Inorganic Chemistry

New Class of Half-Sandwich Ruthenium(II) Arene Complexes Bearing the Water-Soluble CAP Ligand as an in Vitro Anticancer Agent

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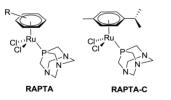
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S Supporting Information

ABSTRACT: Ruthenium(II) arene complexes of 1,4,7triaza-9-phosphatricyclo[5.3.2.1]tridecane (CAP) were obtained. Cytotoxicity studies against cancer cell lines reveal higher activity than the corresponding PTA analogues and, in comparison to the effects on noncancerous cells, the complexes are endowed with a reasonable degree of cancer cell selectivity.

S ince the discovery of cisplatin in 1965,^{1,2} interest in the application of coordination complexes to treat cancer and other diseases has continually increased.³ In particular, when resistance to cisplatin developed,^{4,5} certain classes of ruthenium complexes were found to be active,⁶ including ruthenium(III) coordination complexes⁷⁻⁹ and half-sandwich ruthenium(II) arene organometallic derivatives.¹⁰⁻¹⁴ RAPTA-type compounds (Chart 1) were identified as promising anticancer agents,^{15,16}

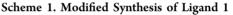
Chart 1. Prototypical RAPTA-Type $(R = H, Me, Me_6)$ and RAPTA-C Structures

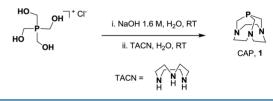


with their water solubility being given by the cage-like 1,3,5triaza-7-phosphaadamantane (PTA) ligand, binding the metal in a κ^{1} -*P* fashion.^{17–19} Several studies evaluating the antitumor activity of RAPTA-C revealed strong antiangiogenic and antimetastatic activities,^{20,21} together with a good efficacy in inhibiting tumor growth even at moderate doses,²² low general toxicity, and tolerance to low pH,^{23,24} suitable for investigation as an orally administered drug. For this class of compounds, the reactivity and stability, uptake into cancer cells, target recognition, and overall cytotoxicity and cancer cell selectivity were found to heavily depend on the choice of phosphine. Indeed, modification of the PTA ligand often led to increased cytotoxicity, but at the same time caused the cancer cell selectivity to be lost.²⁵

We have been interested in the application of Ru-PTA complexes not only as antitumor agents but also as selective

catalysts for hydrogenation reactions.²⁶⁻³³ Recently, the new ligand 1,4,7-triaza-9-phosphatricyclo[5.3.2.1]tridecane (CAP, 1) was reported (Scheme 1).³⁴





CAP has a tris(homoadamantane) cage similar to PTA and was obtained by Mannich-type condensation of 1,4,7triazacyclononane (TACN) and tris(hydroxymethyl)phosphine (THP).^{35,36} CAP has a higher cage flexibility due to the presence of two CH₂ spacers between the N atoms. Electron density distribution analysis showed that the P and N atoms of CAP exhibit comparable electron donicity, bringing about different reactivities of CAP and PTA toward protonation at N sites and alkylation at N and P atoms.^{37–39} In the search for new RAPTA-type compounds with potential anticancer activity, we decided to explore the coordination properties of CAP to ruthenium(II) arene moieties. The obtained complexes were evaluated in vitro against human ovarian (A2780 and A2780cis) cancer and human embryonic kidney (HEK) cell lines, and the preliminary results of these studies are presented here.

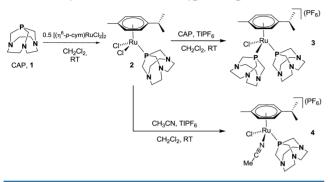
The synthesis of 1 was slightly modified compared to that in the literature. As for PTA,⁴⁰ the air-stable and commercially available tetrakis(hydroxymethyl)phosphonium chloride (THPC; 80% aqueous solution) was first treated with a NaOH solution (1.6 M), producing in situ THP [P-(CH₂OH)₃], which was reacted without isolation with an aqueous solution of TACN at room temperature (Scheme 1). After extraction of the aqueous phase with CHCl₃ and recrystallization of the crude product from hot ethanol (EtOH), 1 was obtained as a white crystalline solid in high purity. Also in our case, formation of the unidentified amorphous solid originally observed could not be avoided.³⁴ Our procedure has the advantage of using the less expensive and easier-to-handle THPC instead of air-sensitive THP and a

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facile workup consisting of a simple extraction and recrystallization.

