bendifallah, R. Belguendouz, L. Hamoudi, K. Arab, 'Biological activity of the *Salvia officinalis*
2537 L. (Lamiaceae) essential oil on *Varroa destructor* infested honeybees", *Plants* **2018**, *7*, 44.
2538
- 2539 [230] R. M. Castillo-Morales, A. L. Carreño Otero, S. C. Mendez-Sanchez, M. A. Navarro Da Silva, E.
2540 E. Stashenko, J. E. Duque, 'Mitochondrial affectation, DNA damage and AChE inhibition induced
2541 by *Salvia officinalis* essential oil on *Aedes aegypti* larvae', *Comp. Biochem. Physiol. C.* **2019**, *221*,
2542 29-37.
2543
- 2544 [231] S. Zhu, X. C. Liu, Z. L. Liu, X. Xu, 'Chemical composition of *Salvia plebeian* R.Br. essential oil
2545 and its larvicidal activity against *Aedes aegypti* L', *Trop. J. Pharm. Res.* **2015**, *14(5)*, 831-836.
2546
- 2547 [232] M. Lee, J. Park, H. Lee, 'Acaricidal toxicities and synergistic activities of *Salvia lavandulifolia* oil
2548 constituents against synanthropic mites', *Pest Manag. Sci.* **2018**, *74*, 2468–2479.
2549
- 2550 [233] F. Sohrabi, M. A. Kohanmoo, 'Fumigant toxicity of plant essential oils against oligonychus
2551 afrasiaticus (MCG.) (Acari: Tetranychidae) and identification of their chemical composition', *J.*
2552 *Essent. Oil Bear. Pl.* **2017**, *20(3)*, 844-850.
2553
- 2554 [234] K. Polatoglu, Ö. C. Karako, Y. Y. Yücel, S. Gücel, B. Demirci, F. Demirci, K. H. Can Baser,
2555 'Insecticidal activity of *Salvia veneris* Hedge. essential oil against coleopteran stored product
2556 insects and *Spodoptera exigua* (Lepidoptera)', *Ind. Crops Prod.* **2017**, *97*, 93-100.
2557
- 2558 [235] N. C. Cárdenas-Ortega, M. M. González-Chávez, R. Figueroa-Brito, A. Flores-Macías, D. Romo-
2559 Asunción, D. E. Martínez-González, V. Pérez-Moreno, M. A. Ramos-López, 'Composition of the
2560 essential oil of *Salvia ballotiflora* (Lamiaceae) and its insecticidal activity', *Molecules* **2015**, *20*,
2561 8048-8059.
2562
- 2563 [236] S. S. M. Najafabadi, A. Bagheri, M. A. Seyahooei, H. Zamani, A. Goodarzi, 'Effects of Thyme
2564 and Rosemary essential oils on population growth parameters of *Macrosiphum rosae* (Hemiptera:
2565 Aphididae) on cut flower rose', *J. Crop Prot.* **2018**, *7(1)*, 51-63.
2566
- 2567 [237] R. J. Wallace, N. R. McEwan, F. M. McIntosh, B. Teferedegne, C. J. Newbold, 'Natural products
2568 as manipulators of rumen fermentation', *Asian Australas J. Anim. Sci.* **2002**, *10*, 1458-1468.
2569
- 2570 [238] G. Cobellis, A. Petrozzi, C. Forte, G. Acuti, M. Orrù, M. C. Marcotullio, A. Aquino, A. Nicolini,
2571 V. Mazza, M. Trabalza-Marinucci, 'Evaluation of the effects of mitigation on methane and

- 2572 ammonia production by using *Origanum vulgare* L. and *Rosmarinus officinalis* L. essential oils
 2573 on *in vitro* rumen fermentation systems', *Sustainability* **2015**, 7, 12856-12869.
 2574
- 2575 [239] G. Cobellis, M. Trabalza-Marinucci, Z. Yu, 'Critical evaluation of essential oils as rumen
 2576 modifiers in ruminant nutrition: A review', *Sci. Total Environ.* **2016**, 545-546, 556-568.
 2577
- 2578 [240] G. Cobellis, M. Trabalza-Marinucci, M. C. Marcotullio, Z. Yu, 'Evaluation of different essential
 2579 oils in modulating methane and ammonia production, rumen fermentation, and rumen bacteria *in*
 2580 *vitro*', *Anim. Feed Sci. Tech.* **2016**, 2015, 25-36.
 2581
- 2582 [241] M. Gunal, B. Pinsky, A. A. Abughazale, 'Evaluating the effects of essential oils on methane
 2583 production and fermentation under *in vitro* conditions', *It. J. Anim. Sci.* **2017**, 16(3), 500-506.
 2584
- 2585 [242] M. Adaszyńska-Skwirzyńska, D. Szczerbińska, 'The effect of lavender (*Lavandula angustifolia*)
 2586 essential oil as a drinking water supplement on the production performance, blood biochemical
 2587 parameters, and ileal microflora in broiler chickens', *Poultry Sci.* **2019**, 98, 358-365.
 2588
- 2589 [243] I. Giannenas, A. Tzora, I. Sarakatsianos, A. Karamoutsios, S. Skoufos, N. Papaioannou, I.
 2590 Anastasiou, I. Skoufos, 'The effectiveness of the use of oregano and laurel essential oils in chicken
 2591 feeding', *Ann. Anim. Sci.* **2016**, 16(3), 779-796.
 2592
- 2593 [244] J. Lejonklev, U. Kidmose, S. Jensen, M. A. Petersen, A. L. F. Helwing, G. Mortensen, M. R.
 2594 Weisbjerg, M. K. Larsen, 'Short communication: Effect of oregano and caraway essential oils on
 2595 the production and flavor of cow milk', *J. Dairy Sci.* **2016**, 99(10), 7898-7903.
 2596
- 2597 [245] E. Tekce, M. Gul., 'Effects of *Origanum Syriacum* essential oil on blood parameters of broilers
 2598 reared at high ambient heat', *Braz. J. Poultry Sci.* **2017**, 19(4), 655-662.
 2599
- 2600 [246] Y. Zou, Q. Xiang, J. Wang, J. Peng, H. Wei, Oregano essential oil improves intestinal morphology
 2601 and expression of tight junction proteins associated with modulation of selected intestinal Bacteria
 2602 and immune status in a pig model, **2016**, Article ID 5436738.
 2603
- 2604 [247] J. Arruda da Cunha, C. de Avila Scheeren, V. Pedroso Fausto, L. Daiane Willrich de Melo, B.
 2605 Henneman, C. Piccinin Frizzo, R. de Almeida Vaucher, A. Castagna de Vargas, B. Baldisserotto,
 2606 'The antibacterial and physiological effects of pure and nanoencapsulated *Origanum majorana*
 2607 essential oil on fish infected with *Aeromonas hydrophila*', *Microb. Pathog.* **2018**, 124, 116-121.
 2608
- 2609 [248] H. Gholipourkanani, N. Buller, A. Lymbery, 'In vitro antibacterial activity of four nano-
 2610 encapsulated herbal essential oils against three bacterial fish pathogens', *Aquac. Res.* **2019**, 50,
 2611 871-875.
 2612
- 2613 [249] M. A. E. Mabrok, A. Wahdan, 'The immune modulatory effect of oregano (*Origanum vulgare* L.)
 2614 essential oil on *Tilapia zillii* following intraperitoneal infection with *Vibrio anguillarum*',
 2615 *Aquacult. Int.* **2018**, 26, 1147-1160.
 2616
- 2617 [250] S. Mizuno, S. Urawa, M. Miyamoto, M. Hatakeyama, Y. Sasaki, N. Koide, S. Tada, H. Ueda,
 2618 'Effects of dietary supplementation with oregano essential oil on prevention of the ectoparasitic
 2619 protozoans *Ichthyobodo salmonis* and *Trichodina truttae* in juvenile chum salmon *Oncorhynchus*
 2620 *keta*', *J. Fish Biol.* **2018**, 93, 528-539.
 2621
- 2622 [251] M. Kacániová, M. Terentjeva, N. Vukovic, C. Puchalski, S. Roychoudhury, S. Kunová, A. Kluga,
 2623 M. Tokár, M. Kluz, E. Ivanišová, 'The antioxidant and antimicrobial activity of essential oils
 2624 against *Pseudomonas* spp. isolated from fish', *Saudi Pharm. J.* **2017**, 25, 1108-1116.
 2625
- 2626 [252] S. Amat, D. Baines, T. W. Alexander, 'A vapour phase assay for evaluating the antimicrobial
 2627 activities of essential oils against bovine respiratory bacterial pathogens', *Lett. Appl. Microbiol.*
 2628 **2017**, 65, 489-495.
 2629

2630 [253] F. Rejes-Jurado, T. Cervantes-Rincon, H. Bach, A. Lopez-Malo, E. Palou, 'Antimicrobial activity
2631 of Mexican oregano (*Lippia berlandieri*), thyme (*Thymus vulgaris*), and mustard (*Brassica nigra*)
2632 essential oils in gaseous phase', *Ind. Crops Prod.* **2019**, *131*, 90-95.
2633

2634 [254] F. C. de Aguiar, A. L. Solarte, C. Tarradas, I. Luque, A. Maldonado, A. Galan-Relano, B. Huerta,
2635 'Antimicrobial activity of selected essential oils against *Streptococcus suis* isolated from pigs',
2636 *Microbiology Open.* **2018**, *7*, e613.
2637

2638 [255] F. C. de Aguiar, A. L. Solarte, C. Tarradas, L. Gomez-Gascon, R. Astorga, A. Maldonado, B.
2639 Huerta, 'Combined effect of conventional antimicrobials with essential oils and their main
2640 components against resistant *Streptococcus suis* strains', *Lett. Appl. Microbiol.* **2019**, *68*, 562-572.
2641

2642 [256] K. Dhakal, F. Tiezzi, J. S. Clay, C. Maltecca, 'Causal relationships between clinical mastitis
2643 events, milk yields and lactation persistency in US Holsteins', *Livest. Sci.* **2016**, *189*, 8-16.
2644

2645 [257] S. Ksouri, S. Djebir, A. A. Bentorki, A. Gouri, Y. Hadeif, A. Benakhla, 'Antifungal activity of
2646 essential oils extract from *Origanum floribundum* Munby, *Rosmarinus officinalis* L. and *Thymus*
2647 *ciliatus* Desf. against *Candida albicans* isolated from bovine clinical mastitis', *J. Mycol. Med.*
2648 **2017**, *27*, 245-249.
2649

2650 [258] B. Grzesiak, B. Kolodziej, A. Głowacka, H. Krukowski, 'The effect of some natural essential oils
2651 against bovine mastitis caused by *Prototheca zopfii* isolates in vitro', *Mycopathologia* **2018**, *183*,
2652 541-550.
2653

2654 [259] M. C. Queiroga, M. Pinto Coelho, S. Macedo Arantes, M. E. Potes, M. R. Martins, 'Antimicrobial
2655 activity of essential oils of lamiaceae aromatic spices towards sheep mastitis-causing
2656 *Staphylococcus aureus* and *Staphylococcus epidermidis*', *J. Essent. Oil Bear. Pl.* **2018**, *21(5)*,
2657 1155-1165.
2658

2659 [260] M. O. Alekish, Z. B. Ismail, M. S. Awawdeh, S. Shatnawi, 'Effects of intramammary infusion of
2660 sage (*Salvia officinalis*) essential oil on milk somatic cell count, milk composition parameters and
2661 selected hematology and serum biochemical parameters in Awassi sheep with subclinical mastitis',
2662 *Vet. World* **2017**, *10*, 895-900.
2663

2664 [261] H. Janacuda-Vidales, E. Pena-Gonzalez, A. D. Alarcon-Rojo, J. Ortega-Gutierrez, N. Aguilar-
2665 Palma, 'Determination of carcass yield, sensory and acceptance of meat from male and female
2666 pigs with dietary supplementation of oregano essential oils', *Ital. J. Anim. Sci.* **2019**, *18(1)*, 668-
2667 678.
2668

2669 [262] P. D. Katsoulos, M. A. Karatzia, C. I. Dovas, G. Filioussis, E. Papadopoulos, E. Kiossis, K.
2670 Arsenopoulos, T. Papadopoulos, C. Boscas, H. Karatzias, 'Evaluation of the in-field efficacy of
2671 oregano essential oil administration on the control of neonatal diarrhea syndrome in calves', *Res.*
2672 *Vet. Sci.* **2017**, *115*, 478-483.
2673

2674 [263] A. Unal, N. Kocabagli, 'Effect of different dosages of oregano oil on performance and some blood
2675 parameters in lambs', *Ankara Üniv. Vet. Fak. Derg.* **2014**, *61*, 199-204.
2676

2677 [264] P. Dudko, A. Junkuszew, W. Bojar, M. Milerski, K. Szczepaniak, J. Le Scouarnec, J. Schmidova,
2678 K. Tomczuk, M. Grzybek, 'Effect of dietary supplementation with preparation comprising the
2679 blend of essential oil from *Origanum vulgare* (lamiaceae) and *Citrus* spp. (citraceae) on coccidia
2680 invasion and lamb growth', *Ital. J. Anim. Sci.* **2018**, *17(1)*, 57-65.
2681

2682 [265] C. Cheng, Z. Liu, Y. Zhou, H. Wei, X. Zhang, M. Xia, Z. Deng, Y. Zou, S. Jiang, J. Peng, 'Effect
2683 of oregano essential oil supplementation to a reduced-protein, amino acid-supplemented diet on
2684 meat quality, fatty acid composition, and oxidative stability of *Longissimus thoracis* muscle in
2685 growing-finishing pigs', *Meat Sci.* **2017**, *133*, 103-109.
2686

- 2687 [266] W. N. El-Hawarry, R. A. Mohamed, S. A. Ibrahim, ‘Collaborating effects of rearing density and
2688 oregano oil supplementation on growth, behavioral and stress response of Nile tilapia
2689 (*Oreochromis niloticus*)’, *Egypt. J. Aquat. Res.* **2018**, *44*, 173-178.
2690
- 2691 [267] N. Paraskevakis, ‘Effects of dietary Greek oregano (*Origanum vulgare* ssp. *hirtum*)
2692 supplementation on rumen fermentation, enzyme profile and microbial communities in goats’, *J.*
2693 *Anim. Physiol. Anim. Nutr.* **2018**, *102*, 701-705.
2694
- 2695 [268] O. Diler, O. Gormez, I. Diler, S. Metin, ‘Effect of oregano (*Origanum onites* L.) essential oil on
2696 growth, lysozyme and antioxidant activity and resistance against *Lactococcus garvieae* in rainbow
2697 trout, *Oncorhynchus mykiss* (Walbaum)’, *Aquac. Nutr.* **2017**, *23*, 844-851.
2698
- 2699 [269] S. Smeti, H. Hajji, K. Bouzid, J. Abdelmoula, F. Munoz, M. Mahouachi, N. Atti, ‘Effects of
2700 *Rosmarinus officinalis* L. as essential oils or in form of leaves supplementation on goat’s
2701 production and metabolic statute’, *Trop. Anim. Health Prod.* **2015**, *47*, 451-457.
2702
- 2703 [270] M. Torki, S. Sedgh-Gooya, H. Mohammadi, ‘Effects of adding essential oils of rosemary, dill and
2704 chicory extract to diets on performance, egg quality and some blood parameters of laying hens
2705 subjected to heat stress’, *J. App. Anim. Res.* **2018**, *46(1)*, 1118-1126.
2706
- 2707 [271] G. Turk, A. O. Ceribasi, U.G. Simsek, S. Ceribasi, M. Guvenc, S. O. Kaya, M. Ciftci, M. Sonmez,
2708 A. Yuce, A. Bayrakdar, M. Yaman, F. Tonbak, ‘Dietary rosemary oil alleviates heat stress-induced
2709 structural and functional damage through lipid peroxidation in the testes of growing Japanese
2710 quail’, *Anim. Rep. Sci.* **2016**, *164*, 133-143.
2711
- 2712 [272] S. Smeti, M. Joy, H. Hajji, J. L. Alabart, F. Munoz, M. Mahouachi, N. Atti, ‘Effects of *Rosmarinus*
2713 *officinalis* L. essential oils supplementation on digestion, colostrum production of dairy ewes and
2714 lamb mortality and growth’, *Anim. Sci. J.* **2015**, *86*, 679-688.
2715
- 2716 [273] S. Smeti, H. Hajji, I. Mekki, M. Mahouachi, N. Atti, ‘Effects of dose and administration form of
2717 rosemary essential oils on meat quality and fatty acid profile of lamb’, *Small Rum. Res.* **2018**, *158*,
2718 62-68.
2719
- 2720 [274] G. Cobellis, G. Acuti, C. Forte, L. Menghini, S. De Vincenzi, M. Orrù, A. Valiani, D. Pacetti, M.
2721 Tralbalza-Marinucci, ‘Use of *Rosmarinus officinalis* in sheep diet formulations: Effects on ruminal
2722 fermentation, microbial numbers and in situ degradability’, *Small Rum. Res.* **2015**, *126*, 10-18.
2723
- 2724 [275] D. Witkowska, J. Sowińska, D. Murawska, P. Matusievičius, A. Kwiatkowska-Stenzel, T.
2725 Mituniewicz, A. Wójcik, ‘Effect of peppermint and thyme essential oil mist on performance and
2726 physiological parameters in broiler chickens’, *S. Afr. J. Anim. Sci.* **2019**, *49(1)*, 29-39.
2727
- 2728 [276] N. Dehghani, M. Afsharmanesh, M. Salarinoi, H. Ebrahimnejad, A. Bitaraf, ‘Effect of
2729 pennyroyal, savory and thyme essential oils on Japanese quail physiology’, *Heliyon* **2018**, *4*,
2730 e00881.
2731
- 2732 [277] G. Moraes Ramos Valladão, S. Umeda Gallani, S. Kotzent, I. Mateus Assane, F. Pilarski, ‘Effects
2733 of dietary thyme essential oil on hemato-immunological indices, intestinal morphology, and
2734 microbiota of Nile tilapia’, *Aquacult. Int.* **2019**, *27*, 399-411.
2735
- 2736 [278] A. A. A. Abdel-Wareth, E. M. M. Taha, K. H. Südekum, J. Lohakare, ‘Thyme oil inclusion levels
2737 in a rabbit ration: Evaluation of productive performance, carcass criteria and meat quality under
2738 hot environmental conditions’, *Anim. Nutr.* **2018**, *4*, 410-416.
2739
- 2740 [279] G. Magi, E. Marini, B. Facinelli, ‘Antimicrobial activity of essential oils and carvacrol, and
2741 synergy of carvacrol and erythromycin, against clinical, erythromycin-resistant group-A
2742 *Streptococci*’, *Front. Microbiol.* **2015**, *6*, 165.
2743

