

Effect of deuteration degree of amphetamine on isotope effect in HPLC was studied

Effect of structure of analyte, mobile phase and chiral selector on isotope effect in HPLC was studied

Possible mechanisms of separation of isotopologues and enantiomers were studied.

1 **Separation of isotopologues of amphetamine with various degree of**  
2 **deuteration on achiral and polysaccharide-based chiral columns in high-**  
3 **performance liquid chromatography**

4

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21 **Abstract**

22 Hydrogen/deuterium (H/D) isotope effects are not unusual in chromatography and such  
23 phenomena have been observed in both gas- and liquid-phase separations. Despite the  
24 numerous reports on this topic, the understanding of mechanisms and the underlying  
25 noncovalent interactions at play remains rather challenging. In our recent study, we reported  
26 baseline separation of isotopologues of some amphetamine (AMP) derivatives on achiral and  
27 polysaccharide-based chiral columns, as well as some correlations between the degree of  
28 separation of enantiomers and isotopologues on (the same) polysaccharide-based chiral  
29 column(s). Following our previous findings on isotope effects in high-performance liquid  
30 chromatography, we report herein a comparative study on the isotope effects observed with  
31 AMP and methamphetamine (MET). The impact of some pivotal factors such as the number  
32 of deuterium atoms part of AMP isotopologues, the structure of its isotopomers, the chemical  
33 structure of the achiral and chiral stationary phases used in this study, and the use of  
34 methanol- vs acetonitrile-containing mobile phases on the isotope effects was examined and  
35 discussed. Quantitative correlations between the observed isotope effects and the  
36 enantioselectivity of the chiral columns used are also shortly discussed. Furthermore,  
37 considering the chromatographic results as benchmark experimental data, we attempted to  
38 elucidate the molecular bases of the observed phenomena using quantum mechanics  
39 calculations.

40

41 **Keywords:** Effect of deuterium atoms / Isotope effect in HPLC / Polysaccharide-based chiral  
42 columns / Separation of enantioisotopologues

43

## 44 1. Introduction

45 Isotopically labelled compounds have attracted increasing interest in the  
46 pharmaceutical industry in the last decade [1-5]. In particular, more and more attention is paid  
47 to the isotope kinetic effect [6] in biological (living) systems and among them in humans, in  
48 the sense that undeuterated and deuterated biologically active compounds may have different  
49 pharmacokinetic, toxicokinetic, and most likely even pharmacodynamic properties. Such  
50 differences led to the approval of some deuterated chemical species as drugs for clinical use  
51 by regulatory bodies since 2017 [1-5]. Contrary to this, in bioanalysis it is believed that  
52 isotope effects do not play any significant role or don't even exist and therefore, isotopically  
53 labelled (primarily deuterated) analogues of target analytes are considered to be optimal  
54 internal standards in bioanalysis with mass-spectrometric (MS) detection [7,8]. Thus, remarks  
55 like this "*Ideally, a stable-isotope labeled internal standard is preferred whenever possible, as*  
56 *it has exactly the same structure as the analyte and co-elutes with it.*" [7] are quite common in  
57 the scientific literature. This belief translates into the following practical consequences:  
58 nonlabelled and isotopically labelled compounds extract from biological matrixes most likely  
59 to the same degree, have the same solubility in specific solvents, have the same pK values, the  
60 same retention in given chromatographic systems (column + mobile phase), etc. Such  
61 assumptions mean that isotopologues cannot be chromatographically separated. However,  
62 when mass-spectrometric detection is used, isotopologues can be differentiated and  
63 selectively detected and quantified based on their different molecular mass. The coelution of  
64 the undeuterated and deuterated forms of a given analyte is generally considered  
65 advantageous in LCMS analysis with the expectation that both species experience the same  
66 degree of ion enhancement or suppression in the ion source [7,8].