The RAPTA-C analogue κ^{1} -*P*-[(η^{6} -*p*-cymene)RuCl₂(CAP)] (2, RACAP-C) was obtained by reacting [(η^{6} -*p*-cymene)-RuCl₂]₂ with 2 equiv of CAP in CH₂Cl₂ at room temperature,





affording 2 in 87% yield after purification (Scheme 2). The ³¹P{¹H} NMR spectrum of **2** in CDCl₃ shows a singlet at 52.83 ppm, a rather different chemical shift than RAPTA-C (-36.63 ppm)¹⁹ and in line with the gold(I) CAP derivative (CAP- H_2 ₃Au⁷⁺₄ (50.99 ppm).³⁴ The substantial ³¹P{¹H} NMR downfield shift is not unexpected because of the different Pdonor properties of 1.37 In the corresponding ¹H NMR spectrum, the cage protons of coordinated CAP give a multiplet in the range 3.07–2.92 ppm for $N(CH_2)_2N$ and a doublet centered at 3.70 ppm (${}^{2}J_{HP} = 7.2$ Hz) due to PCH₂N. This assignment was verified by ¹H{³¹P} NMR spectroscopy, with the doublet turning into a singlet upon P decoupling. The aromatic protons of the *p*-cymene ring give a broad singlet at 5.48 ppm, similar to the signal at 5.46 ppm reported for [$(\eta^6$ -pcymene)RuCl₂(PTA)].¹⁹ In the ¹³C{¹H} NMR spectrum of 2, the η^6 -coordinated arene ring affords two doublets for the aromatic CH C atoms at 88.55 and 83.97 ppm with ${}^{2}J_{CP} = 2.5$ and 3.4 Hz, respectively. The proposed structure of 2 was also confirmed by electrospray ionization mass spectrometry (ESI-MS), with a peak envelope centered at m/z 470.2 corresponding to the cation $[(\eta^{6}-p-cymene)RuCl(CAP)]^{+}$, i.e., the expected $[M - Cl]^+$ ion.

The ruthenium(II) complex κ^1 -P-[(η^6 -p-cymene)RuCl- $(CAP)_2$]PF₆ (3) was obtained in ca. 77% yield by reacting 2 with 1 equiv of CAP and a stoichiometric amount of TlPF₆ (Scheme 2). The ³¹P{¹H} NMR spectrum of 3 in acetone- d_6 contains a singlet at 51.60 ppm, corresponding to the two equivalent phosphines and a septet at -144.26 ppm ($J_{\rm PF}$ = 707.5 Hz) attributable to the PF_6^- counterion (confirmed by the IR peak at 841.95 cm⁻¹, KBr). In the ¹H NMR spectrum, two doublets at 6.55 and 6.26 ppm ($J_{\rm HH}$ = 6.4 Hz) corresponding to the aromatic protons of the arene ligand were observed, and the protons of PCH₂N give two sets of multiplets in the range 3.85-3.67 ppm, which afford a defined AB system (J_{HAHB} = 16.0 Hz) in the ¹H{³¹P} NMR spectrum (see the Supporting Information). In line with the data obtained for the PTA analogue,⁴¹ the ESI-MS spectrum of 3 contains a peak at m/z 669.3 consistent with the ion $[(\eta^6-p$ cymene)RuCl(CAP)₂]⁺ and a peak of lower relative intensity at m/z 470.2 corresponding to $[(\eta^{\circ}-p-\text{cymene})\text{RuCl}(\text{CAP})]^+$, due to the loss of one phosphine ligand.

The complex κ^1 -P-[(η^6 -p-cymene)RuCl(MeCN)(CAP)]PF₆ (4) was obtained from 2 by chloride abstraction with $TlPF_6$ and ligand substitution with CH_3CN (Scheme 2). The ${}^{31}P{}^{1}H$ NMR spectrum of 4 exhibits a singlet at 48.17 ppm corresponding to the P atom in the CAP ligand and a septet at -144.27 ppm (J_{PF} = 707.8 Hz) corresponding to the PF₆⁻ ion (IR peak at 839.16 cm⁻¹). The presence of coordinated acetonitrile in 4 was confirmed by the singlet at 2.52 ppm in the ¹H NMR spectrum and by the two singlets at 128.55 and 3.91 ppm in the ${}^{13}C{}^{1}H$ NMR spectrum, with the the former attributed to $C \equiv N$ and the latter to CH_3CN .⁴² The ¹H NMR spectrum of 4 showed two well-distinguished doublets centered at 1.26 and 1.25 ppm (${}^{3}J_{\rm HH}$ = 6.8 Hz) for the arene CH₃ isopropyl groups upon loss of symmetry in the piano-stool geometry. The solubilities of 2-4 in water were determined at 20 °C. Complex 2 was found to be much less water-soluble than RAPTA-C, with $S(H_2O)_{20 \ ^\circ C}$ = 0.86 versus 10 g/L. The ionic complexes 3 and 4 were slightly more soluble than 2, with values of 1.2 and 1.1 g/L, respectively.