- 2744 [280] L. Ait Said, K. Zahlane, I. Ghalbane, S. El Messoussi, A. Romane, C. Cavaleiro, L. Salgueiro,
2745 'Chemical composition and antibacterial activity of *Lavandula coronopifolia* essential oil against
2746 antibiotic-resistant bacteria', *Nat. Prod. Res.* **2015**, *29(6)*, 582-585.
2747
- 2748 [281] B. Kot, K. Wierzchowska, A. Gruzewska, D. Lohinau, 'The effects of selected phytochemicals on
2749 biofilm formed by five methicillin-resistant *Staphylococcus aureus*', *Nat. Prod. Res.* **2018**, *32(11)*,
2750 1299-1302.
2751
- 2752 [282] A. Khadir, M. Bendahou, F. Benbelaid, M. A. Abdoune, C. Bellahcene, F. Zenati, A. Muselli, J.
2753 Paolini, J. Costa, 'Chemical Composition and Anti-MRSA Activity of Essential Oil and Ethanol
2754 Extract of *Lavandula multifida* L. from Algeria', *J. Essent. Oil Bear. Pl.* **2016**, *19(3)*, 712-718.
2755
- 2756 [283] M. T. Milenković, D. D. Božić, V. N. Slavkovska, B. S. Lakušić, 'Synergistic effects of *Salvia*
2757 *officinalis* L. essential oils and antibiotics against methicillin-resistant *Staphylococcus aureus*',
2758 *Arch. Biol. Sci., Belgrade*, **2015**, *67(3)*, 949-956.
2759
- 2760 [284] P. S. X. Yap, T. Krishnan, B. C. Yiap, C. P. Hu, K. -G. Chan, S. H. E. Lim, 'Membrane disruption
2761 and anti-quorum sensing effects of synergistic interaction between *Lavandula angustifolia*
2762 (lavender oil) in combination with antibiotic against plasmid-conferred multi-drug-resistant
2763 *Escherichia coli*', *J. App. Microb.* **2014**, *116*, 1119-1128.
2764
- 2765 [285] A. Alexopoulos, S. Plessas, A. Kimbaris, M. Varvatou, I. Mantzourani, M. Fournomiti, V. Tzouti,
2766 A. Nerantzaki, E. Bezirtzoglou, 'Mode of antimicrobial action of *Origanum vulgare* essential oil
2767 against clinical pathogens', *Curr. Res. Nutr. Food Sci.* **2017**, *5(2)*, 109-115.
2768
- 2769 [286] E. Perrin, V. Maggini, I. Maida, E. Gallo, K. Lombardo, M.P. Madarena, S. Buroni, V. C.
2770 Scoffone, F. Firenzuoli, A. Mengoni, R. Fani, 'Antimicrobial activity of six essential oils against
2771 *Burkholderia cepacia* complex: insights into mechanism(s) of action', *Future Microbiol.* **2017**, *13*,
2772 59-67.
2773
- 2774 [287] O. Ghafari, A. Sharifi, A. Ahmadi, B. Nayeri Fasaee, 'Antibacterial and anti-PmrA activity of plant
2775 essential oils against fluoroquinolone-resistant *Streptococcus pneumoniae* clinical isolates', *Lett.*
2776 *Appl. Microbiol.* **2018**, *67*, 564-569.
2777
- 2778 [288] H. Cui, X. Zhang, H. Zhou, C. Zhao, L. Lin, 'Antimicrobial activity and mechanisms of *Salvia*
2779 *sclarea* essential oil', *Bot. Stud.* **2015**, *56*, 16.
2780
- 2781 [289] D. Schillaci, E. M. Napoli, M. G. Cusimano, M. Vitale, G. Ruberto, '*Origanum vulgare* subsp.
2782 *hirtum* essential oil prevented biofilm formation and showed antibacterial activity against
2783 planktonic and sessile bacterial cells', *J. Food Prot.* **2013**, *76(10)*, 1747-1752.
2784
- 2785 [290] J. Bezerra dos Santos Rodrigues, N. Targino de Souza, J. O. Alcântara Scarano, J. M. de Sousa,
2786 M. Cariry Lira, R. C. Bressan Queiroz de Figueiredo, E. Leite de Souza, M. Magnani, 'Efficacy
2787 of using oregano essential oil and carvacrol to remove young and mature *Staphylococcus aureus*
2788 biofilms on food-contact surfaces of stainless steel', *LWT Food Sci. Tech.* **2018**, *93*, 293-299.
2789
- 2790 [291] N. M. Wijesundara, H. P. Vasantha Rupasinghe, 'Essential oils from *Origanum vulgare* and *Salvia*
2791 *officinalis* exhibit antibacterial and anti-biofilm activities against *Streptococcus pyogenes*',
2792 *Microb. Path.* **2018**, *117*, 118-127.
2793
- 2794 [292] J. Kang, L. Liu, X. Wu, Y. Sun, Z. Liu, 'Effect of thyme essential oil against *Bacillus cereus*
2795 planktonic growth and biofilm formation', *Appl. Microbiol. Biotechnol.* **2018**, *102*, 10209-10218.
2796
- 2797 [293] S. H. Mohamed, M. S. M. Mohamed, M. S. Khalil, M. Azmy, M. I. Mabrouk, 'Combination of
2798 essential oil and ciprofloxacin to inhibit/eradicate biofilms in multidrug-resistant *Klebsiella*
2799 *pneumoniae*', *J. Appl. Microb.* **2018**, *125*, 84-95.
2800

- 2801 [294] A. Vidács, E. Kerekes, R. Rajkó, T. Petkovits, N.S. Alharbi, J. M. Khaled, C. Vágvölgyi, J. Krisch,
2802 'Optimization of essential oil-based natural disinfectants against *Listeria monocytogenes* and
2803 *Escherichia coli* biofilms formed on polypropylene surfaces', *J. Mol. Liq.* **2018**, *255*, 257-262.
2804
- 2805 [295] K. Habbadi, T. Meyer, L. Vial, V. Gaillard, R. Benkirane, A. Benbouazza, I. Kerzaon, E. Achbani,
2806 C. Lavire, 'Essential oils of *Origanum compactum* and *Thymus vulgaris* exert a protective effect
2807 against the phytopathogen *Allorhizobium vitis*', *Environ. Sci. Pollut. R.* **2018**, *25*, 29943–29952.
2808
- 2809 [296] A. Gormez, S. Bozari, D. Yanmis, M. Gulluce, G. Agar, F. Sahin, 'The use of essential oils of
2810 *Origanum rotundifolium* as antimicrobial agent against plant pathogenic bacteria', *J. Essent. Oil*
2811 *Bear. Pl.* **2016**, *19(3)*, 656-663.
2812
- 2813 [297] M. E. Carezzano, J. P. Sotelo, E. Primo, E. B. Reinoso, M. F. Paletti Rovey, M. S. Demo, W. F.
2814 Giordano, M. de las M. Oliva, 'Inhibitory effect of *Thymus vulgaris* and *Origanum vulgare*
2815 essential oils on virulence factors of phytopathogenic *Pseudomonas syringae* strains', *Plant biol.*
2816 **2017**, *19*, 599-607.
2817
- 2818 [298] G. Schött, S. Liesegang, F. Gaunitz, A. Gleß, S. Basche, C. Hannig, K. Speer, 'The chemical
2819 composition of the pharmacologically active Thymus species, its antibacterial activity against
2820 *Streptococcus mutans* and the antiadherent effects of *T. vulgaris* on the bacterial colonization of
2821 the in situ pellicle', *Fitoterapia* **2017**, *121*, 118-128.
2822
- 2823 [299] Ö. I. Karadaghoglu, N. Ulusoy, K. H. Can Baser, A. Hanoglu, I. Sık, 'Antibacterial activities of
2824 herbal toothpastes combined with essential oils against *Streptococcus mutans*', *Pathogens* **2019**,
2825 *8*, 20.
2826
- 2827 [300] B. Kulaksiz, S. Er, N. Üstundag-Okur, G. Saltan-Isçan, 'Investigation of antimicrobial activities
2828 of some herbs containing essential oils and their mouthwash formulations', *Turk. J. Pharm. Sci.*
2829 **2018**, *15(3)*, 370-375.
2830
- 2831 [301] E. Bona, S. Cantamessa, M. Pavan, G. Novello, N. Massa, A. Rocchetti, G. Berta, E. Gamalero,
2832 'Sensitivity of *Candida albicans* to essential oils: are they an alternative to antifungal agents?', *J.*
2833 *Appl. Microb.* **2016**, *121*, 1530-1545.
2834
- 2835 [302] M. H. Minoeianhaghighi, L. Sepehrian, H. Shokri, 'Antifungal effects of *Lavandula binaludensis*
2836 and *Cuminum cyminum* essential oils against *Candida albicans* strains isolated from patients with
2837 recurrent vulvovaginal candidiasis', *J. Mycol. Med.* **2017**, *27*, 65-71.
2838
- 2839 [303] S. Dolatabadi, Z. Salari, M. Mahboubi, 'Antifungal effects of *Ziziphora tenuior*, *Lavandula*
2840 *angustifolia*, *Cuminum cyminum* essential oils against clinical isolates of *Candida albicans* from
2841 women suffering from vulvovaginal candidiasis', *Infectio* **2019**, *23(3)*, 222-226.
2842
- 2843 [304] K. Gucwa, S. Milewski, T. Dymerski, P. Szweda, 'Investigation of the antifungal activity and
2844 mode of action of *Thymus vulgaris*, *Citrus limonum*, *Pelargonium graveolens*, *Cinnamomum*
2845 *cassia*, *Ocimum basilicum*, and *Eugenia caryophyllus* essential oils', *Molecules* **2018**, *23*, 1116.
2846
- 2847 [305] Á. Blaskó, Z. Gazdag, P. Gróf, G. Máté, S. Sárosi, J. Krisch, C. Vágvölgyi, L. Makszin, M. Pesti,
2848 'Effects of clary sage oil and its main components, linalool and linalyl acetate, on the plasma
2849 membrane of *Candida albicans*: an *in vivo* EPR study', *Apoptosis* **2017**, *22*, 175-187.
2850
- 2851 [306] D. Scalas, N. Mandras, J. Roana, R. Tardugno, A. M. Cuffini, V. Ghisetti, S. Benvenuti, V. Tullio,
2852 'Use of *Pinus sylvestris* L. (Pinaceae), *Origanum vulgare* L. (Lamiaceae), and *Thymus vulgaris* L.
2853 (Lamiaceae) essential oils and their main components to enhance itraconazole activity against
2854 azole susceptible/not-susceptible *Cryptococcus neoformans* strains', *BMC Complement. Altern.*
2855 *Med.* **2018**, *18*, 43.
2856

- 2857 [307] P. Kumari, R. Mishra, N. Arora, A. Chatrath, R. Gangwa, P. Roy, R. Prasad, 'Antifungal and anti-
2858 biofilm of essential oil active components against *Cryptococcus neoformans* and *Cryptococcus*
2859 *laurentii*', *Front. Microbiol.* **2017**, *8*, 2161.
2860
- 2861 [308] M. Nikkhah, M. Hashemi, M. B. Habibi Najafi, R. Farhoosh, 'Synergistic effects of some essential
2862 oils against fungal spoilage on pear fruit', *Int. J. Food Microbiol.* **2017**, *257*, 285-294.
2863
- 2864 [309] A. Sarkhosh, B. Schaffer, A.I. Vargas, A.J. Palmateer, P. Lopez, A. Soleymani, 'In Vitro
2865 evaluation of eight plant essential oils for controlling Colletotrichum, Botryosphaeria, Fusarium
2866 and Phytophthora fruit rots of avocado, mango and papaya', *Plant Protect. Sci.* **2018**, *54(3)*, 153-
2867 162.
2868
- 2869 [310] F. Dianeze, M. Santos, C. Parra, M. J. Navarro, R. Blanco, F. J. Gea, 'Screening of antifungal
2870 activity of 12 essential oils against eight pathogenic fungi of vegetables and mushroom', *Lett.*
2871 *Appl. Microbiol.* **2018**, *67*, 400-410.
2872
- 2873 [311] M. P. Santamarina, M. D. Ibáñez, M. Marqués, J. Roselló, S. Giménez, M. A. Blázquez,
2874 'Bioactivity of essential oils in phytopathogenic and postharvest fungi control', *Nat. Prod. Res.*
2875 **2017**, *31(22)*, 2675-2679.
2876
- 2877 [312] M. Bill, L. Korsten, F. Remize, M. Glowacz, D. Sivakumar, 'Effect of thyme oil vapours exposure
2878 on phenylalanine ammonia-lyase (PAL) and lipoxygenase (LOX) genes expression, and control of
2879 anthracnose in 'Hass' and 'Ryan' avocado fruit', *Sci. Hort.* **2017**, *224*, 232-237.
2880
- 2881 [313] D. Tančinová, J. Medo, Z. Mašková, D. Foltinová, J. Árvay, 'Effect of essential oils of Lamiaceae
2882 plants on the *Penicillium commune*', *J. Microbiol. Biotech. Food Sci.* **2019**, *8(4)*, 1111-1117.
2883
- 2884 [314] T. Krzysko-Łupicka, W. Walkowiak, M. Białon, 'Comparison of the fungistatic activity of selected
2885 essential oils relative to *Fusarium graminearum* isolates', *Molecules* **2019**, *24*, 311.
2886
- 2887 [315] A. Salamone, G. Scarito, G. Camerata Scovazzo, G. Fascella, 'Control of powdery mildew in cut
2888 roses using natural products in the greenhouse', *Floricult. Ornamental Biotech.* **2009**, *3(1)*, 121-
2889 125.
2890
- 2891 [316] T. A. Misharina, M. B. Terenina, N. I. Krikunova, M. G. Semenova, 'Inhibition of autoxidation of
2892 polyunsaturated fatty acids by clove and oregano essential oils', *Appl. Biochem. Microbiol.* **2019**,
2893 *55(1)*, 67-72.
2894
- 2895 [317] D. D. Jayasena, C. Jo, 'Potential application of essential oils as natural antioxidants in meat and
2896 meat products: A review', *Food Rev. Int.* **2014**, *30*, 71-90.
2897
- 2898 [318] S. Patel, 'Plant essential oils and allied volatile fractions as multifunctional additives in meat and
2899 fish-based food products: a review', *Food Addit. Contam. Part A* **2015**, *32(7)*, 1049-1064.
2900
- 2901 [319] M. Al-Hijazeen, E. J. Lee, A. Mendonca, D. U. Ahn, 'Effect of oregano essential oil (*Origanum*
2902 *vulgare* subsp. *hirtum*) on the storage stability and quality parameters of ground chicken breast
2903 meat', *Antioxidants*, **2016**, *5*, 18, antiox5020018.
2904
- 2905 [320] M. Al-Hijazeen, 'Effect of direct adding oregano essential oil (*Origanum syriacum* L.) on quality
2906 and stability of chicken meat patties', *Food Sci. Technol.* **2018**, *38(1)*, 123-130.
2907
- 2908 [321] K. Ünal, A. Samet Babaoglu, M. Karakaya, 'Effect of oregano, sage and rosemary essential oils
2909 on lipid oxidation and color properties of minced beef during refrigerated storage', *J. Essent. Oil*
2910 *Bear. Pl.* **2014**, *17(5)*, 797-805.
2911
- 2912 [322] M. Křížek, E. Dadáková, F. Vácha, T. Pelikánová, K. Matějková, 'The effects of two essential oil
2913 and UV-light irradiation treatments on the formation of biogenic amines in vacuum packed fillets
2914 of carp (*Cyprinus carpio*)', *LWT-Food Sci. Tech.* **2018**, *95*, 268-273.