67 Over the past decades, significant isotope effects have been observed in gas  
68 chromatography [9-16], in various modes of liquid chromatography, such as ion-exchange  
69 [17], normal phase [18], reversed-phase [19-23] and chiral [24-26] chromatography, as well as

70 in electrokinetic chromatography [27,28]. Two types of isotope effects are known in  
71 chromatography, i.e. the “normal” and the “inverse” isotope effect. The former refers to cases  
72 when the heavier deuterated isotopologues retain longer compared to lighter protiated  
73 counterparts, whereas the “inverse” isotope effect is observed when lighter isotopologues  
74 retain longer compared to heavier counterparts. The presence of either isotope effect is  
75 contrary to the expectations of most analysts, and a challenge in bioanalysis [23]. On the other  
76 hand, the isotope dilution method is based on the presence of such an isotope effect and  
77 enables using isotopically labelled internal standards without the use of MS detection [13,19].  
78 In addition, Rudaz and co-authors have demonstrated that for quantification with a mass  
79 spectrometric detector it may be advantageous that nonlabelled and isotopically labelled  
80 internal standard elute with different retention factors. This means that the presence of the  
81 isotope effect favors adequate quantification [24].

82 In a recent study, we observed that the isotope effect was tunable and dependent on  
83 mobile phase composition in high-performance liquid chromatography (HPLC) for several  
84 AMP derivatives [26]. Under the studied conditions, a stronger isotope effect was observed in  
85 acetonitrile-containing mobile phases compared to methanol-containing ones with both chiral  
86 and achiral columns and both “normal” and “inverse” isotope effects were observed. Whereas  
87 the former was favored in polar organic solvents, increasing the content of the aqueous  
88 component in the reversed-phase mobile phase favored an “inverse” isotope effect.  
89 Preliminary quantum mechanics calculations supported the hypothesis that polar, hydrogen  
90 bonding-type noncovalent interactions are involved in the “normal” isotope effect, while  
91 apolar, hydrophobic-type interactions underlie the “inverse” isotope effect.

92 Following our previous study, we describe herein a study on the isotope effects  
93 observed on AMP and MET isotopologues. In this frame, the impact of the number of  
94 deuterium atoms part of AMP isotopologues, the structure of isotopomers, the chemical

95 structure of the achiral and chiral stationary phases used in this study, and the use of  
96 methanol- vs. acetonitrile-containing mobile phases on the isotope effects are reported and  
97 examined. Quantitative correlations between the isotope effect and the enantioselectivity of  
98 the chiral columns used are also shortly discussed. With the aim of elucidating the molecular  
99 bases of the observed phenomena, quantum mechanics calculations were performed focusing  
100 on the vibrational degree of freedom calculated for low-energy conformers of both AMP and  
101 MET isotopologues as well as related zero-point vibrational energies as descriptors useful to  
102 differentiate computationally isotopologues and isotopomers.

103

## 104 **2. Experimental**

### 105 *2.1. Materials*

106 The chiral test compounds amphetamine (AMP), amphetamine- $d_{5(\text{ring})}$  (AMP- $d_{5(\text{ring})}$ ),  
107 methamphetamine (MET) and methamphetamine- $d_5$  (MET- $d_5$ ) were commercially available  
108 from Cerilliant (Round Rock, TX, USA). S-(+)-amphetamine (S-(+)-AMP), amphetamine-  
109  $d_{5(\text{side chain})}$  (AMP- $d_{5(\text{side chain})}$ ), amphetamine- $d_6$  (AMP- $d_6$ ), amphetamine- $d_8$  (AMP- $d_8$ ) and  
110 amphetamine- $d_{11}$  (AMP- $d_{11}$ ) were purchased from Sigma Aldrich (Milan, Italy). Standards  
111 were stored at  $-20^\circ\text{C}$  until used in analysis. The structures of the studied analytes are shown  
112 in Fig 1. HPLC-grade methanol, acetonitrile and water were supplied by Carlo Erba  
113 (Cornaredo, Italy). Ammonium hydroxide (25% w/w aqueous solution) and ammonium  
114 bicarbonate (98.5% purity) were purchased from Honeywell Fluka™ (Morristown, NJ, USA).  
115 The chiral columns Lux-AMP, Lux i-Amylose-3 (with amylose tris(3-chloro-5-  
116 methylphenylcarbamate) as a chiral selector) and Lux Cellulose-3 (based on cellulose tris(4-  
117 methylbenzoate) as a chiral selector), as well as all achiral columns such as Kinetex 2.6  $\mu\text{m}$   
118 Phenyl-Hexyl, Kinetex 2.6  $\mu\text{m}$  Biphenyl and Luna Omega 1.6  $\mu\text{m}$  Polar C18 were provided  
119 by Phenomenex Inc. (Torrance, CA, USA). Lux Cellulose-3 and Lux i-Amylose-3 both

120 packed with the particles of 5  $\mu\text{m}$  nominal size and the Lux AMP column packed with the  
121 particles of 3  $\mu\text{m}$  nominal size all were of 250 x 4.6 mm dimensions. All 3 achiral columns  
122 were of 100 x 2.1 mm dimensions.