The thermal stability of **2** in D_2O was determined by variable-temperature NMR experiments. No changes were observed in the corresponding ${}^{31}P{}^{1}H$ NMR spectra (singlet at 57.48 ppm) in the temperature range 25–60 °C for a total of 30 h of experiment.

The stabilities of complexes 2-4 under pseudopharmacological conditions were also determined in aqueous NaCl/D₂O (100 mM) at 37 $^{\circ}\mathrm{C}$ for 72 h, monitoring every 24 h by $^{1}\mathrm{H}$ and ³¹P{¹H} NMR (see the Supporting Information). Complex 2 was stable under these conditions; i.e., the NMR spectra remained unchanged after 72 h, showing a ³¹P{¹H} NMR singlet at 57.31 ppm. Complex 3 showed after 7 h the presence of singlets at 56.77 ppm (ca. 47% by integration), due to the original complex, and at 57.31 ppm (ca. 30%), due to the formation of 2 upon CAP-Cl⁻ exchange. Two less intense singlets were also observed at 58.46 ppm (10%) and 54.44 ppm (13%). After 48 h, the 3:2 ratio reached ca. 1:1, and it was maintained until the end of the experiment. Similarly, complex 4, characterized by a ${}^{31}P{}^{1}H$ NMR singlet at 54.30 ppm, immediately underwent MeCN-Cl⁻ exchange to form complex 2 (ca. 60%). The ratio between 2 and 4 remained constant during the following 72 h (Figures S16–S21 in the Supporting Information).

Suitable crystals for single-crystal X-ray diffraction of 2 and 3 were obtained from the slow diffusion of dry EtOH in a CH₂Cl₂ solution of each complex. The molecular structure of 2 (Figure 1) consists of a Ru^{II} center coordinated by a η^6 -*p*-cymene unit, two Cl atoms, and a κ^1 -*P* CAP ligand. The CAP ligand does not experience significant changes of the [333] conformation observed in the free ligand.⁴³ Accordingly, the intramolecular P…N (d_{PN}) and N…N (d_{NN}) distances of CAP in 2 range from 2.824 to 2.808 Å and from 3.028 to 3.064 Å, respectively (comparable to that found for uncoordinated CAP, i.e., an average $d_{\rm PN}$ of 2.886 Å and an average $d_{\rm NN}$ of 3.1132 Å).³⁷ The Ru–P bond length in 2 is significantly longer compared to that observed in the analogous PTA structure [i.e., 2.3180(5) vs 2.296(2) and 2.298(2) Å].¹⁹ The same applies to the Ru–C(pcymene) bond lengths [i.e., 2.198(2) and 2.275(2) Å in 2 versus 2.179(9) and 2.210(10) Å in the PTA analogue].¹⁹

The replacement of one Cl⁻ in **2** by a CAP ligand, to give the monocationic ruthenium species **3** (Figure 2), leads to an elongation of the Ru–P [i.e., 2.3180(5) Å (**2**) vs 2.3254(7) and 2.3353(7) Å (**3**·2MeOH·H₂O)] and Ru–C(*p*-cymene) [i.e., 1.709 Å (**2**) vs 1.768 Å (**3**·2MeOH·H₂O)] bonds compared to

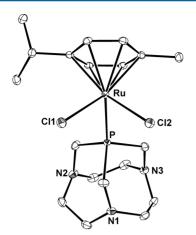


Figure 1. Molecular structure of **2**. Thermal ellipsoids are presented at a 30% probability level, and H atoms are omitted for clarity. Selected bond distances (Å) and angles (deg): Ru-Cl1 = 2.4184(5), Ru-Cl2 = 2.4205(5), Ru-P = 2.3180(5), Ru-centroid (C1–C6) = 1.709; Cl1–Ru–Cl2 = 87.601(19), Cl1–Ru–P = 86.937 (19), Cl2–Ru–P = 83.775(18).