- 2915 [323] N. M. Costa Menezes, W. Figueiredo Martins, D. A. Longhi, G. M. Falcão de Aragão, 'Modeling
2916 the effect of oregano essential oil on shelf-life extension of vacuum-packed cooked sliced ham',
2917 *Meat Sci.* **2018**, *139*, 113-119.
2918
- 2919 [324] Z. Stojanovic-Radic, M. Pejcic, N. Jokovic, M. Jokanovic, M. Ivic, B. Sojic, S. Skaljic, P.
2920 Stojanovic, T. Mihajilov-Krstev, 'Inhibition of *Salmonella Enteritidis* growth and storage stability
2921 in chicken meat treated with basil and rosemary essential oils alone or in combination', *Food*
2922 *Contr.* **2018**, *90*, 332-343.
2923
- 2924 [325] R. Iseppi, C. Sabia, S. de Niederhäusern, F. Pellati, S. Benvenuti, R. Tardugno, M. Bondi, P.
2925 Messi, 'Antibacterial activity of *Rosmarinus officinalis* L. and *Thymus vulgaris* L. essential oils
2926 and their combination against foodborne pathogens and spoilage bacteria in ready-to-eat
2927 vegetables', *Nat. Prod. Res.* **2019**, *33*(24), 3568-3572.
2928
- 2929 [326] M. H. Mohammad Abdeldaiem, H. G. Mohammad Ali, M. F. Ramadan, 'Impact of different
2930 essential oils on the characteristics of refrigerated carp (*Cyprinus carpio*) fish fingers', *Food meas.*
2931 **2017**, *11*, 1412-1420.
2932
- 2933 [327] A. C. Pelaez Vital, A. Guerrero, M. Garcia Ornaghi, E. M. Barbosa Carvalho Kempinski, C. Sary,
2934 J. de Oliveira Monteschio, P. T. Matumoto-Pintro, R. Pereira Ribeiro, I. Nunes do Prado, 'Quality
2935 and sensory acceptability of fish fillet (*Oreochromis niloticus*) with alginate-based coating
2936 containing essential oils', *J. Food Sci. Technol.* **2018**, *55*(12), 4945-4955.
2937
- 2938 [328] Sarengaowa, W. Hu, A. Jiang, Z. Xiu, K. Fenga, 'Effect of thyme oil-alginate-based coating on
2939 quality and microbial safety of fresh-cut apples', *J. Sci. Food Agric.* **2018**, *98*, 2302-2311.
2940
- 2941 [329] M. H. Abdeldaiem, H. G. Mohammad, M. F. Ramadan, 'Improving the quality of silver carp fish
2942 fillets by gamma irradiation and coatings containing rosemary oil', *J. Aquat. Food Prod. Technol.*
2943 **2018**, *27*(5), 568-579.
2944
- 2945 [330] M. Llana-Ruiz-Cabello, S. Pichardo, J. María Bermudez, A. Baños, J. J. Ariza, E. Guillamón, S.
2946 Aucejo, A. M. Cameán, 'Characterisation and antimicrobial activity of active polypropylene films
2947 containing oregano essential oil and Allium extract to be used in packaging for meat products',
2948 *Food Addit. Contam. Part A* **2018**, *35*(4), 783-792.
2949
- 2950 [331] Z. Dong, F. Xu, I. Ahmed, Z. Li, H. Lin, 'Characterization and preservation performance of active
2951 polyethylene films containing rosemary and cinnamon essential oils for Pacific white shrimp
2952 packaging', *Food Contr.* **2018**, *92*, 37-46.
2953
- 2954 [332] S. Kwon, Y. Chang, J. Han, 'Oregano essential oil-based natural antimicrobial packaging film to
2955 inactivate *Salmonella enterica* and yeasts/molds in the atmosphere surrounding cherry tomatoes',
2956 *Food Microbiol.* **2017**, *65*, 114-121.
2957
- 2958 [333] S. Ketkaew, P. Kasemsiri, S. Hiziroglu, W. Mongkolthananuk, R. Wannasutta, U. Pongsa, P.
2959 Chindaprasirt, 'Effect of oregano essential oil content on properties of green biocomposites based
2960 on cassava starch and sugarcane bagasse for bioactive packaging', *J. Polym. Environ.* **2018**, *26*,
2961 311-318.
2962
- 2963 [334] L. Lahnine, S. Mghazli, M. Mahrouz, N. Hidar, M. Ouhammou, M. Mouhib, S. Zantar, H. Hanine,
2964 H. Osson, M.A. Misdaq, 'Decontamination by gamma irradiation at lowdoses of *Thymus*
2965 *satureioides* and its impact on physico-chemical quality', *Food Bioprod. Process* **2017**, *104*, 48-
2966 56.
2967
- 2968 [335] C. Kirkin, G. Gunes, 'Modified atmosphere packaging and gamma-irradiation of some herbs and
2969 spices: Effects on antioxidant and antimicrobial properties', *J. Food Process Preserv.* **2018**, *42*,
2970 e13678.
2971
2972

- 2973 [336] S. A. Mahgoub, R. M. El-Mekawi, M. E. Abd El-Hack, W. R. El-Ghareeb, G. M. Suliman, A. N.
2974 Alowaimer, A. A. Swelum, 'Inactivation of *Listeria monocytogenes* in ready-to-eat smoked
2975 turkey meat by combination with packaging atmosphere, oregano essential oil and cold
2976 temperature', *AMB Expr.* **2019**, *9*, 54.
2977
- 2978 [337] M. Boskovic, J. Djordjevic, J. Ivanovic, J. Janjic, N. Zdravkovic, M. Glisic, N. Glamoclija, B.
2979 Baltic, V. Djordjevic, M. Baltic, 'Inhibition of *Salmonella* by thyme essential oil and its effect on
2980 microbiological and sensory properties of minced pork meat packaged under vacuum and modified
2981 atmosphere', *Int. J. Food Microbiol.* **2017**, *258*, 58-67.
2982
- 2983 [338] A. Russo, V. Cardile, A. C. E. Graziano, C. Formisano, D. Rigano, M. Canzoneri, M. Bruno, F.
2984 Senatore, 'Comparison of essential oil components and in vitro anticancer activity in wild and
2985 cultivated *Salvia verbenaca*', *Nat. Prod. Res.* **2015**, *29(17)*, 1630-1640.
2986
- 2987 [339] Y. Zhang, R. Chen, Y. Wang, C. Quing, W. Wang, Y. Yang, 'In vitro and in vivo efficacy studies
2988 of *Lavender angustifolia* essential oil and its active constituents on the proliferation of human
2989 prostate cancer', *Integr. Cancer Ther.* **2017**, *16(2)*, 215-226.
2990
- 2991 [340] B. Justus, C. C. Kanunfre, J. M. Budel, M. Ferreira de Faria, V. Raman, J. Padilha de Paula, P. V.
2992 Farago, 'New insights into the mechanisms of French lavender essential oil on non small- cell lung
2993 cancer cell growth', *Ind. Crops Prod.* **2019**, *136*, 28-36.
2994
- 2995 [341] S. Tariq, S. Wani, W. Rasool, K. Shafi, M. A. Bhat, A. Prabhakar, A. H. Shalla, M.A. Rather, 'A
2996 comprehensive review of the antibacterial, antifungal and antiviral potential of essential oils and
2997 their chemical constituents against drug-resistant microbial pathogens', *Micr. Path.* **2019**, *134*,
2998 103580.
2999
- 3000 [342] J. Reichling, P. Schnitzler, U. Suschke, R. Saller, 'Essential oils of aromatic plants with
3001 antibacterial, antifungal, antiviral, and cytotoxic properties – an overview', *Forsch.*
3002 *Komplementmed* **2009**, *16*, 79–90.
3003
- 3004 [343] D. H. Gilling, M. Kitajima, J. R. Torrey, K. R. Bright, 'Antiviral efficacy and mechanisms of
3005 action of oregano essential oil and its primary component carvacrol against murine norovirus', *J.*
3006 *App. Microbiol.* **2014**, *116*, 1149-1163.
3007
- 3008 [344] H. Choi, 'Chemical constituents of essential oils possessing anti-influenza A/WS/33 virus
3009 activity', *Osong Public Health Res. Perspect.* **2018**, *9(6)*, 348–353.
3010
- 3011 [345] G. Feriotto, N. Marchetti, V. Costa, S. Beninati, F. Tagliati, C. Mischiati, 'Chemical composition
3012 of essential oils from *Thymus vulgaris*, *Cymbopogon citratus*, and *Rosmarinus officinalis*, and their
3013 effects on the HIV-1 Tat protein function", *Chem. Biodiversity* **2018**, *15*, e1700436.
3014
- 3015 [346] V. Cagno, B. Sgorbini, C. Sanna, C. Cagliero, M. Ballero, A. Civra, M. Donalisio, C. Bicchi, D.
3016 Lembo, P. Rubiolo, 'In vitro anti-herpes simplex virus-2 activity of *Salvia desoleana* Atzei & V.
3017 Picci essential oil', *PLoS ONE* **2017**, *12(2)*, e0172322.
3018
- 3019 [347] E. M. de Lavor, A. W. Calvancate Fernandes, R. Braga de Andrade Teles, A. E. Barbosa Pereira
3020 Leal, R. Gonçalves de Oliveira Júnior, M. Gama e Silva, A. P. de Oliveira, J. Cabral Silva, M. Tais
3021 de Moura Fontes Araújo, H. D. Melo Coutinho, I. R. Alencar de Menezes, L. Picot, J. R. Guedes
3022 da Silva Almeida, 'Essential oils and their major compounds in the treatment of chronic
3023 inflammation: A review of antioxidant potential in preclinical studies and molecular mechanisms',
3024 *Oxid. Med. Cell. Longev.* **2018**, Article ID 6468593.
3025
- 3026 [348] G. R. Gandhi, A. B. Silva Vasconcelos, G. H. Haran, V. K. da Silva Calisto, G. Jothi, J. de Souza
3027 Siqueira Quintans, L. E. Cuevas, N. Narain, L. J. Quintans Júnior, R. Cipolotti, R. Queiroz Gurgel,
3028 'Essential oils and its bioactive compounds modulating cytokines: A systematic review on anti-
3029 asthmatic and immunomodulatory properties', *Phytomedicine* **2019**, 152854.
3030

- 3031 [349] C. Cheng, Y. Zou, J. Peng, 'Oregano essential oil attenuates RAW264.7 cells from
3032 lipopolysaccharide-induced inflammatory response through regulating NADPH oxidase
3033 activation-driven oxidative stress', *Molecules* **2018**, *23*, 1857.
3034
- 3035 [350] A.Tosun, S. Khan, Y. S. Kim, Á. Calín-Sánchez,X. Hysenaj, Á. A. Carbonell-Barrachina,
3036 'Essential oil composition and anti-inflammatory activity of *Salvia officinalis* L (Lamiaceae) in
3037 murin macrophages', *Trop. J. Pharm. Res.* **2014**, *13(6)*, 937-942.
3038
- 3039 [351] M. Zuzarte, J. M. Alves-Silva, M. Alves, C. Cavaleiro, L. Salgueiro, M. T. Cruz, 'New insights
3040 on the anti-inflammatory potential and safety profile of *Thymus carnosus* and *Thymus*
3041 *camphoratus* essential oils and their main compounds', *J. Ethnopharmacol.* **2018**, *225*, 10-17.
3042
- 3043 [352] J. C. R. Goncalves, D. Andrade de Meneses, A. Pereira de Vasconcelos, C. Alves Piauilino, F. R.
3044 de Castro Almeida, E. M. Napoli, G. Ruberto, D. A. Machado de Araujo, 'Essential oil
3045 composition and antinociceptive activity of *Thymus capitatus*', *Pharm. Biol.* **2017**, *55(1)*, 782-786.
3046
- 3047 [353] L. Caputo, L. F. Souza, S. Alloisio, L. Cornara, V. De Feo, '*Coriandrum sativum* and *Lavandula*
3048 *angustifolia* essential oils: chemical composition and activity on central nervous system', *Int. J.*
3049 *Mol. Sci.* **2016**, *17*, 1999.
3050
- 3051 [354] V. López, B. Nielsen, M. Solas, M. J. Ramírez, A. K. Jäger, 'Exploring pharmacological
3052 mechanisms of Lavender (*Lavandula angustifolia*) essential oil on central nervous system targets',
3053 *Front. Pharmacol.* **2017**, *8*, 280.
3054
- 3055 [355] B. Ali, N. A. Al-Wabel, S. Shams, A. Ahamad, S. A. Khan, F. Anwar, 'Essential oils used in
3056 aromatherapy: A systemic review', *Asian Pac. J. Trop. Biomed.* **2015**, *5(8)*, 601-611.
3057
- 3058 [356] K. Sowndhararajan, S. Kim, 'Influence of fragrances on human psychophysiological activity: with
3059 special reference to human electroencephalographic response', *Sci. Pharm.* **2016**, *84*, 724-751.
3060
- 3061 [357] C. Ayik, D. Özden, 'The effects of preoperative aromatherapy massage on anxiety and sleep
3062 quality of colorectal surgery patients: A randomized controlled study', *Complement. Ther. Med.*
3063 **2018**, *36*, 93-99.
3064
- 3065 [358] A. Nasiri, M. A. Mahmodi, 'Aromatherapy massage with lavender essential oil and the prevention
3066 of disability in ADL in patients with osteoarthritis of the knee: A randomized controlled clinical
3067 trial', *Complement. Ther. Clin.* **2018**, *30*, 116-121.
3068
- 3069 [359] M. Hassanzadeh, F. Kiani, S. Bouya, M. Zarei, 'Comparing the effects of relaxation technique and
3070 inhalation aromatherapy on fatigue in patients undergoing hemodialysis', *Complement. Ther. Clin.*
3071 **2018**, *31*, 210-214.
3072
- 3073 [360] N. Tugut, G. Demirel, M. Baser, E. E. Ata, S. Karakus, 'Effects of lavender scent on patients'
3074 anxiety and pain levels during gynecological examination', *Complement. Ther. Clin.* **2017**, *28*, 65-
3075 69.
3076
- 3077 [361] M. O. Villareal, A. Ikeya, K. Sasaki, A. Ben Arfa, M. Neffati, H. Isoda, 'Anti-stress and neuronal
3078 cell differentiation induction effects of *Rosmarinus officinalis* L. essential oil', *BMC Compl.*
3079 *Altern. Med.* **2017**, *17*, 549.
3080
- 3081 [362] T. Satou, Y. Hanashima, I. Mizutani, K. Koike, 'The effect of inhalation of essential oil from
3082 *Rosmarinus officinalis* on scopolamine-induced Alzheimer's type dementia model mice', *Flavour*
3083 *Fragr. J.* **2017**, *33*, 230-234.
3084
- 3085 [363] A. Bouyahya, N. Dakka, F. Lagrouh, J. Abrini, Y. Bakri, 'In vitro antiproliferative and
3086 antidermatophyte activities of essential oils from three Moroccan medicinal plants', *JBAPN* **2018**,
3087 *8(3)*, 144-153.
3088

- 3089 [364] M. G. Donadu, D. Usai, V. Mazzarello, P. Molicotti, S. Cannas, M. G. Bellardi, S. Zanetti, 'Change
3090 in Caco-2 cells following treatment with various lavender essential oils', *Nat. Prod. Res.* **2017**,
3091 *31(18)*, 2203-2206.
3092
- 3093 [365] S. Gezici, N. Sekeroglu, A. Kijjoa, 'In vitro anticancer activity and antioxidant properties of
3094 essential oils from *Populus alba* L. and *Rosmarinus officinalis* L. from South Eastern Anatolia of
3095 Turkey', *Indian J. Pharm. Educ.* **2017**, *51(3)*, S498-S503.
3096
- 3097 [366] S. R. Balusamy, H. Perumalsamy, Md. Amdad Huq, B. Balasubramanian, 'Anti-proliferative
3098 activity of *Origanum vulgare* inhibited lipogenesis and induced mitochondrial mediated apoptosis
3099 in human stomach cancer cell lines', *Biomed. Pharmacother.* **2018**, *108*, 1835-1844.
3100
- 3101 [367] K. R. Begnini, F. Nedel, R. G. Lund, P. H. de Azambuja Carvalho, M. R. Alves Rodrigues, F. T.
3102 Alves Beira, F. A. Burkert Del-Pino, 'Composition and antiproliferative effect of essential oil of
3103 *Origanum vulgare* against tumor cell lines', *J. Med. Food* **2014**, *17(10)*, 1-5.
3104
- 3105 [368] H. S. Elshafie, M. F. Armentano, M. Carmosino, S. A. Bufo, V. De Feo, I. Camele, 'Cytotoxic
3106 activity of *Origanum vulgare* L. on hepatocellular carcinoma cell line HepG2 and evaluation of its
3107 biological activity', *Molecules* **2017**, *22*, 1435.
3108
- 3109 [369] F. Öke Altuntaş, I. Demirtaş, 'Real-Time cell analysis of the cytotoxicity of *Origanum acutidens*
3110 essential oil on HT-29 and HeLa cell lines', *Turk. J. Pharm. Sci.* **2017**, *14(1)*, 29-33.
3111
- 3112 [370] G. Privitera, T. Luca, S. Castorina, R. Passanisi, G. Ruberto, E. Napoli, 'Anticancer activity of
3113 *Salvia officinalis* essential oil and its principal constituents against hormone-dependent tumour
3114 cells', *Asian Pac. J. Trop. Biomed.* **2019**, *9(1)*, 24-28.
3115
- 3116 [371] G. Privitera, E. Napoli, T. Luca, G. Ruberto, S. Castorina, 'In vitro anti-proliferative effect of
3117 *Salvia officinalis* essential oil and its three main components on human lung cancer cells', *AJPCT*
3118 **2014**, *2(10)*, 1159-1168.
3119
- 3120 [372] R. Russo, M. T. Corasaniti, G. Bagetta, L. A. Morrone, 'Exploitation of cytotoxicity of some
3121 essential oils for translation in cancer therapy', *Evid.-Based Complementary Altern. Med.* **2015**,
3122 397821.
3123
- 3124 [373] H. Durgba, R. Thirugnanasampandan, G. Ramya, M. G. Ramanth, 'Inhibition of inducible nitric
3125 oxide synthase gene expression (iNOS) and cytotoxic activity of *Salvia sclarea* L. essential oil',
3126 *J. King Saud. Univ. Sci.* **2016**, *28*, 390-395.
3127
- 3128 [374] A. Russo, V. Cardile, A. C. E. Graziano, R. Avola, M. Bruno, D. Rigano, 'Involvement of Bax
3129 and Bcl-2 in induction of apoptosis by essential oils of three lebanese *Salvia* species in human
3130 prostate cancer cells', *Int. J. Mol. Sci.* **2018**, *19*, 292.
3131
- 3132 [375] S. M. El-Darier, A. M. D. El-Ahwany, E. T. Elkenany, A. A. Abdeldaim, 'An in vitro study on
3133 antimicrobial and anticancer potentiality of thyme and clove oils', *Rend. Lincei Sci. Fis. Nat.* **2018**,
3134 *29*, 131-139.
3135
- 3136 [376] M. Stupar, M. Lj. Grbić, A. Džamić, N. Unković, M. Ristić, A. Jelikić, J. Vukojević, 'Antifungal
3137 activity of selected essential oils and biocide benzalkonium chloride against the fungi isolated from
3138 cultural heritage objects', *S. Afr. J. Bot.* **2014**, *93*, 118-124.
3139
- 3140 [377] N. S. Geweely, H. A. Afifi, D. M. Ibrahim, M. M. Soliman, 'Efficacy of essential oils on fungi
3141 isolated from archaeological objects in Saqqara excavation, Egypt', *Geomicrobiol. J.* **2019**, *36(2)*,
3142 148-168.
3143
- 3144 [378] V. Rotolo, M. L. De Caro, A. Giordano, F. Palla, 'Solunto archaeological park in Sicily: life under
3145 mosaic tesserae', *Fl. Medit.* **2018**, *28*, 233-245.
3146