123

## 124 *2.2. High-performance liquid chromatography tandem mass spectrometry (HPLC-MS/MS)* 125 *analysis*

126 A HPLC 1290 Infinity II (Agilent Technologies Italia S.p.a., Milan, Italy) instrument  
127 coupled with a mass spectrometer (6470A Triple Quadrupole LC-MS) equipped with an  
128 electrospray ionization source (ESI) operating in both positive and negative mode was used.  
129 Data were acquired with MassHunter® Workstation Qualitative Analysis 10.0 Software  
130 (Agilent). Analysis of all compounds were performed on three chiral and three achiral  
131 columns (for their characteristics see above). Isocratic elution mode was adopted in all  
132 experiments. In case of Lux AMP column which is unique columns with pH stability in the  
133 range 1.0-11.5, methanol with 0.1% ammonium hydroxide was initially used as mobile phase,  
134 while later the mobile phase was composed of methanol and 5 mM ammonium bicarbonate  
135 (pH=11.0, adjusted with ammonium hydroxide) in water in the ratio 95/5 (v/v). Also, the  
136 content of methanol was decreased in 5% steps down to 20/80 (v/v) ratio of methanol to 5  
137 mM ammonium bicarbonate in water. The same procedure was used with acetonitrile as  
138 mobile phase organic component. In case of the other five columns (two chiral and three  
139 achiral) the same approach was used, but without adjusting the pH of 5 mM ammonium  
140 bicarbonate solution which was used at its native pH=7.7. For the chiral columns (of 4.6 mm  
141 i.d.) 1 ml/min mobile phase flow rate was used.

142 In case of the Kinetex 2.6  $\mu\text{m}$  Phenyl-Hexyl column, 0.5 ml/min flow rate was used  
143 with methanol, while 0.4 ml/min with acetonitrile containing mobile phases. The Kinetex 2.6

144  $\mu\text{m}$  Biphenyl column was operated at 0.4 ml/min mobile phase flow rate. The Luna Omega  
145 1.6  $\mu\text{m}$  Polar C18 column was operated at 0.4 ml/min flow rate with acetonitrile in  
146 combination with 5 mM ammonium bicarbonate in water as mobile phase. The same column  
147 operated with methanol in combination with 5 mM ammonium bicarbonate in water  
148 experienced excessive backpressure. For this reason, all experiments using such mobile phase  
149 were executed at 0.2 ml/min mobile phase flow rate. Autosampler and column oven  
150 temperatures were set to 10° C and 25° C, respectively. The mass spectrometer was operated  
151 in scheduled multiple reaction monitoring (MRM) mode for the analyte and internal standard,  
152 with two transitions each (Table 1). Scan speed (dwell time) was set to 0.023 sec. ESI  
153 conditions were optimized as follows: capillary voltage 3500 V, source temperature 300° C,  
154 cone gas flow rate 10 L/min, desolvation gas flow rate 12 L/min.

155

### 156 2.3. Computations

157 The 3D structures of AMP and MET were prepared by using the build function, and  
158 model kits and tools provided by Spartan' 10 Version 1.1.0 (Wavefunction Inc., Irvine, CA,  
159 USA) [29] for building and editing organic molecules. On this basis, the structures of these  
160 molecules were generated, and their refinement was performed by a MMFF procedure. Then,  
161 each structure was submitted to a conformational search through a systematic algorithm by  
162 using the MMFF force field, spanning all shapes accessible to the molecule without regard to  
163 energy. After elimination of three high-energy conformers (Boltzmann Distribution %  $\leq 0.1$ ), a  
164 set of 15 energetically accessible conformers was selected in both cases. For each conformer,  
165 geometry optimization was performed in gas phase at density functional theory (DFT) level  
166 with the B3LYP functional and the 6-31G(d) basis set. For each compound, two low-energy  
167 conformers were selected and their geometry was optimized with Gaussian 16W (Wallingford,  
168 CT, USA) [30] using tight convergence criteria, D3 version of Grimme's dispersion with the