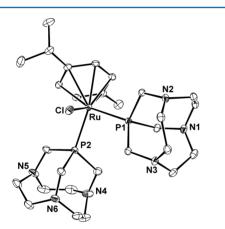


Figure 2. Molecular structure of $3.2 \text{MeOH} \cdot \text{H}_2\text{O}$. Thermal ellipsoids are presented at a 30% probability level, and H atoms, counteranions, and solvent molecules are omitted for clarity. Selected bond distances (Å) and angles (deg): Ru–Cl = 2.4026(7), Ru–P1 = 2.3254(7), Ru–P2 = 2.3353(7), Ru–centroid (C1–C6) = 1.768; Cl–Ru–P1 = 83.59(2), Cl–Ru–P2 = 87.90(3), P1–Ru–P2 = 93.37(3).

2, whereas the Ru–Cl bond shortens from 2.4205(5) Å (2) to 2.4026(7) Å (3·2MeOH·H₂O). The overall conformation of the coordinated CAP is essentially identical with that found in 2. The significantly different Ru–P bond lengths found for 2 and 3·2MeOH·H₂O correlate with the slightly higher positive ³¹P coordination shift found for 2 in solution [i.e., 52.83 ppm, d_{RuP} = 2.3180(5) Å for 2, vs 51.60 ppm, d_{RuP} = 2.3254(7) and 2.3353(7) Å for 3·2MeOH·H₂O]. The stronger Ru–*p*-cymene interaction in 2 compared to 3, observed in the solid state, is thus maintained in solution. Accordingly, the aromatic ¹H NMR signals assigned to *p*-cymene in 2 are shifted upfield compared to 3. In addition, the ¹³C{¹H} NMR signals of the aromatic C atoms of *p*-cymene in 2 have small ²*J*_{CP} values of 2.5 and 3.4 Hz, whereas in 3, only ¹³C{¹H} NMR singlets are observed.

Compounds 1–4 were tested for their cytotoxicity to human ovarian A2780 carcinoma cells and the A2780cisR variant with acquired resistance to cisplatin as well as against noncancerous HEK293 cells, using MTT assay (Table 1). All complexes

Table 1. Cytotoxicity (IC ₅₀ , μ M, 72 h) of 1–4 to A2780 and
A2780cisR Cancer Cells and Non-cancerous HEK293 Cells

compound	A2780	A2780cisR	HEK293	
1	>200	>200	>200	
2	55.3 ± 18.6	108 ± 10	102 ± 26	
3	48.1 ± 2.2	99.2 ± 15.9	80.7 ± 12.4	
4	65.2 ± 18.0	70.6 ± 3.1	163 ± 46	
RAPTA-C ^a	230	270	>1000	
^{<i>a</i>} Values taken for comparison from ref 45.				

induced a dose- and cell-dependent decrease in the cell viability, with 1 exerting only minor cytotoxic effects (not reaching the IC_{50} value in the tested concentration range). The similar IC_{50} values of the complexes are likely due to the same species formed by 3 and 4 in aqueous solution after ligand exchange reactions leading to 2. Compared to RAPTA-C, the compounds are considerably more cytotoxic toward all cell lines. However, only complex 4 shows essentially equal cytotoxicity toward both the A2780 and A2780cisR cells. In comparison to complexes with other PTA-type ligands, complexes 2–4 maintain their cancer cell selectivity by a 2-fold stronger growth inhibition effect on A2780 cancer cells compared to noncancerous HEK293 cells. Usually modification of the PTA ligand has often led to more cytotoxic complexes but at the same time reduces the cancer cell selectivity.^{30,44}

In conclusion, novel ruthenium(II) arene half-sandwich complexes bearing the cage-type CAP were prepared and characterized. Compared to RAPTA-C, the direct CAP analogue **2** is more cytotoxic to cancer cells and exhibits a reasonable degree of cancer cell selectivity. Other PTA-type ligands previously studied have not maintained the desirable cancer cell selectivity exhibited by the CAP derivatives.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.inorg-chem.7b00915.

General methods and materials, synthetic procedures, NMR and MS spectra, tables for X-ray crystal structure determination, stability tests, and details of in vitro cytotoxicity tests (PDF)

X-ray crystallographic data in CIF format (CIF) X-ray crystallographic data in CIF format (CIF)

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Author Contributions

All authors have given approval to the final version of the manuscript.

Notes

The authors declare no competing financial interest.

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