- 3147 [379] L. Levinskaite, A. Paskevicius, “Fungi in water-damaged buildings of Vilnius old vity and their
3148 susceptibility towards disinfectants and essential oils”, *Indoor Built Environ.* **2013**, 22(5), 766–
3149 775.
- 3150 [380] M. Mironescu, C. Georgescu, ‘Activity of some essential oils against common spoilage fungi of
3151 buildings’, *Acta Univ. Cibiniensis, Ser. E: Food Technol.* **2010**, 2, 41-46.
- 3152 [381] M. Stupar, M. L. Grbic, G. Subakov Simic, A. Jelkic, J. Vukojevic, M. Sabovljevic, ‘Sub-aerial
3153 biofilms investigation and new approach in biocide application in cultural heritage conservation:
3154 Holy Virgin Church (Gradac Monastery, Serbia)’, *Indoor Built Environ.* **2014**, 23(4), 584–593.
- 3155 [382] S. Borrego, O. Valdes, I. Vivar, P. Lavin, P. Guiamet, P. Battistoni, S. Gomez de Saravia, P.
3156 Borges, ‘Essential oils of plants as biocides against microorganisms isolated from cuban and
3157 argentine documentary heritage’, *ISRN Microbiology* **2012**, 826786.
- 3158 [383] P. Lavin, S. Gómez de Saravia, P. Guiamet, ‘*Scopulariopsis* sp. and *Fusarium* sp. in the
3159 documentary heritage: evaluation of their biodeterioration ability and antifungal effect of two
3160 essential oils’, *Microb. Ecol.* **2016**, 7(3), 628-633.
- 3161 [384] K. Pietrzak, A. Otlewska, D. Danielewicz, K. Dybka, D. Pangallo, L. Kraková, A. Puskárová, M.
3162 Bucková, V. Scholtz, M. Durovic, B. Surma-Slusarska, K. Demnerová, B. Gutarowska,
3163 ‘Disinfection of archival documents using thyme essential oil, silver nanoparticles misting and low
3164 temperature plasma’, *J. Cult. Herit.* **2017**, 24, 69-77.
- 3165 [385] M. Di Vito, M. G. Bellardi, P. Colaizzi, D. Ruggiero, C. Mazzucca, L. Micheli, S. Sotgiu, S.
3166 Iannuccelli, M. Michelozzi, F. Mondello, P. Mattarelli, M. C. Sclocchi, ‘Hydrolates and gellan: an
3167 eco-innovative synergy for safe cleaning of paper artworks’, *Stud. Conserv.* **2018**, 63(1), 13-23.
- 3168 [386] R. Pathania, H. K. Ravinder Kaushik, M. Azhar Khan, “Essential oil nanoemulsions and their
3169 antimicrobial and food applications”, *Curr. Res. Nutr Food Sci Jour.* **2018**, 6(3), 626-643.
- 3170 [387] G. A. Cardoso-Ugarte, N. Ramírez-Corona, A. López-Malo, E. Palou, M. F. San Martín-González,
3171 M. T. Jiménez-Munguía, “Modeling phase separation and droplet size of W/O emulsions with
3172 oregano essential oil as a function of its formulation and homogenization conditions”, *J. Disper.*
3173 *Sci. Technol.* **2018**, 39(7), 1065-1073.
- 3174 [388] R. Moghimi, A. Aliahmadi, D. J. McClements, H. Rafati, “Investigations of the effectiveness of
3175 nanoemulsions from sage oil as antibacterial agents on some food borne pathogens”, *LWT - Food*
3176 *Sci. Technol.* **2016**, 71, 69-76.
- 3177 [389] S. Gharehaghadeh, N. Karimi, S. Forghani, M. Nourazarian, S. Gharehaghadeh, V. Jabbari, M.
3178 S. Khiabani, H. S. Kafil, “Application of *Salvia multicaulis* essential oil-containing nanoemulsion
3179 against food-borne pathogens”, *Food Biosci.* **2017**, 19, 128-133.
- 3180 [390] V. Ryu, D. J. McClements, M. G. Corradini, L. McLandsborough, “Effect of ripening inhibitor
3181 type on formation, stability, and antimicrobial activity of thyme oil nanoemulsion”, *Food Chem.*
3182 **2018**, 245, 104-111.
- 3183 [391] A. S. Doost, F. Devlieghere, A. Dirckx, P. Van der Meeren, “Fabrication of *Origanum compactum*
3184 essential oil nanoemulsions stabilized using Quillaja saponin biosurfactant”, *J. Food Process.*
3185 *Preserv.* **2018**, 42, e13668.
- 3186 [392] M. J. Martin-Piñero, P. Ramirez, J. Muñoz, M. C. Alfaro, “Development of rosemary essential oil
3187 nanoemulsions using a wheat biomass-derived surfactant”, *Colloid Surface B* **2019**, 173, 486-492.
- 3188 [393] M. Jose Martin, L. A. Trujillo, M. C. Garcia, M. C. Alfaro, J. Muñoz, “Effect of emulsifier HLB
3189 and stabilizer addition on the physical stability of thyme essential oil emulsions”, *J. Disper. Sci.*
3190 *Technol.* **2018**, 39(11), 1627-1634.

- 3205 [394] M. M. A. Rashed, C. Zhang, A. D. S. Ghaleb, J. Li, A. Nagi, H. Majeed, A. M. Bakry, J. Haider,
3206 Z. Xu, Q. Tong, “Techno-functional properties and sustainable application of nanoparticles based
3207 *Lavandula angustifolia* essential oil fabricated using unsaturated lipidcarrier and biodegradable
3208 wall material”, *Ind. Crops Prod.* **2019**, 136, 66-76.
3209
- 3210 [395] K. J. Figueroa-Lopez, A. A. Vicente, M. A. M. Reis, S. Torres-Giner, J. M. Lagaron,
3211 “Antimicrobial and antioxidant performance of various essential oils and natural extracts and their
3212 Incorporation into biowaste derived Poly(3-hydroxybutyrate-co-3-hydroxyvalerate) layers made
3213 from electrospun ultrathin fibers”, *Nanomaterials* **2019**, 9, 144.
3214
- 3215 [396] M. H. Alkhatib, S. M. AlMotwaa, H. M. Alkreathy, “Incorporation of ifosfamide into various
3216 essential oils –based nanoemulsions ameliorates its apoptotic effect in the cancers cells”, *Scientific
3217 Reports* **2019**, 9, 695.
3218
- 3219 [397] R. D. De Oliveira-Filho, A. Roncalli Avles e Silva, R. De Azevedo Moreira, N. Accioly Pinto
3220 Nogueira, “Biological activities and pharmacological applications of cyclodextrins complexed
3221 with essential oils and their volatile components: A systematic review”, *Curr. Pharm. Des.* **2018**,
3222 24, 1-13.
3223
- 3224 [398] C. Yuan, Y. Wang, Y. Liu, Bo Cui, “Physicochemical characterization and antibacterial activity
3225 assessment of lavender essential oil encapsulated in hydroxypropyl-beta-cyclodextrin”, *Food
3226 Chem.* **2019**, 130, 104-110.
3227
- 3228 [399] M. T. Yilmaz, A. Yilmaz, P. Kubra Akman, F. Bozkurt, E. Dertli, A. Basahel, B. Al-Sasi, O.
3229 Taylan, O. Sagdic, “Electrospraying method for fabrication of essential oil loaded-chitosan
3230 nanoparticle delivery systems characterized by molecular, thermal, morphological and antifungal
3231 properties”, *Innov. Food Sci. Emerg. Technol.* **2019**, 52, 166-178.
3232
- 3233 [400] J. Hu, Y. Zhang, Z. Xiao, X. Wang, “Preparation and properties of cinnamon-thyme-ginger
3234 composite essential oil nanocapsules”, *Ind. Crops Prod.* **2018**, 122, 85-92,
3235
- 3236 [401] H. Avci, H. Ghorbanpoor, M. Nurbas, “Preparation of *Origanum minutiflorum* oil-loaded core–
3237 shell structured chitosan nanofibers with tunable properties”, *Polym. Bull.* **2018**, 75, 4129–4144.
3238
- 3239 [402] W. Weisany, S. Samadi, J. Amini, S. Hossaini, S. Yousefi, F. Maggi, “Enhancement of the
3240 antifungal activity of thyme and dill essential oils against *Colletotrichum nymphaeae* by nano-
3241 encapsulation with copper NPs”, *Ind. Crops Prod.* **2019**, 132, 213-225.
3242
- 3243 [403] C. Carbone, C. Martins-Gomes, C. Caddeo, A. M. Silva, T. Musumeci, R. Pignatello, G. Puglisi,
3244 E. B. Souto, “Mediterranean essential oils as precious matrix components and active ingredients
3245 of lipid nanoparticles”, *Int. J. Pharm.* **2018**, 548, 217-226.
3246
- 3247 [404] L. Risaliti, A. Kehagia, E. Daoultzi, D. Lazari, M. C. Bergonzi, S. Vergkizi-Nikolakaki, D.
3248 Hadjipavlou-Litina, A. R. Bilia, “Liposomes loaded with *Salvia triloba* and *Rosmarinus officinalis*
3249 essential oils: *In vitro* assessment of antioxidant, antiinflammatory and antibacterial activities”, *J.
3250 Drug Deliv. Sci. Tec.* **2019**, 51, 493-498.
3251
- 3252 [405] M. Asprea, I. Leto, M. C. Bergonzi, A. R. Bilia, “Thyme essential oil loaded in nanocochleates:
3253 Encapsulation efficiency, *in vitro* release study and antioxidant activity”, *LWT - Food Sci. Tech.*
3254 **2017**, 497e502.
3255
- 3256 [406] A. P. Perez, N. Perez, C. M. Suligoy Lozano, M. J. Altube, M. A. de Farias, R. Villares Portugal,
3257 F. Buzzola, M. J. Morilla, E. Lilia Romero, “The anti MRSA biofilm activity of *Thymus vulgaris*
3258 essential oil in nanovesicles”, *Phytomedicine* **2019**, 57, 339-351.
3259
- 3260 [407] G. Granata, S. Stracquadanio, M. Leonardi, E. Napoli, G. M. L. Consoli, V. Cafiso, S. Stefani, C.
3261 Geraci, “Essential oils encapsulated in polymer-based nanocapsules as potential candidates for
3262 application in food preservation”, *Food Chem.* **2018**, 269, 286-292.

3263 [408] M. Alboofetileh, M. Rezaei, H. Hosseini, M. Abdollahi, “Morphological, physico-mechanical, and
3264 antimicrobial properties of sodium alginate-montmorillonite nanocomposite films incorporated
3265 with marjoram essential oil”, *J. Food Process. Preserv.* **2018**, 42, e13596.
3266

3267 [409] Q. Liu, W. Wang, J. Qi, Q. Huang, J. Xiao, “Oregano essential oil loaded soybean polysaccharide
3268 films: Effect of Pickering type immobilization on physical and antimicrobial properties”, *Food*
3269 *Hydrocol.* **2019**, 87, 165-172.
3270

3271 [410] M. Volić, I. Pajić-Lijaković, V. Djordjević, Z. Knežević-Jugović, I. Pećinar, Z. Stevanović-Dajić,
3272 Đ. Veljović, M. Hadnadjev, B. Bugarski, “Alginate/soy protein system for essential oil
3273 encapsulation with intestinal delivery”, *Carbohydr. Polym.* **2018**, 200, 15-24.
3274

3275 [411] O. Vega, J. J. Araya, M. Chavarria, E. Castellon, “Antibacterial biocomposite materials based on
3276 essential oils embedded in sol–gel hybrid silica matrices”, *J. Sol-Gel Sci. Technol.* **2016**, 79, 584–
3277 595.
3278

3279 [412] A. Jobdeedamrong, R. Jenjob, D. Crespy, “Encapsulation and release of essential oils in functional
3280 silica nanocontainers”, *Langmuir* **2018**, 34, 13235-13243.
3281

3282 [413] A. Dzimitrowicz, G. C. diCenzo, P. Swatek, P. Cyganowski, A. Stencel, D. Pogoda, P. Jamroz, P.
3283 Pohl, “Size-defined synthesis of magnetic nanorods by *Salvia hispanica* essential oil with
3284 electromagnetic excitation properties useful in microwave imaging”, *J. Magn. Mater.* **2019**, 480,
3285 87-96.
3286

3287 [414] P. Asadi Fard, S. Shakooryavan, S. Akbari, “The relationship between odour intensity and
3288 antibacterial durability of encapsulated thyme essential oil by PPI dendrimer on cotton fabrics”, *J.*
3289 *Text. I.* **2018**, 109(6), 832-841.
3290

3291 [415] M. Soheili, M. Salami, ‘*Lavandula angustifolia* biological characteristics: An in vitro study, *J.*
3292 *Cell Physiol.* **2019**, 234, 16424–16430
3293

3294 [416] E. Alexa, C. Danciu, I. Radulov, D. Obistioiu, R. M. Sumalan, A. Morar, C.A. Dehelean,
3295 ‘Phytochemical screening and biological activity of *Mentha piperita* L. and *Lavandula*
3296 *angustifolia* Mill. extracts’, *Anal. Cel. Pathol.* **2018**, Article ID 2678924.
3297

3298 [417] L. Gayoso, M. Roxo, R. Y. Cavero, M. I. Calvo, D. Ansorena, I. Astiasarán, M. Wink,
3299 ‘Bioaccessibility and biological activity of *Melissa officinalis*, *Lavandula latifolia* and *Origanum*
3300 *vulgare* extracts: Influence of an in vitro gastrointestinal digestion’, *J. Func. Foods* **2018**, 44, 146–
3301 154.
3302

3303 [418] F. Pereira, R. Baptista, D. Ladeiras, A. M. Madureira, G. Teixeira, C. Rosado, A. S. Fernandes, L.
3304 Ascensão, C. Oliveira-Silva, C. Pinto-Reis, P. Rijo, ‘Production and characterization of
3305 nanoparticles containing methanol extracts of Portuguese lavenders’, *Measurement* **2015**, 74, 170–
3306 177.
3307

3308 [419] B. Rubin, J. Manso, H. Monticelli, L. Bertazza, M. Redaelli, F. Sensi, M. Zorzan, C. Scaroni, C.
3309 Mian, M. Iacobone, D. Armanini, C. Bertolini, S. Barollo, M. Boscaro, R. Pezzani, ‘Crude extract
3310 of *Origanum vulgare* L. induced cell death and suppressed MAPK and PI3/Akt signaling pathways
3311 in SW13and H295R cell lines’, *Nat. Prod. Res.* **2019**, 33, 1646–1649.
3312

3313 [420] F. De Santis, N. Poerio, A. Gismondi, V. Nanni, G. Di Marco, R. Nisini, M. C. Thaller, A. Canini,
3314 M. Fraziano, ‘Hydroalcoholic extract from *Origanum vulgare* induces a combined anti-
3315 mycobacterial and anti-inflammatory response in innate immune cells’, *PLoS ONE* **2019**, 14,
3316 e0213150.
3317

3317 [421] B. Ponzilacqua, G. E. Rottinghaus, B. R. Landers, C. A. F. Oliveira, ‘Effects of medicinal herb
3318 and Brazilian traditional plant extracts on in vitro mycotoxin decontamination’, *Food Contr.* **2019**,
3319 *100*, 24–27.
3320