169 original D3 damping function [31], and the solvation model based on density (SMD)  
170 (acetonitrile, methanol, and water) variation of IEFPCM (integral equation formalism for  
171 polarizable continuum model) of Truhlar and co-workers [32]. Computation of electrostatic  
172 potential ( $V$ ) values mapped on electron density isosurfaces ( $V_S$ ) was performed. Search for the  
173 exact location of  $V_{S,max}$  was extracted from the .wfn files through the Multiwfn code [33] and  
174 through its module enabling quantitative analyses of molecular surfaces (isovalue 0.001 au)  
175 [34]. The .wfn files for AMP and MET, and the vibrational wavenumbers for AMP and MET  
176 isotopologues were calculated by using Gaussian 16W (Wallingford, CT, USA) [30]  
177 [DFT/B3LYP/6-31G(d)]. The energy of the optimized structures of the complexes of MET with  
178 benzene, 1,3-dimethylbenzene, and 1-chloro-3-methylbenzene were calculated at DFT level  
179 with the B3LYP functional, the 6-31+G(d,p) as basis set, and the D3 version of Grimme's  
180 dispersion with Becke-Johnson damping [35].

181

### 182 **3. Results and Discussion**

183 Separation and enantioseparation of AMP and MET isotopologues, i.e. AMP, AMP-  
184  $d_{5(\text{side chain})}$ , AMP- $d_{5(\text{ring})}$ , AMP- $d_6$ , AMP- $d_8$ , AMP- $d_{11}$ , MET and MET- $d_5$ , were systematically  
185 explored by using the Kinetex Biphenyl (Supporting Information, Tables S1-S3 and Figs. S1-  
186 S3) and Phenyl-Hexyl achiral columns, and Lux AMP (Tables S4-S6, Figs. S4 and S5), Lux  
187 Cellulose-3 and Lux *i*-Amylose-3 (Tables S7-S9, Figs. S6 and S7), as chiral columns, at  
188 different contents of water in aqueous MeOH and ACN mixtures used as mobile phases.

189

#### 190 *3.1. Effect of achiral and chiral column stationary phase on isotope effect*

191 The isotope effect observed with the Biphenyl column in methanol was weak. It was  
192 positive (i.e. the heavier isotopologues retained longer compared to the lighter ones) in

193 methanol containing 0.1% (v/v) ammonium hydroxide and methanol containing a low amount  
194 of aqueous phase and turned negative (i.e. lighter isotopologues retained longer compared to  
195 the heavier ones) with increasing amount of aqueous component in the mobile phase. The  
196 same trend was observed for MET as for AMP and its isotopologues (Supporting Information,  
197 Table S3 and Fig. S3). The effect observed with the Phenyl-Hexyl column was very similar to  
198 the one observed on the Biphenyl column.

199           On the chiral Lux AMP column AMP and its isotopologues were retained very long  
200 (for over 40 minutes) and peaks were severely distorted in methanol without any additive. In  
201 this mobile phase, the positive isotope effect was observed. However, in contrast to the  
202 abovementioned achiral columns, there was no positive isotope effect observed, not even in  
203 methanol containing 0.1% ammonium hydroxide. In fact, there was a very weak, almost  
204 negligible, negative isotope effect in methanol with 0.1% ammonium hydroxide [ $\Delta RT$  (AMP-  
205 AMP-d<sub>5</sub>(side chain) = 0.001 min)] that increased with increasing content of the aqueous  
206 component in the mobile phase (Figs. 2a-c). The separation of isotopologues and enantiomers  
207 correlated very well with each other (Figs. 2a-c). Specifically, with increasing aqueous  
208 content in the mobile phase both, the extent of the negative isotope effect, as well as the  
209 separation selectivity of AMP-enantiomers increased significantly (Fig. 2). The same  
210 observation was made for MET under the same experimental conditions (Fig. 3).

211           On the chiral i-Amylose-3 column there was no measurable isotope effect in methanol  
212 containing 0.1% ammonium hydroxide but with addition of 5% (v/v) aqueous buffer a  
213 significant positive isotope effect appeared (Fig. 4) which is different from the observation  
214 made with the Lux AMP column. With increasing content of the aqueous mobile phase, the  
215 positive isotope effect reversed to negative and at a high content of aqueous component also  
216 enantioseparation appeared (This is another example of a correlation between the negative  
217 isotope effect and enantioseparation) (Fig. 4). With MET, positive isotope effect was observed

218 on i-Amylose-3 while no enantioseparation in any methanol-containing mobile phases (Table  
219 S9).