- 3321 [422] S. C. Bosco Stivanin, E. Forgiarini Vizzotto, M. de Paris, M. Balbinotti Zanela, L. Teixeira Passos,
3322 I. D. Veber Angelo, V. Fischer, 'Addition of oregano or green tea extracts into the diet for Jersey
3323 cows in transition period. Feeding and social behavior, intake and health status. Plant extracts for
3324 cows in the transition period', *Anim. Feed Sci. Techn.* **2019**, *257*, 114265.
3325
- 3326 [423] R. Lelešius, A. Karpovaitė, R. Mickienė, T. Drevinskas, N. Tiso, O. Ragažinskienė, L. Kubilienė,
3327 A. Maruška, A. Šalomska, 'In vitro antiviral activity of fifteen plant extracts against avian
3328 infectious bronchitis virus', *BMC Veter. Res.* **2019**, *15*, 178-187.
3329
- 3330 [424] M. F. Mahomoodally, G. Zengin, M. O. Aladag, H. Ozparlak, A. Diuzheva, J. Jekó, Z. Cziáky, M.
3331 Z. Aumeeruddy, 'HPLC-MS/MS chemical characterization and biological properties of *Origanum*
3332 *onites* extracts: a recent insight', *Int. J. Environ. Health Res.* **2019**, *29(6)*, 607-621.
3333
- 3334 [425] Z. Slim, M. Jancheva, S. Grigorakis, D.P. Makris, 'Polyphenol extraction from *Origanum*
3335 *dictamnus* using low-transition temperature mixtures composed of glycerol and organic salts: Effect
3336 of organic anion carbon length', *Chem. Eng. Comm.* **2018**, *205(10)*, 1494-1506.
3337
- 3338 [426] A. M. Hegazy, A. S. Abdel-Azeem, H. M. Zeidan, K. S. Ibrahim, E. M. El Sayed, 'Hypolipidemic
3339 and hepatoprotective activities of rosemary and thyme in gentamicin-treated rats', *Hum. Exper.*
3340 *Toxic.* **2018**, *37*, 420-430
3341
- 3342 [427] W. A. M. Mohamed, M. Yasmina Abd-Elhakim, S. M. Farouk, 'Protective effects of ethanolic
3343 extract of rosemary against lead-induced hepato-renal damage in rabbits', *Exper. Toxic. Pathol.*
3344 **2016**, *68*, 451-461.
3345
- 3346 [428] S. Hosseini, M. Imenshahidi, H. Hosseinzadeh, G. Karimi, 'Effects of plant extracts and bioactive
3347 compounds on attenuation of bleomycin-induced pulmonary fibrosis', *Biomed. Pharmac.* **2018**,
3348 *107*, 1454-1465.
3349
- 3350 [429] L. Wen, W. Tian-Cong, H. Dong-Mei, H. Yue, F. Ting, G. Wen-Jie, X. Qiang, 'Carnosic acid
3351 enhances the anti-lung cancer effect of cisplatin by inhibiting myeloid-derived suppressor cells',
3352 *Chinese J. Nat. Med.* **2018**, *16*, 0907-0915.
3353
- 3354 [430] M. G. Rahbardar, B. Amin, S. Mehri, S. J. Mirnajafi-Zadeh, H. Hosseinzadeh, 'Anti-inflammatory
3355 effects of ethanolic extract of *Rosmarinus officinalis* L. and rosmarinic acid in a rat model of
3356 neuropathic pain', *Biomed. Pharmac.* **2017**, *86*, 441-449.
3357
- 3358 [431] E. Horváthová, A. Srančíková, E. Regendová-Sedláčková, M. Melušová, V. Meluš, J. Netriová,
3359 Z. Krajčovičová, D. Slameňová, M. Pastorek, K. Kozics, 'Enriching the drinking water of rats with
3360 extracts of *Salvia officinalis* and *Thymus vulgaris* increases their resistance to oxidative stress',
3361 *Mutagenesis* **2016**, *31*, 51-59
3362
- 3363 [432] Z. El Gabbas, K. Bezza, J. Laadraoui, R. Makbal, R. Aboufatima, A. Chait, '*Salvia officinalis*
3364 induces antidepressant-like effect, anxiolytic activity and learning improvement in hippocampal
3365 lesioned and intact adult rats', *Bangladesh J. Pharmac.* **2018**, *13*, 367-378.
3366
- 3367 [433] M. Cheurfa, R. Allem, 'Effect of some plant extracts on pathogenic bacteria responsible for
3368 gastroenteritis', *Phytothérapie* **2017**, *15*, 395-400.
3369
- 3370 [434] M. I. Kazeem, A. O. T. Ashafa, M. O. Nafiu, 'Biological activities of three Nigerian spices –
3371 *Laurus nobilis* Linn, *Murraya koenigii* (L) Spreng and *Thymus vulgaris* Linn.', *Trop. J. Pharmac.*
3372 *Res.* **2015**, *14*, 2255-2261.
3373
- 3374 [435] M. Proks, A. Heghes, A. Cheveresan, S. Nita, M. Voicu, V. Buda, D. Ionescu, L. Nita, C.
3375 Trandafirescu, V. Paunescu, 'Thyme leaves aqueous extract and its formulations: A comparative
3376 study based on chemical structures and biological activity', *Rev. Chim.* **2019**, *70*, 1875-1878.
3377

- 3378 [436] T. Khouya, M. Ramchoun, A. Hmidani, S. Amrani, H. Harnafi, M. Benlyas, Y.F. Zegzouti, C.
3379 Alem, 'Anti-inflammatory, anticoagulant and antioxidant effects of aqueous extracts from
3380 Moroccan thyme varieties', *Asian Pacific J. Trop. Biomed.* **2015**, *5*, 636–644.
3381
- 3382 [437] D. R. Johnson, E. A. Decker, 'The role of oxygen in lipid oxidation reactions: a review', *Ann. Rev.*
3383 *Food Sci. Techn.* **2015**, *6*, 171–190.
3384
- 3385 [438] M. Aminzare, M. Hashemi, E. Ansarian, M. Bimkar, H. H. Azar, M. R. Mehrasbi, S.
3386 Daneshamooz, M. Raeisi, B. Jannat, A. Afshari, 'Using natural antioxidants in meat and meat
3387 products as preservatives: a review', *Adv. Anim. Vet. Sci.* **2019**, *7(5)*, 417-426.
3388
- 3389 [439] M. Hashemi, A. Ehsani, A. Afshari, M. Aminzare, M. Raeisi M. 'Chemical composition and
3390 antifungal effect of *Echinophora platyloba* essential oil against *Aspergillus flavus*, *Penicillium*
3391 *expansum* and *Fusarium graminearum*', *J. Chem. Health Risks.* **2016**, *6*, 91-97.
3392
- 3393 [440] J. Dai, R. J. Mumper, 'Plant phenolics: Extraction, analysis and their antioxidant and anticancer
3394 properties', *Molecules* **2010**, *15*, 7313–7352
3395
- 3396 [441] M. Alanon, L. Marchante, M. Diaz-Maroto, M. Perez-Coello, M. 'Extraction of natural flavorings
3397 with antioxidant capacity from cooperage by-products by green extraction procedure with
3398 subcritical fluids', *Ind. Crops Prod.* **2017**, *103*, 222-232.
3399
- 3400 [442] A. B. Falowo, P. O. Fayemi, Muchenje 'Natural antioxidants against lipid–protein oxidative
3401 deterioration in meat and meat products: A review', *Food Res. Int.* **2014**, *64*,171-181
3402
- 3403 [443] J. Souza-Ribeiro, M. J. M. Cordeiro-Santos, L. K. Rosa-Silva, L. C. Lavinsky Pereira, I. Alves-
3404 Santos, S. C. da Silva-Lannes, M. Viana-da Silva, 'Natural antioxidants used in meat products: A
3405 brief review', *Meat Sci.* **2019**, *148*, 181–188
3406
- 3407 [444] T. L. Mc Carthy, J. P. Kerry, J. F. Kerry, P. B. Lynch, D. J. Buckley, 'Assessment of the
3408 antioxidant potential of natural food and plant extracts in fresh and previously frozen pork patties',
3409 *Meat Sci.* **2001**, *57*, 177–184
3410
- 3411 [445] T. Li, J. Li, W. Hu, X. Zhang, X. Li, J. Zhao, 'Shelf-life extension of crucian carp (*Carassius*
3412 *auratus*) using natural preservatives during chilled storage', *Food Chem.* **2012**, *135*, 140–145.
3413
- 3414 [446] L. R. Nissen, D. V. Byrne, G. Bertelsen, L. H. Skibsted, 'The antioxidative activity of plant extracts
3415 in cooked pork patties as evaluated by descriptive sensory profiling and chemical analysis', *Meat*
3416 *Sci.*, **2004**, *68*, 485–495.
3417
- 3418 [447] H. Zhang, J. Wu, X. Guo. 'Effects of antimicrobial and antioxidant activities of spice extracts on
3419 raw chicken meat quality', *Food Sci. Hum. Wellness* **2016**, *5*, 39–48
3420
- 3421 [448] F. A. Khalafalla, F. H. M. Ali, A. -R. H. A. Hassan, 'Quality improvement and shelf-life extension
3422 of refrigerated Nile tilapia (*Oreochromis niloticus*) fillets using natural herbs', *Beni-Suef J- Bas-*
3423 *Appl. Sci.* **2015**, *4(1)*, 33-40.
3424
- 3425 [449] C. Axel, E. Zannini, E. K. Arendt, 'Mold spoilage of bread and its biopreservation: A review of
3426 current strategies for bread shelf life extension', *Crit. Rev. Food Sci. Nutr.* **2017**, *57*, 3528-3542
3427
- 3428 [450] L. da Cruz-Cabral, V. Fernández-Pinto, A. Patriarca, 'Application of plant derived compounds to
3429 control fungal spoilage and mycotoxin production in foods', *Int. J. Food Microbiol.* **2013**, *166*, 1–
3430 14.
3431
- 3432 [451] O. C. Matos, M. G. Barreiro, Safety use of bioactive products of plant origin for the control of post
3433 harvested fungal diseases of "Rocha" pear. IV Simposium Ibérico de Maturação e pós-colheita:
3434 Frutos Hortícolas. Livro de Actas, 2004, pp. 525–529
3435

- 3436 [452] B. Shan, Y. -Z. Cai, J. D. Brooks, H. Corke, 'Potential application of spice and herb extracts as
3437 natural preservatives in cheese', *J. Med. Food* **2011**, *14*, 3.
3438
- 3439 [453] P. R Salgado, C. M. Ortiz, Y. S. Musso, L. Di Giorgio, A. N. Mauri, 'Edible films and coatings
3440 containing bioactives', *Cur. Opin. Food Sci.* **2015**, *5*, 86–92, and references therein.
3441
- 3442 [454] C. Frezza, A. Venditti, M. Serafini, A. Bianco, "Chapter 4 – Phytochemistry, Chemotaxonomy,
3443 Ethnopharmacology, and Nutraceuticals of Lamiaceae", in Atta-Ur-Rahman (Ed.) *Studies in Natural*
3444 *Product Chemistry* **2019**, *62*, pp. 125-178.
3445
- 3446 [455] D. Piñeros-Hernandez, C. Medina-Jaramillo, A. López-Cordóba, S. Goyanes, 'Edible cassava
3447 starch films carrying rosemary antioxidant extracts for potential use as active food packaging',
3448 *Food Hydrocol.* **2017**, *63*, 488-495.
3449
- 3450 [456] J. Gómez-Estaca, P. Montero, F. Fernández-Martín, A. Alemán, M. C. Gómez-Guillén, 'Physical
3451 and chemical properties of tuna-skin and bovine hide gelatin films with added aqueous oregano
3452 and rosemary extracts', *Food Hydrocol.* **2009**, *23(5)*, 1334-1341.
3453
- 3454 [457] A. C. Seydim, G. Sarikus, 'Antimicrobial activity of whey protein based edible films incorporated
3455 with oregano, rosemary and garlic essential oils', *Food Res. Int.* **2006**, *39(5)*, 639-644.
3456
- 3457 [458] D. Georgantelis, G. Blekas, P. Katikou, I. Ambrosiadis, D. J. Fletouris, 'Effect of rosemary
3458 extract, chitosan and α -tocopherol on lipid oxidation and colour stability during frozen storage of
3459 beef burgers', *Meat Sci.* **2007**, *75(2)*, 256–264.
3460
- 3461 [459] T. Li, W. Hu, J. Li, X. Zhang, J. Zhu, X. Li, 'Coating effects of tea polyphenols and rosemary
3462 extract combined with chitosan on the storage quality of large yellow croaker (*Pseudosciaena*
3463 *crocea*)', *Food Contr.* **2012**, *25(1)*, 101–106.
3464
- 3465 [460] A. G. Ponce, S. I. Roura, C. E. Del Valle, M. R. Moreira, 'Antimicrobial and antioxidant activities
3466 of edible coating enriched with natural plant extract: in vitro and in vivo studies', *Postharv. Biol.*
3467 *Techn.* **2008**, *49(2)*, 294–300.
3468
- 3469 [461] M. L. Rooney, Overview of Active Packaging. 1995, in M. I. Rooney (Ed.) *Active Food*
3470 *Packaging*. Blackie Academic and Professional, Glasgow, UK, 1995, pp. 1–37, ISBN 978–1-
3471 4613–5910–4
3472
- 3473 [462] R. Dobrucka, R. Cierpiszewski, 'Active and Intelligent Packaging Food – Research and
3474 Development – A Review', *Polish J. Food Nutr. Sci.* **2014**, *64(1)*, 7-15.
3475
- 3476 [463] T. Hutton, *Food Packaging: An introduction*. Key topics in food science and technology. Chipping
3477 Campden, Gloucestershire, UK: Campden and Chorleywood Food Research Association Group,
3478 **2003**, *7*, 108.
3479
- 3480 [464] D. Restuccia, U. G. Spizzirri, O. I. Parisi, G. Cirillo, M. Curcio, F. Iemma, F. Puoci, G. Vinci, N.
3481 Picci, 'New EU regulation aspects and global market of active and intelligent packaging for food
3482 industry applications', *Food Contr.* **2010**, *21*, 1425–1435.
3483
- 3484 [465] C. Nerin, L. Tovar, D. Djenane, J. Camo, J. Salafranca, J. A. Beltrán, P. Roncalés, 'Stabilization
3485 of beef meat by a new active packaging containing natural antioxidants', *J. Agric. Food Chem.*
3486 **2006**, *54*, 7840-7846.
3487
- 3488 [466] C. Xiao, L. Zhu, W. Luo, X. Song, Y. Deng, 'Combined action of pure oxygen pretreatment and
3489 chitosan coating incorporated with rosemary extracts on the quality of fresh-cut pears', *Food*
3490 *Chem.* **2010**, *121*, 1003–1009.
3491

- 3492 [467] S. Nimsha, A. Weerakkody, A. Nola Caffin, S. Mark, A. Turner, A. Gary, B. C. Dykes, 'In vitro
3493 antimicrobial activity of less-utilized spice and herb extracts against selected foodborne bacteria',
3494 *Food Contr.* **2010**, *21(1)*, 1408- 1414.
3495
- 3496 [468] J. Camo, A. Lorés, D. Djenane, J. A. Beltrán, P. Roncalés, 'Display life of beef packaged with an
3497 antioxidant active film as a function of the concentration of oregano extract', *Meat Sci.* **2011**, *88*,
3498 174–178.
3499
- 3500 [469] K. Bentayeb, P. Vera, C. Rubio, C. Nerin, 'Adaptation of the ORAC assay to the common
3501 laboratory equipment and subsequent application to antioxidant plastic films', *Anal. Bioanal.*
3502 *Chem.* **2009**, *394*, 903–910.
3503
- 3504 [470] P. De Cruz, L. Prideaux, J. Wagner, S. C. Ng, C. McSweeney, C. Kirkwood, M. Morrison, M. A.
3505 Kamm, 'Characterization of the gastrointestinal microbiota in health and inflammatory bowel
3506 disease' *Inflamm. Bowel Dis.* **2012**, *18*, 372-390.
3507
- 3508 [471] L. V. Hooper, D. R. Littman, A. J. Macpherson, 'Interactions between the microbiota and the
3509 immune system', *Science* **2012**, *336*, 1268–1273.
3510
- 3511 [472] C. Chang, H. Lin, 'Dysbiosis in gastrointestinal disorders', *Best Pract. Res. Clin. Gastroenter.*
3512 **2016**, *30*, 3–15.
3513
- 3514 [473] N. A. Nasef, S. Mehta, L. R. Ferguson, 'Dietary interactions with the bacterial sensing machinery
3515 in the intestine: the plant polyphenol case', *Front. Genet.* **2014**, *5*, 1-14.
3516
- 3517 [474] F. De Filippis, N. Pellegrini, L. Vannini, I. B. Jeffery, A. La Stora, L. Laghi, D. I. Serrazanetti,
3518 R. Di Cagno, I. Ferrocino, C. Lazzi, S. Turrioni, L. Cocolin, P. Brigidi, E. Neviani, M. Gobbetti,
3519 P.W. O'Toole, D. Ercolini, 'High-level adherence to a Mediterranean diet beneficially impacts the
3520 gut microbiota and associated metabolome', *Gut* **2016**, *65*, 1812–1821.
3521
- 3522 [475] T. A. Thumann, E. -M. Pferschy-Wenziga, C. Moissl-Eichinger, R. Bauer, 'The role of gut
3523 microbiota for the activity of medicinal plants traditionally used in the European Union for
3524 gastrointestinal disorders', *J. Ethnopharm.* **2019**, *245*, 112153.
3525
- 3526 [476] G. A. Gonçalves, R. C. G. Corrêa, L. Barros, M. I. Dias, R. C. Calhelha, V. G. Correa, A. Bracht,
3527 R. M. Peralta, I. C. F. R. Ferreira, 'Effects of *in vitro* gastrointestinal digestion and colonic
3528 fermentation on a rosemary (*Rosmarinus officinalis* L) extract rich in rosmarinic acid', *Food*
3529 *Chem.* **2019**, *271*, 393-400.
3530
- 3531 [477] A. R. Madureira, D. Campos, B. Gullon, C. Marques, L. M. Rodríguez-Alcalá, C. Calhau, J. L.
3532 Alonso, B. Sarmiento, A. M. Gomes, M. Pintado, 'Fermentation of bioactive solid lipid
3533 nanoparticles by human gut microflora', *Food Funct.* **2016**, *7*, 516–529.
3534

Table 1. Extract of the main recent publications on endemisms of Lavender and chemical variability of its essential oils

Species	Plant material	Origin	Yield (%)	Main components	Ref.
<i>L. hybrida</i>	Whole plant	Romania	N.R.	linalyl acetate (23%), linalool (22%), terpinene-4-ol (17%)	[45]
<i>L. hybrida</i> ‘Budrovka’	Leaves	Croatia	3.3	linalool (57%), linalyl acetate (10%), 1,8-cineole (8%)	[36]
<i>L. hybrida</i>	Aerial parts	Iran	0.5-1.5	1,8-cineole (32-48%), borneol (17-26%), camphor (8-13%)	[46]
<i>L. hybrida</i> ‘Grosso’	Aerial parts	Italy	8.2-8.5	linalool (34-43%), linalyl acetate (15-17%), 1,8-cineole (8%)	[35]
<i>L. hybrida</i> ‘ SuperA’	Whole plant	Italy	8.8-9.3	linalool (34-39%), linalyl acetate (16-21%), 1,8- cineole (6-8%)	[35]
<i>L. hybrida</i> ‘SuperA’	Aerial parts	Turkey	N.R.	linalool (37%), linalyl acetate (33%), camphor (5%)	[39]
<i>L. hybrida</i> ‘ Sumiens’	Whole plant	Italy	6.4-6.9	linalool (40-41%), 1,8-cineole (16%), borneol (4%)	[41]
<i>L. hybrida</i> “Grey hedge”	Aerial parts	Turkey	N.R.	linalool (29%), 1,8-cineole (16%), borneol (8%)	[39]
<i>L. angustifolia</i>	Aerial parts	Algeria	2.0	1,8-cineole (29%), camphor (25%), borneol (4%)	[41]
<i>L. angustifolia</i>	Aerial parts	India	0.55	linalyl acetate (39%), linalool (30%), terpinene-4-ol (4%)	[40]
<i>L. angustifolia</i>	Inflorescens	Croatia	0.9	linalool (54%), linalyl acetate (12%), lavandulol (7%)	[36]
<i>L. angustifolia</i>	Inflorescens	Turkey	3.5-6.0	linalool (32-50%), linalyl acetate (2-18%)	[37]
<i>L. angustifolia</i>	Whole plant	Turkey	N.R.	linalyl acetate (29-47%), linalool (28-36%)	[39]
<i>L. angustifolia</i>	Whole plant	Spain	N.R.	linalool (37-54%), linalyl acetate (21- 36%), (<i>E</i>)-caryophyllene (1-3%)	[38]
<i>L. angustifolia</i>	Inflorescens	Poland	N.R.	linalool (fresh and dried 35%), linalyl acetate (fresh 23%, dried 20%)	[34]
<i>L. angustifolia</i>	Aerial parts	Poland	N.R.	linalool (fresh 31%, dried 27%), linalyl acetate (fresh and dried 23%)	[34]
<i>L. angustifolia</i> ‘mailette’	Aerial parts	Italy	4.5-5.1	linalool (46-48%), linalyl acetate (26%), α -terpineol (7-57%)	[35]
<i>L. mairei</i> (w)	Aerial parts	Morocco	1.0	carvacrol (78%), terpinolene (3%), octen-3-ol (2%)	[47]
<i>L. mairei</i> (w)	Aerial parts	Morocco	1.2	carvacrol (77%), terpinolene (3%), octen-3-ol (2%)	[47]
<i>L. pinnata</i>	Inflorescens	Spain	N.R.	carvacrol (68%), caryophyllene oxide (15%), spathulenol (12%)	[48]
<i>L. pinnata</i>	Leaves	Spain	N.R.	carvacrol (84%), carvacrol methyl ether (3%)	[48]
<i>L. pubescens</i>	Aerial parts	Yemen	1.3	carvacrol (73%), carvacrol methyl ether (7%), caryophyllene oxide (6%)	[49]
<i>L. stricta</i>	Aerial parts	Iran	0.2-0.4	α -pinene (58-63%), linalool (9%), 3-methyl-2-methyl butanoate (7-8%)	[50]
<i>L. stoechas</i>	Whole plant	Spain	0.3-1.0	fenchone (33-37%), camphor (16-24%), 1,8-cineole (17-18%)	[42]
<i>L. stoechas</i>	Aerial parts	Italy	0.7-1.1	fenchone (45-60%), camphor (8-2%), 1,8-cineole (1-16%)	[43]
<i>L. stoechas</i>	Inflorescens	Italy	0.4-0.5	fenchone (53-71%), camphor (7-12%), 1,8-cineole (1-13%)	[44]
<i>L. stoechas</i>	Aerial parts	Algeria	1.5	fenchone (50%), camphor (14%), bornyl acetate (6%)	[51]
<i>L. latifolia</i>	Whole plant	Spain	N.R.	linalool (35-51%), 1,8-cineole (26-32%), camphor (10-18%)	[38]

(w) = wild; N.R. = not reported

Table 2. Extract of the main recent publications on endemisms of *Oregano* and chemical variability of its essential oils

Species	Plant material	Origin	Yield (%)	Main components	Ref.
<i>O. acutidens</i>	Aerial parts	Turkey	0.5	carvacrol (62%), <i>p</i> -cymene (16%), thymol (13%)	[72]
<i>O. compactum</i>	Aerial parts	Morocco	1.2-4.4	carvacrol (2-64%), <i>p</i> -cymene (7-43%), thymol (<1-42.%)	[67]
<i>O. compactum</i>	Aerial parts	Morocco	0.6-2.9	carvacrol (2-79%), thymol (<1-56%), <i>p</i> -cymene (4-48%)	[68]
<i>Origanum x majoricum</i>	Aerial parts	Argentina	N.R.	<i>trans</i> -sabinene hydrate (24-28%), thymol (12-17%), γ -terpinene (7-8%)	[59]
<i>O. vulgare</i>	Aerial parts	Kosovo	0.4-0.8	sabinene (2-12%), 1,8- cineole (1-14%), caryophyllene oxide (<1-38%)	[58]
<i>O. vulgare</i> (w)	Aerial parts	Italy	2.1-6.2	thymol (21-63%), γ -terpinene (6-24%), <i>p</i> -cymene (4-12%)	[55]
<i>O. vulgare</i>	Aerial parts	Iran	0.1-1.8	carvacrol (<1-47%), linalyl acetate (<1-44%), <i>Z</i> - β -bisabolene (nd-40%)	[57]
<i>O. vulgare</i>	Aerial parts	Spain	0.8-2.0	carvacrol (59-77%), γ -terpinene (2-11%), <i>p</i> -cymene (4-8%)	[56]
<i>O. vulgare</i> ssp. <i>vulgare</i>	Aerial parts	Argentina	N.R.	<i>trans</i> -sabinene-hydrate (23-27%), thymol (14-17%), terpinene-4-ol (8-11%)	[59]
<i>O. vulgare</i> ssp. <i>vulgare</i>	Aerial parts	Poland	0.3-0.9	sabinene (2-25%), 1,8-cineole (<1-15%), linalool (3-32%)	[74]
<i>O. vulgare</i> ssp. <i>vulgare</i> (w)	Aerial parts	Montenegro	0.7-1.2	germacrene D (16-28%), α -terpineol (15-18%), (<i>E</i>)-caryophyllene (8-15%)	[64]
<i>O. vulgare</i> ssp. <i>vulgare</i> (w)	Aerial parts	India	0.2-2.1	thymol (14-86%), carvacrol (nd-63%)	[75]
<i>O. vulgare</i> ssp. <i>hirtum</i>	Aerial parts	Argentina	N.R.	<i>trans</i> -sabinene hydrate (18-23%), thymol (17-19%), γ -terpinene (7-8%)	[59]
<i>O. vulgare</i> ssp. <i>hirtum</i>	Aerial parts	Italy	2.3-6.2	thymol (24-54%), γ -terpinene (10- 31%), <i>p</i> -cymene (5-19%)	[61-62]
<i>O. vulgare</i> ssp. <i>hirtum</i>	Aerial parts	Italy	1.8-6.4	thymol (11-54%), <i>p</i> -cymene (3-19%), γ -terpinene (8-53%)	[60]
<i>O. vulgare</i> ssp. <i>hirtum</i>	Aerial parts	Italy	1.0-2.7	thymol/carvacrol (65-85%), linalool (<1-3%)	[63]
<i>O. vulgare</i> ssp. <i>hirtum</i> (w)	Aerial parts	Montenegro	3.0	carvacrol (74%), <i>p</i> -cymene (8%), γ -terpinene (7%)	[64]
<i>O. vulgare</i> ssp. <i>gracile</i>	Aerial parts	Turkey	0.2-0.5	thymol (8-33%), <i>p</i> -cymene (3-24%), γ -terpinene (9-30%)	[76]
<i>O. vulgare</i> ssp. <i>gracile</i>	Leaves	Iran	1.4	carvacrol (47%), γ -terpinene (14%), <i>p</i> -cymene (14%)	[77]
<i>O. vulgare</i> ssp. <i>gracile</i>	Flowers	Iran	2.4	carvacrol (61%), γ -terpinene (17%), <i>p</i> -cimene (7%)	[77]
<i>O. dictamnus</i>	Aerial parts	Greece	1.2	<i>p</i> -cymene (33%), carvacrol (15%), linalool (7%)	[71]
<i>O. microphyllum</i>	Aerial parts	Greece	0.3	terpinen-4-ol (16%), carvacrol (13%), sabinene (8%)	[71]
<i>O. libanoticum</i>	Aerial parts	Lebanon	0.16	hexadecanoic acid (11%), linalool (9%), 3-methyl-2-methyl butanoate (7-8%)	[50]

<i>O. libanoticum</i>	Aerial parts	Lebanon	0.16	(<i>E</i>)-caryophyllene (27%), caryophyllene oxide (23%), germacrene D (17%)	[70]
<i>O. dubium</i>	Aerial parts	Turkey	8.0-11.1	carvacrol (82-91%), γ -terpinene (2-6%), <i>p</i> -cymene (2-4%)	[78]
<i>O. ehrenbergii</i>	Aerial parts	Lebanon	3.8-4.5	carvacrol (48-90%), <i>p</i> -cymene (1-21%), γ -terpinene (<1-7%)	[69]
<i>O. ehrenbergii</i>	Aerial parts	Lebanon	3.2	carvacrol (79%), <i>p</i> -cymene (4%), γ -terpinene (3%)	[70]
<i>O. floribundum</i>	Aerial parts	Algeria	2.5-5.8	carvacrol (32-61%), γ -terpinene (3-26%), <i>p</i> -cymene (9-43%)	[73]
<i>O. syriacum</i>	Aerial parts	Lebanon	1.6	carvacrol (61%), <i>p</i> -cymene (8%), γ -terpinene (8%)	[70]
<i>O. syriacum</i> (w)	Whole plant	Egypt	4.6	thymol (32%), γ -terpinene (14%), linalool (9%)	[79]
<i>O. syriacum</i>	Whole plant	Egypt	5.5	carvacrol (81%), <i>p</i> -cymene (9%), γ -terpinene (2%)	[79]
<i>O. hypericifolium</i>	Aerial parts	Turkey	1.5	<i>p</i> -cymene (33%), thymol (23%), γ -terpinene (15%)	[80]
<i>O. syleum</i>	Aerial parts	Turkey	1.1	<i>p</i> -cymene (13%), carvacrol (13%), α -pinene (11%)	[80]
<i>O. majorana</i>	Aerial parts	Turkey	1.9	linalool (38%), thymol (26%), γ -terpinene (12%)	[80]
<i>O. majorana</i>	Aerial parts	Tunisia	1.8	terpinen-4-ol (23%), <i>cis</i> -sabinene hydrate (18%), γ -terpinene (11%)	[81]
<i>O. majorana</i>	Leaves	Egypt	2.3-2.6	<i>cis</i> -sabinene hydrate (15-34%), γ -terpinene (9-29%), terpinen-4-ol (14-35%)	[82]
<i>O. onites</i>	Aerial parts	Turkey	2.0	thymol (42%), linalool (14%), carvacrol (9%)	[80]
<i>O. onites</i>	Inflorescens	Greece	3.0-7.0	carvacrol (69-93%), <i>p</i> -cymene (nd-10%), γ -terpinene (nd-8%)	[83]
<i>O. minutiflorum</i>	Aerial parts	Turkey	1.1	carvacrol (91%), linalool (3%), (<i>E</i>)-caryophyllene (1%)	[84]
<i>O. rotundifolium</i>	Aerial parts	Turkey	1.0	carvacrol (57%), <i>p</i> -cymene (13%), (<i>Z</i>)- β -ocimene (5%)	[85]

(w) = wild; nd = not detected; N.R. = not reported

Table 3. Extract of the main recent publications on endemism of Rosemary and chemical variability of its essential oils

Species	Plant material	Origin	Yield (%)	Main components	Ref.
<i>R. eriocalix</i>	Stems	Algeria	0.02	camphor (41%), camphene (10%), α -pinene (8%)	[87]
<i>R. eriocalix</i>	Leaves	Algeria	0.7	camphor (37%), α -pinene (18%), camphene (16%)	[87]
<i>R. eriocalix</i>	Flowers	Algeria	0.9	camphor (30%), α -pinene (15%), camphene (13%)	[87]
<i>R. officinalis</i> (w)	Aerial parts	Italy	0.8-3.6	α -pinene (10-46%), 1,8-cineole (<1-67%), camphor (<1-27%)	[88]
<i>R. officinalis</i>	Aerial parts	Jordan	0.7	1,8-cineole (31%), α -pinene (17%), camphor (12%)	[89]
<i>R. officinalis</i>	Aerial parts	Italy	0.1-0.9	α -pinene (14-34%), 1,8-cineole (11-28%)	[90]
<i>R. officinalis</i> (w)	Aerial parts	Tunisia	0.7-1.7	α -pinene (7-10%), 1,8-cineole (23-53%), camphor (7-28%)	[91]
<i>R. officinalis</i> (w)	Aerial parts	Tunisia	1.5-2.1	1,8-cineole (29-60%), camphor (6-28%), α -pinene (7-13%)	[92]
<i>R. officinalis</i> (w)	Aerial parts	Algeria	0.7-1.9	1,8-cineole (2-48%), camphor (3-41%), α -pinene (14-29%)	[93]
<i>R. officinalis</i> (w)	Aerial parts	Italy	0.7-2.8	1,8-cineole (<1-63%), α -pinene (12-40%), camphor (3-31%)	[94]
<i>R. officinalis</i>	Leaves	Iran	0.5-2.6	1,8-cineole (5-28%), camphor (2-25%), α -pinene (14-21%)	[95]
<i>R. officinalis</i> (w)	Leaves	Iran	0.6-2.3	1,8-cineole (6-27%), camphor (2-25%), α -pinene (15-21%)	[96]
<i>R. officinalis</i> (w)	Aerial parts	Italy	0.8-2.3	1,8-cineole (8-59%), α -pinene (14-27%), camphor (3-15%)	[97]
<i>R. officinalis</i> (w)	Aerial parts	Italy	N.R.	1,8-cineole (nd-48%), α -pinene (9-44%), camphor (<1-23%)	[98]
<i>R. officinalis</i>	Aerial parts	Tunisia	N.R.	1,8-cineole (24%), camphene (13%), camphor (13%)	[99]
<i>R. officinalis</i>	Aerial parts	Italy	N.R.	1,8-cineole (2-48%), α -pinene (9-44%), camphor (1-13%)	[100]

(w) = wild; nd = not detected; N.R. = not reported

Table 4. Extract of the main recent publications on endemism of Sage and chemical variability of its essential oils

Species	Plant material	Origin	Yield (%)	Main components	Ref.
<i>S. algeriensis</i>	Leaves	Algeria	0.8	benzaldehyde (10%), eugenol (9%), phenylethyl alcohol (8%)	[118]
<i>S. algeriensis</i>	Flowers	Algeria	1.5	viridiflorol (71%), globulol (9%), α -cadinene (3%)	[118]
<i>S. fruticos</i> ssp. <i>thomasi</i>	Aerial parts	Italy	0.2	1,8-cineole (11-26%), β -pinene (13-14%), camphor (8-9%)	[111]
<i>S. hispanica</i>	Aerial parts	Italy	0.05	(<i>Z</i>)-caryophyllene (12%), (<i>E</i>)-caryophyllene (11%), humulene (5%)	[105]
<i>S. trichoclada</i>	Aerial parts	Turkey	N.R.	caryophyllene oxide (25%), spathulenol (15%), β -pinene(12%)	[106]
<i>S. virgate</i>	Aerial parts	Turkey	N.R.	1,8-cineole (20%), α -copaene (19%), germacrene D (18%)	[106]
<i>S. virgate</i>	Aerial parts	Iran	0.03	(<i>E</i>)-caryophyllene (34-38%), caryophyllene oxide (26-29%), β -farnesene (9-10%)	[109]
<i>S. ceratophilla</i>	Aerial parts	Turkey	N.R.	germacrene D (24%), α - copaene (19%), 1,8-cineole (8%)	[106]
<i>S. multicaulis</i>	Aerial parts	Turkey	N.R.	caryophyllene oxide (23%), spathulenol (13%), β -pinene (8%)	[106]
<i>S. multicaulis</i>	Aerial parts	Iran	0.2-0.5	(<i>E</i>)-caryophyllene (11-35%), α -pinene (12-17%), linalyl acetate (10-14%)	[108]
<i>S. multicaulis</i>	Aerial parts	Lebanon	0.7-1.3	<i>trans</i> -nerolidol (nd-12%), 1,8-cineole (7-16%), δ -cadinene (2-5%)	[119]
<i>S. lavandulifolia</i>	Aerial parts	Spain	N.R.	1,8-cineole (19%), camphor (11%), α -pinene (10%)	[112]
<i>S. nemorosa</i>	Aerial parts	Iran	N.R.	caryophyllene oxide (11-35%), spathulenol (4-37%), (<i>E</i>)-caryophyllene (2-27%)	[109]
<i>S. officinalis</i>	Aerial parts	Ethiopia	1.0-1.2	α -thujone (20-26%), 1,8-cineole (17-22%), camphor (13-18%)	[102]
<i>S. officinalis</i>	Aerial parts	Montenegro	1.8-2.8	α -thujone (17-40%), camphor (13-35%), 1,8-cineole (6-12%)	[103]
<i>S. officinalis</i>	Aerial parts	Spain	0.8-1.5	α -thujone (23-42%), camphor (11-20%), 1,8-cineole (5-16%)	[104]
<i>S. reuteriana</i>	Aerial parts	Iran	0.2-0.4	α -gurjunene (5-14%), β -elemene (5-14%), germacrene D (3-7%)	[120]
<i>S. reuteriana</i>	Aerial parts	Iran	0.1-0.3	caryophyllene oxide (30-52%), spathulenol (12-15%), (<i>E</i>)-caryophyllene (5-10%)	[108]
<i>S. sclarea</i>	Flowers	Iran	0.08	germacrene D (21%), α -bulnesene (12%), limonene (12%)	[121]
<i>S. sclarea</i>	Leaves	Iran	0.05	germacrene D (21%), caryophyllene (17%), caryophyllene oxide (10%)	[121]
<i>S. syriaca</i>	Aerial parts	Iran	0.1	germacrene B (26-36%), germacrene D (18-24%), bicyclogermacrene (9-15%)	[108]
<i>S. tomentosa</i>	Aerial parts	Greece	1.1-3.3	β -pinene (1-30%), α -pinene (11-25%), α -thujone (nd-24%)	[122]
<i>S. sericeo-tomentosa</i>	Aerial parts	Turkey	N.R.	sabinyl acetate (80%), α -pinene (3-4%)	[123]
<i>S. adenophylla</i>	Aerial parts	Turkey	0.28	α -pinene (16%), β -pinene (14%)	[124]
<i>S. viscosa</i>	Aerial parts	Turkey	0.16	β -pinene (25%), myrcene (9%), humulene (8%)	[124]
<i>S. argentea</i>	Aerial parts	Italy	0.27	14-hydroxy- α -humulene (40%), 1,3,8- <i>p</i> -menthatriene (12%)	[125]

<i>S. aristata</i>	Aerial parts	Iran	0.02	carvacrol (23%), 1,8-cineole (8%), α -pinene (7%)	[126]
<i>S. aucheri</i> spp. <i>blancoana</i>	Aerial parts	Morocco	1.4	camphor (27%), camphene (22%), α -pinene (21%)	[127]
<i>S. aucheri</i> spp. <i>blancoana</i> (w)	Aerial parts	Morocco	1.2	camphor (57%), camphene (6%), (<i>E</i>)-caryophyllene (5%)	[127]
<i>S. bicolor</i>	Aerial parts	Egypt	0.3	α -pinene (25%), α -thujone (13%), (<i>E</i>)-caryophyllene (7%)	[128]
<i>S. broussonetti</i>	Aerial parts	Italy	<0.01	α -gurjunene (6-16%), α -pinene (11-15%), germacrene D (7-14%)	[129]
<i>S. buchananii</i>	Aerial parts	Algeria	0.9	1,8-cineole (23%), α -pinene (20%), camphene (17%)	[113]
<i>S. bucharica</i>	Aerial parts	Pakistan	0.17	1,8-cineole (26%), camphor (16%), (<i>E</i>)-caryophyllene (11%)	[114]
<i>S. chudaei</i>	Aerial parts	Algeria	0.55	bornyl acetate (21%), β -eudesmol (14%), (<i>E</i>)-caryophyllene (11%)	[130]
<i>S. desoleana</i>	Aerial parts	Italy	N.R.	germacrene D (4-26%), linalyl acetate (<1-38%), 1,8-cineole (8-11%)	[131]
<i>S. forskahlei</i> (w)	Aerial parts	Turkey	0.1	germacrene D (19%), δ -cadinene (13%), neophytadiene (13%)	[110]
<i>S. forskahlei</i>	Aerial parts	Turkey	0.01	(<i>E</i>)-caryophyllene (24%), α -selinene (13%), naphtalene (11%)	[110]
<i>S. hypoeuca</i>	Inflorescens	Iran	0.1-0.8	bicyclogermacrene (2-37%), (<i>E</i>)-caryophyllene (2-22%)	[132]
<i>S. kiansiensis</i>	Leaves	China	0.1	epimanol (46%), globulol (6%), α -guaiene (4%)	[133]
<i>S. lachnocalyx</i>	Aerial parts	Iran	N.R.	bicyclogermacrene (41%), (<i>E</i>)-caryophyllene (11%), spathulenol (9%)	[134]
<i>S. lachnocalyx</i>	Aerial parts	Iran	N.R.	bicyclogermacrene (18%), α -pinene (15%), β -pinene (14%)	[134]
<i>S. leriifolia</i>	Aerial parts	Iran	0.4	β -pinene (14%), 1,8-cineole (14%), α -pinene (10%)	[135]
<i>S. limbata</i>	Aerial parts	Iran	0.4	caryophyllene oxide (12%), terpinen-4-ol (9%), sabinene (8%)	[136]
<i>S. montbretti</i>	Aerial parts	Turkey	<0.1	(<i>E</i>)-caryophyllene (33%), β -pinene (10%), humulene (8%)	[107]
<i>S. palaestina</i>	Leaves	Palestine	0.1-0.7	1,8-cineole (52-63%), camphor (1-9%), (<i>E</i>)-caryophyllene (1-5%)	[115]
<i>S. ringens</i>	Aerial parts	Macedonia	0.2	1,8-cineole (32%), camphene (17%), borneol (12%)	[116]
<i>S. sahendica</i>	Aerial parts	Iran	1.5-1.8	α -pinene (14-15%), β -pinene (11-13%), germacrene D (3-8%)	[137]
<i>S. santolinifolia</i>	Aerial parts	Iran	1.6	α -pinene (49%), β -eudesmol (20%), camphene (8%)	[138]
<i>S. sclareopsis</i>	Leaves	Iran	N.R.	viridiflorol (24 %), humulene (18 %), α -muurolene (9%)	[139]
<i>S. sclareopsis</i>	Stems	Iran	N.R.	octadecane (21%), caryophyllene oxide (8%), nonanal (6%)	[139]
<i>S. sclareopsis</i>	Flowers	Iran	N.R.	linalool (28%), hexyl 2-methyl butyrate (5%), <i>n</i> -hexyl acetate (5%)	[139]
<i>S. sharifi</i>	Aerial parts	Iran	0.2	germacrene D (30%), bicyclogermacrene (16%), β -caryophyllene (12%)	[140]
<i>S. veneris</i>	Aerial parts	Cyprus	2.0	1,8-cineole (51%), camphor (9%), camphene (6%).	[117]

(w) = wild; nd = not detected; N.R. = not reported

Table 5. Extract of the main recent publications on endemisms of Thyme and chemical variability of its essential oils

Species	Plant material	Origin	Yield (%)	Main components	Ref.
<i>T. mastichina</i>	Aerial parts	Spain	N.R.	1,8-cineole (59-68%), linalool (1-10%), β -pinene (4-5%)	[112]
<i>T. mastichina</i>	Flowers	Spain	N.R.	limonene+1,8-cineole (71%), myrcene (10%)	[148]
<i>T. mastichina</i>	Fruits	Spain	N.R.	limonene+1,8-cineole (78%), myrcene (6%)	[148]
<i>T. capitata</i>	Aerial parts	Italy	1.4-5.6	carvacrol (3-83%), <i>p</i> -cymene (4-40%), γ -terpinene (<1-10%)	[144]
<i>T. capitata</i>	Aerial parts	Spain	2.8-5.6	carvacrol (69-76%), γ -terpinene (7-8%), <i>p</i> -cymene (7%)	[147]
<i>T. capitata</i>	Flowers	Spain	N.R.	carvacrol (76%), <i>p</i> -cymene (7%), γ -terpinene (5%)	[148]
<i>T. capitata</i>	Fruits	Spain	N.R.	carvacrol (74%), <i>p</i> -cymene (9%), γ -terpinene (4%)	[148]
<i>T. capitata</i>	Aerial parts	Tunisia	1.4-2.4	carvacrol (73-76%), <i>p</i> -cymene (5-13%), γ -terpinene (1-8%)	[146]
<i>T. capitata</i>	Aerial parts	Italy	4.2-8.1	carvacrol (62-81%), <i>p</i> -cymene (5-11%), γ -terpinene (2-9%)	[145]
<i>T. caespititius</i>	Flowers	Spain	N.R.	α -terpineol (42%), <i>p</i> -cymene (11%), γ -terpinene (6%)	[147]
<i>T. caespititius</i>	Fruits	Spain	N.R.	α -terpineol (53%), <i>p</i> -cymene (10%), γ -terpinene (4%)	[147]
<i>T. zygis</i>	Flowers	Spain	N.R.	thymol (45%), <i>p</i> -cymene (23%), γ -terpinene (11%)	[147]
<i>T. zygis</i>	Fruits	Spain	N.R.	thymol (53%), <i>p</i> -cymene (30%), γ -terpinene (5%)	[147]
<i>T. spicata</i>	Aerial parts	Greece	2.4-4.8	carvacrol (67-88%), <i>p</i> -cymene (1-17%), γ -terpinene (<1-7%)	[149]
<i>T. serpyllum</i>	Aerial parts	Poland	0.2-1.0	geranyl acetate <1-23%), α -terpineol (5-22%), myrcene (1-16%)	[150]
<i>T. serrulatus</i>	Aerial parts	Ethiopia	N.R.	carvacrol (7-81%), thymol (7-66%)	[151]
<i>T. schimperi</i>	Aerial parts	Ethiopia	N.R.	carvacrol (35-72%), thymol (16-54%)	[151]
<i>T. pannonicus</i>	Aerial parts	Serbia	0.3-1.3	geranial (29-46%), neral (23-36%)	[152]

(w) = wild; nd = not detected; N.R. = not reported

Table 6. Recent studies on repellent and insecticidal activity of essential oils

Essential oil	Target insect	Evaluated parameters	Ref.
<i>L. dentata</i>	<i>Culiseta longiareolata</i>	Larvicidal activity	[213]
<i>L. dentata</i>	<i>Culex pipiens</i>	Larvicidal activity	[213]
<i>L. luisieri</i>	<i>Hyalomma lusitanicum</i>	Larvicidal activity	[214]
<i>L. angustifolia</i>	<i>Lucilla sericata</i>	Toxicity and oviposition deterrence	[215]
<i>L. angustifolia</i>	<i>Plodia interpunctella</i>	Toxicity	[216]
<i>L. angustifolia</i>	<i>Plutella xylostella</i>	Toxicity	[217]
<i>L. angustifolia</i>	<i>Sitophilus granarius</i>	Contact/fumigant toxicity, repellent, antifeedant and nutritional effects	[218]
<i>L. stoechas</i>	<i>Orgyia trigotephras</i>	Toxicity	[219]
<i>Lavandin</i>	<i>Spodoptera littoralis</i>	Biocidal	[220]
<i>Several</i>	<i>Hyalomma scupense</i>	Acaricidal activity (adults and larvae)	[221]
<i>O. vulgare</i>	<i>Tribolium confusum</i>	Repellence	[222]
<i>O. vulgare</i>	<i>Sitophilus oryzae</i>	Repellence	[222]
<i>O. onites</i>	<i>Amblyomma americanum</i>	Repellence	[223]
<i>O. onites</i>	<i>Aedes aegypti</i>	Repellence	[223]
<i>Origanum</i>	<i>Aedes albopictus</i>	Larvicidal effects and repellence	[224]
<i>O. syriacum</i>	<i>Spodoptera littoralis</i>	Insecticidal	[225]
<i>O. syriacum</i>	<i>Myzus persicae</i>	Insecticidal	[225]
<i>O. syriacum</i>	<i>Musca domestica</i>	Insecticidal	[225]
<i>O. scabrum</i>	<i>Mosquito vectors</i>	Insecticidal, ovicidal	[226]
<i>O. elongatum</i>	<i>Varroa destructor</i>	Acaricidal	[227]
<i>R. officinalis</i>	<i>Tribolium castaneum</i>	Fumigant and contact toxicity	[228]
<i>S. officinalis</i>	<i>Varroa destructor</i>	Acaricidal	[229]
<i>S. officinalis</i>	<i>Aedes aegypti</i>	Larvicidal	[230]
<i>S. plebeian</i>	<i>Aedes aegypti</i>	Larvicidal	[231]
<i>S. lavandulifolia</i>	<i>Synanthropic mites</i>	Acaricidal	[232]
<i>S. mirzayanii</i>	<i>Oligonychus afrasiaticus</i>	Acaricidal	[233]
<i>S. veneris</i>	<i>Spodoptera exigua</i>	Insecticidal	[234]
<i>S. ballotiflora</i>	<i>Spodoptera frugiperda</i>	Insectistatic and insecticidal	[235]
<i>T. satureioides</i>	<i>Varroa destructor</i>	Acaricidal	[227]
<i>T. vulgaris</i>	<i>Macrosiphum rosae</i>	Insecticidal	[236]

Table 7. Use of essential oils in animal feeding

Essential oil	Target animal	Way of administration	Evaluated parameters	Ref.
<i>L. angustifolia</i>	Broiler chickens	Drinking water	Production, blood biochemical parameters, ileal microflora	[242]
<i>L. graveolens</i>	Pigs	Food supplement	Carcass yield, sensory and acceptance of meat	[261]
<i>O. vulgare</i>	Calves	Oral administration	Neonatal diarrhea	[262]
<i>O. vulgare</i>	Lambs	Food supplement	Performance, carcass characteristics, blood parameters	[263]
<i>O. vulgare</i>	Lambs	Food supplement	Growth	[264]
<i>O. vulgare</i>	Broiler chickens	Food supplement	Growth, intestinal microflora, gut integrity	[243]
<i>O. vulgare</i>	Pigs	Food supplement	Meat quality, fatty acid composition, oxidative stability of <i>Longissimus thoracis</i> muscle	[265]
<i>O. vulgare</i>	Nile tilapia	Food supplement	Growth, behavioral, stress response	[266]
<i>O. vulgare</i>	Cows	Food supplement	Sensory properties and production levels of milk, feed consumption, methane emission	[244]
<i>O. vulgare</i>	Goats	Food supplement	Rumen fermentation, enzyme profile	[267]
<i>O. onites</i>	Rainbow trout	Food supplement	Growth, lysozyme, antioxidant activity	[268]
<i>O. syriacum</i>	Broiler chickens	Food supplement	Blood parameters	[245]
<i>R. officinalis</i>	Goats	Food supplement	Milk production	[269]
<i>R. officinalis</i>	Hens	Food supplement	Performances, egg quality, blood parameters	[270]
<i>R. officinalis</i>	Japanese quail	Food supplement	Heat stress-induced testes, structural and functional damage	[271]
<i>R. officinalis</i>	Dairy ewes, lambs	Food supplement	Digestion, colostrum production of dairy ewes, lambs mortality and growth	[272]
<i>R. officinalis</i>	Lambs	Food supplement	Meat quality	[273]
<i>R. officinalis</i>	Sheeps	Food supplement	Ruminal fermentation	[274]
<i>T. vulgaris</i>	Broiler chickens	Food supplement	Growth performances, blood parameters	[275]
<i>T. vulgaris</i>	Japanese quail	Food supplement	Performance, organs weight, intestinal morphology, serum lipids	[276]
<i>T. vulgaris</i>	Nile tilapia	Food supplement	Hemato-immunological indices, intestinal morphology, microbiota	[277]
<i>T. vulgaris</i>	Rabbits	Food supplement	Productive performance, carcass criteria and meat quality	[278]

Table 8. Selection of recent *in-vitro* studies on cytotoxicity of essential oil^a

Essential oil	Cell lines	Test	Ref.
<i>L. stoechas</i>	RD, L20B, RD	MTT	[363]
<i>L. angustifolia</i>	Caco-2	Microscopic evaluation of cell monolayers	[364]
<i>O. vulgare</i>	AGS	MTT, inverted phase- microscope	[366]
<i>O. vulgare</i>	MCF-7, HT-29	Sulforhodamine B assay	[367]
<i>O. vulgare</i>	HepG2, HEK293	MTT, inverted phase-contrast microscope	[368]
<i>O. acutidens</i>	HAT-29, HeLa	Real time cell analysis	[369]
<i>R. officinalis</i>	RDL20B, RD	MTT	[363]
<i>R. officinalis</i>	A549, H1299, MCF-7, HUVEC	MTT	[365]
<i>S. officinalis</i>	MCF.7, LNCaP, HeLa	MTT	[370]
<i>S. officinalis</i>	A549, NCI-H226	MTT	[371]
<i>S. verbenacea</i>	M14	MTT, LGD, COMET, Caspase colorimetric	[372]
<i>S. sclarea</i>	HeLa	MTT, RT-PCR	[373]
<i>S. aurea</i>	DU-145	MTT, LDH, Comet, Caspase colorimetric	[374]
<i>S. judaica</i>	DU-145	MTT, LDH, Comet, Caspase colorimetric	[374]
<i>S. viscosa</i>	DU-145	MTT, LDH, Comet, Caspase colorimetric	[374]
<i>T. capitatus</i>	MCF-7	Sulforhodamine B assay	[375]

^a MTT: tetrazolium dye MTT 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide colorimetric assay; LGD: Lowest genotoxic dose assay; RT-PCR: Reverse transcriptase-Polymerase Chain Reaction; LDH: Lactate dehydrogenase cytotoxicity assay.

Captions of figures

Figure 1.

The main genera of the Lamiaceae family.

Figure 2.

Number of medicinal plants in different plant families [7-9].

Figure 3.

Comparison between the average content of the main components of the Sicilian oregano essential oils (A) and a commercial (B) sample.

Figure 4.

Spider diagram of the sensory profile of the a Sicilian oregano and a commercial oregano sample.

Figure 5.

The photo on the left shows the typical rows of an oregano cultivation, the photo on the right shows a particular of oregano flowers.

Figure 6.

The photo on the left shows the mechanical harvesting of oregano, in the photo on the right the machinery for the preparation of the bunches of oregano

Figure 7.

The photo on the left shows alternate rows of rosemary and sage cultivation, the photo on the right shows a particular of sage inflorescens.

Figure 8.

Selection of compounds isolated from Lamiaceae species (glc=glucoside, glu=glucuronide, rhm=rhamnoside).

Figure 9.

Typical HPLC profiles of oregano (A), rosemary (B), sage (C), and thyme (D) ethyl acetate extracts.

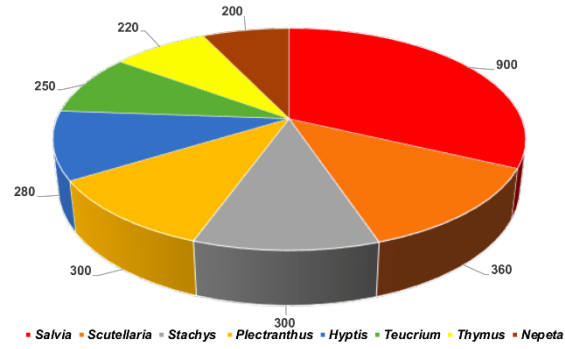


Figure 1.

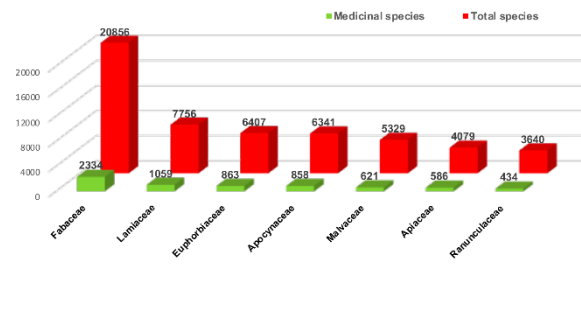


Figure 2.

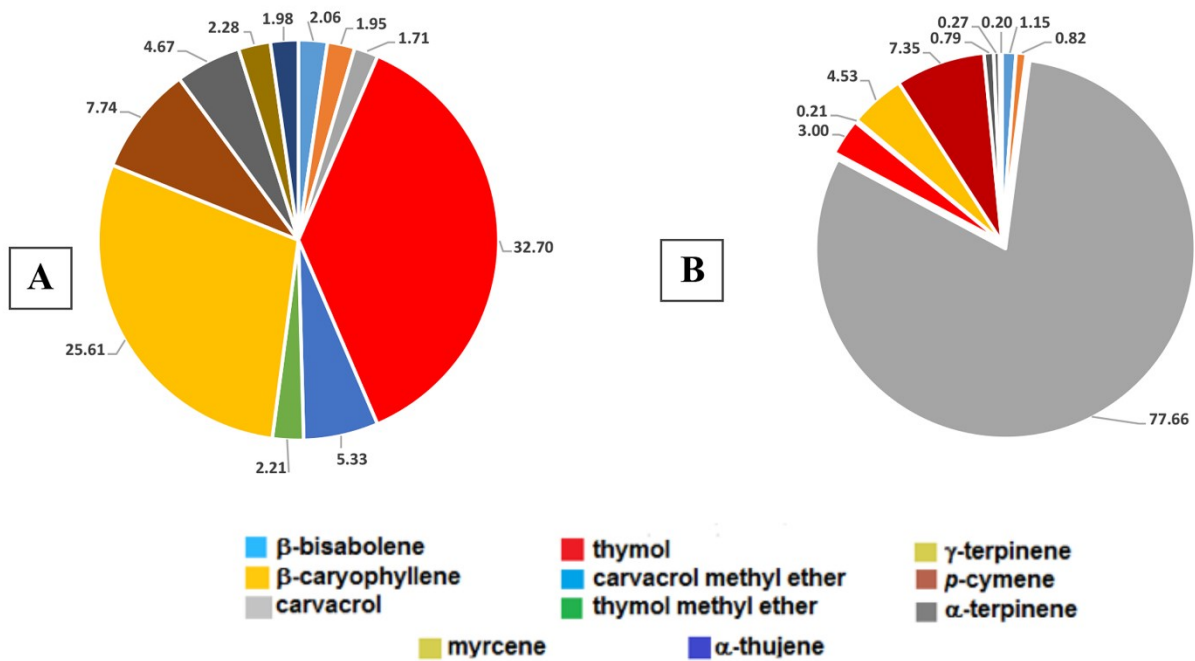


Figure 3.

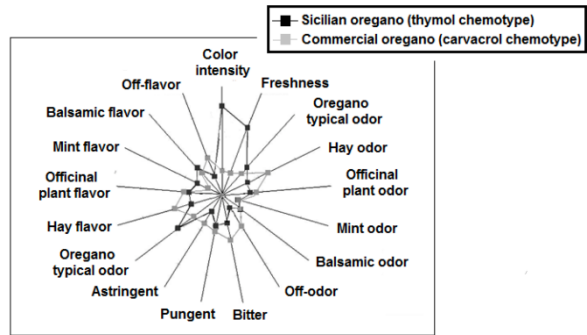


Figure 4.



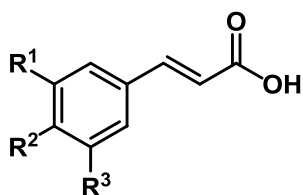
Figure 5.



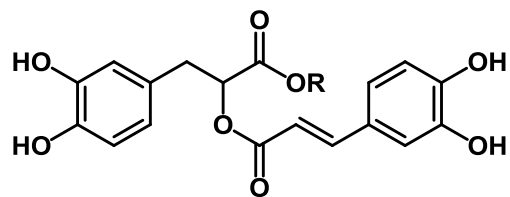
Figure 6.



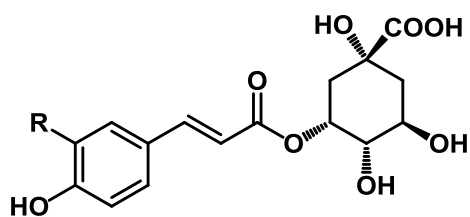
Figure 7.



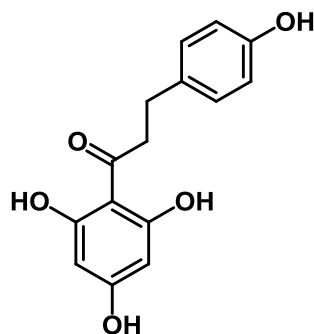
- Cinnamic acid (**39**) $R^1=R^2=R^3=H$
p-Coumaric acid (**2**) $R^1=R^3=H, R^2=OH$
 Caffeic acid (**1**) $R^1=R^2=OH, R^3=H$
 Ferulic acid (**3**) $R^1=OCH_3, R^2=OH, R^3=H$
 iso-Ferulic acid (**26**) $R^1=OH, R^2=OCH_3, R^3=H$
 Sinapic acid (**41**) $R^1=R^3=OCH_3, R^2=OH$



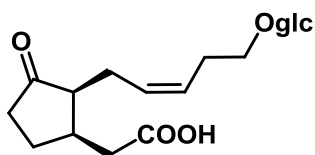
- Rosmarinic acid (**4**) ($R=H$)
 Rosmarinic acid methyl ester (**31**) ($R=CH_3$)



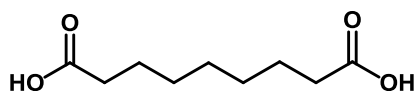
- 3-O-4-Coumaroylquinic acid (**16**) ($R=H$)
 Chlorogenic acid (**36**) ($R=OH$)



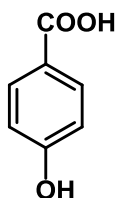
Phloridzin (**54**)



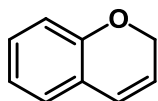
Tuberonic acid glucoside (**18**)



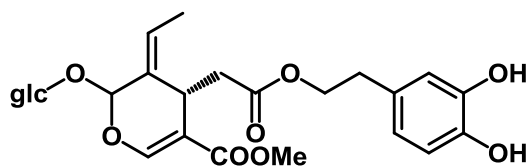
Azelaic acid (**20**)



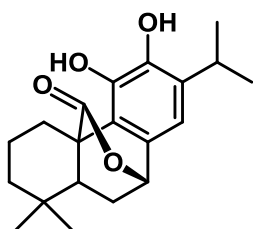
p-hydroxybenzoic acid (**51**)



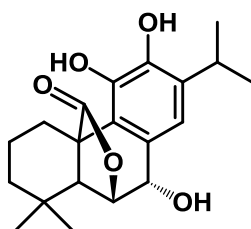
Coumarin (**38**)



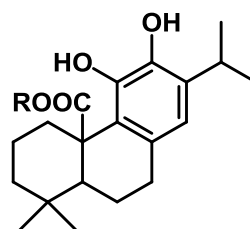
Oleuropein (**42**)



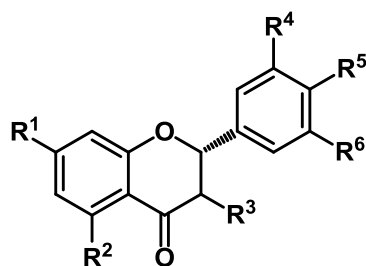
Carnosol (**43**)



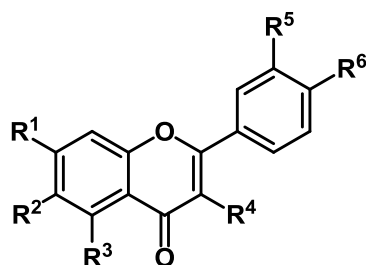
Rosmanol (**44**)



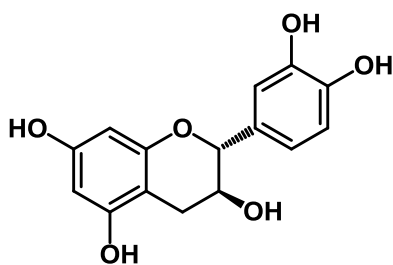
- Carnosic acid (**49**) ($R=H$)
 Methyl carnosate (**50**) ($R=CH_3$)



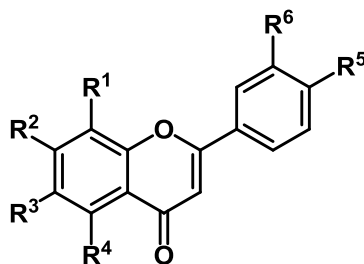
Aromadendrin (7)	$R^1=R^2=R^3=R^5=OH, R^4=R^6=H$
Dihydrorobinetin (23)	$R^1=R^3=R^4=R^5=R^6=OH, R^2=H$
Eriodictyol (8)	$R^1=R^2=R^4=R^5=OH, R^3=R^6=H$
Eriodictyol 7-O-rutinoside (6)	$R^1=Orut, R^2=R^4=R^5=OH, R^3=R^6=H$
Hesperetin (55)	$R^1=R^2=R^4=OH, R^3=R^6=H, R^5=OCH_3$
Naringenin (9)	$R^1=R^2=R^5=OH, R^3=R^4=R^6=H$
Sakuranetin (32)	$R^1=OCH_3, R^2=R^5=OH, R^3=R^4=R^6=H$
Taxifolin (58)	$R^1=R^2=R^4=R^5=OH, R^3=R^6=H$



Casticin (22)	$R^1=R^2=R^4=R^6=OCH_3, R^3=R^5=OH$
Kaempferol (27)	$R^1=R^3=R^4=R^6=OH, R^2=R^5=H$
Quercetin (30)	$R^1=R^3=R^4=R^5=R^6=OH, R^2=H$
Rutin (40)	$R^1=R^3=R^5=R^6=OH, R^2=H, R^4=Orut$

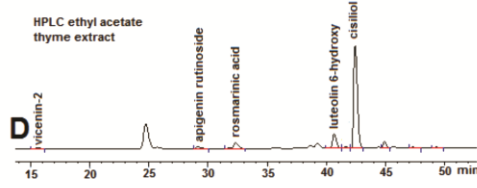
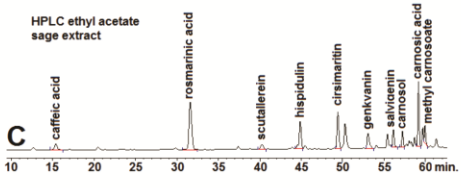
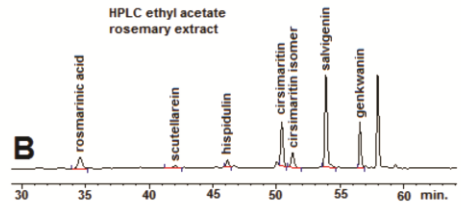
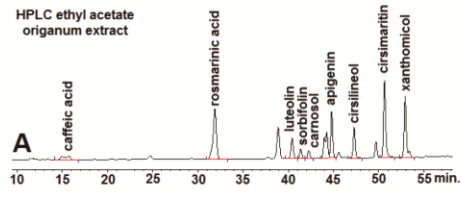


Catechin (37)



Acacetin (17)	$R^1=R^3=R^6=H, R^2=R^4=OH, R^5=OCH_3$
Apigenin (12)	$R^1=R^3=R^6=H, R^2=R^4=R^5=OH$
Apigenin 7,4'-dimethyl ether (19)	$R^1=R^3=R^6=H, R^4=OH, R^2=R^5=OCH_3$
Apigenin 7-O-glucuronide (35)	$R^1=R^3=R^6=H, R^2=Oglu, R^4=R^5=OH$
Apigenin 7-O-glucoside (53)	$R^1=R^3=R^6=H, R^2=Oglc, R^4=R^5=OH$
Apigenin 7-O-rutinoside (59)	$R^1=R^3=R^6=H, R^2=Orut, R^4=R^5=OH$
Baicalin (21)	$R^1=R^5=R^6=H, R^2=Oglu, R^3=R^4=OH$
Cirsiliol (11)	$R^1=H, R^2=R^3=OCH_3, R^4=R^5=R^6=OH$
Cirsilineol (13)	$R^1=H, R^2=R^3=R^6=OCH_3, R^4=R^5=OH$
Cirsimaritin (14)	$R^1=R^6=H, R^2=R^3=OCH_3, R^4=R^5=OH$
Chrysoeriol (62)	$R^1=R^3=H, R^2=R^4=R^5=OH, R^6=OCH_3$
Gardenin B (24)	$R^1=R^2=R^3=R^5=OCH_3, R^4=OH, R^6=H$
Gardenin B glucoside (60)	$R^1=R^2=R^3=R^5=OCH_3, R^4=Oglc, R^6=H$
Genkwanin (25)	$R^1=R^3=R^6=H, R^2=OCH_3, R^4=R^5=OH$
Luteolin (5)	$R^1=R^3=H, R^2=R^4=R^5=R^6=OH$
Luteolin 7-O-glucoside (56)	$R^1=R^3=H, R^2=Oglc, R^4=R^5=R^6=OH$
Luteoilin 7-O-glucuronide (45)	$R^1=R^3=H, R^2=Oglu, R^4=R^5=R^6=OH$
Luteolin 6-hydroxy (57)	$R^1=H, R^2=R^3=R^4=R^5=R^6=OH$
Luteolin 6-hydroxy-7-O-glucoside (46)	$R^1=H, R^2=Oglc, R^3=R^4=R^5=R^6=OH$
Luteolin 7-O-rutinoside (52)	$R^1=R^3=H, R^2=Orut, R^4=R^5=R^6=OH$
Nevadensin (28)	$R^1=R^3=R^5=OCH_3, R^2=R^4=OH, R^6=H$
Oroxylin 7-O-glucoside (29)	$R^1=R^5=R^6=H, R^2=OCH_3, R^3=R^4=OH$
Salvigenin (33)	$R^1=R^6=H, R^2=R^3=R^5=OCH_3, R^4=OH$
Scutallarin (47)	$R^1=R^6=H, R^2=Oglu, R^3=R^4=R^5=OH$
Scutallarein 7-O-glucoside (48)	$R^1=R^6=H, R^2=Oglc, R^3=R^4=R^5=OH$
Sorbifolin (10)	$R^1=R^6=H, R^2=Orhm, R^3=R^4=R^5=OH$
Vicenin-2 (34)	$R^1=R^3=C-glc; R^2=R^4=R^5=OH, R^6=H$
Thymusin (61)	$R^1=R^2=OCH_3, R^3=R^4=R^6=OH$
Xanthomicol (15)	$R^1=R^2=OCH_3, R^3=R^4=R^5=OH, R^6=H$

Figure 8. Selection of compounds isolated from Lamiaceae species (glc=glucoside, glu=glucuronide, rhm=rhamnoside).





Rosemary

Lamiaceae



Thyme



Oregano



Sage



Lavender

Graphical Abstract