220 A rather weak (mostly negative) isotope effect was observed on the Lux Cellulose-3  
221 column for both AMPs and MET in methanol-containing mobile phases, while a weak  
222 positive isotope effect was observed in ACN-containing mobile phases (Data not shown). The  
223 enantiomers of AMP and MET were not separated on this column under the conditions  
224 studied.

225 From these experimental results some remarks emerged:

- 226 1. The negative isotopic effect was favoured by mobile phases containing higher  
227 percentages of water. In all cases, retention times increased significantly as the  
228 water content in the mobile phase increased (typical reversed-phase behaviour  
229 although at lower content of water (up to 20% mostly) HILIC-like retention was  
230 observed).
- 231 2. With the two amylose-based columns, i.e. Lux AMP and i-Amylose-3, the type and  
232 extent of the isotope effect as well as of enantioseparation could be affected by the  
233 stereoelectronic features of the pendant groups.

234

235 3.2. *Isotope effect in relation to methanol vs. acetonitrile and amphetamines vs.*

236 *methamphetamines*

237 As mentioned in subsection 3.1 only a weak isotope effect was observed for both,  
238 AMP and MET in methanol on the Biphenyl column while a significant difference was  
239 observed on the same column in ACN between these compounds. In particular, only a  
240 marginal isotope effect was observed for AMPs in ACN containing 0.1% ammonium  
241 hydroxide ( $\Delta$ RT = 0.001 min), while a quite strong positive isotope effect was observed for

242 MET in the same separation system (Fig. 5). The observation on the Phenyl-Hexyl column in  
243 ACN vs. methanol was very similar to that on the Biphenyl column (Data not shown).

244 On the chiral Lux AMP column, good separation of AMP enantiomers and only very  
245 weak (almost undetectable) positive isotope effect was observed in ACN containing 0.1 %  
246 ammonium hydroxide. With the addition of an initial amount of the aqueous component (see  
247 experimental section) HILIC behavior was observed as reported in our multiple studies  
248 leading to a reduction in retention factors and enantioseparation [26,36-39]. Already at 5%  
249 (v/v) content of the aqueous component in the mobile phase the isotope effect turned negative  
250 and gradually increased in correlation with enantioseparation (Table S5 and Figure S5). MET  
251 showed a significant difference from AMP with positive isotope effect observed in ACN and  
252 lower content of aqueous buffer. This positive isotope effect disappeared and then flipped to  
253 negative isotope effect with increasing content of the aqueous component in the mobile phase  
254 without achieving separation of enantiomers (Supporting information, Table S6).

255 On the Lux i-Amylose-3 column there was no measurable isotope effect for AMP, and  
256 its enantiomers were not separated in ACN modified with 0.1% (v/v) ammonium hydroxide,  
257 while for MET under the same experimental conditions there was a quite strong positive  
258 isotope effect, as well as MET enantiomers were significantly separated from each other (Fig.  
259 6).

260 The experimental results obtained for the separation and enantioseparation performed  
261 with acetonitrile-containing mobile phases showed that under these conditions the positive  
262 effect tended to be favored, while the negative effect could be observed with mobile phases  
263 containing higher percentages of water. In this regard, it is worth mentioning that acetonitrile  
264 favors and disfavors hydrogen-bonding (HB) and hydrophobic interactions between the chiral  
265 selector and analyte species, respectively, in contrast to methanol which manifests the  
266 opposite effects.

267

268 3.3. *Correlations between the extent of the isotope effect and the number and location of*  
269 *deuterium atoms in the molecule*

270 There are a few reports in the literature on the effect of the number of deuterium atoms  
271 part of the isotopologue on the extent of the isotope effect [20,25]. This aspect was  
272 systematically examined in the present study. As shown in Fig. 7, a certain correlation was  
273 observed, and in some cases the isotope effect increased with increasing number of deuterium  
274 atoms in the molecule. As already mentioned in earlier studies [20,25] the isotope effect does  
275 not increase linearly with the number of deuterium atoms part of the molecule and the so-  
276 called specific deuterium effect (incremental isotope effect per added deuterium atom to the  
277 molecule) decreases with increasing the degree of deuteration of a molecule. Moreover, it is  
278 obvious that not just the number of deuterium atoms but also their location in the structure of  
279 molecule plays a certain role in the observed effect. On one hand, this observation suggests  
280 that the separation of isotopomers is possible and correlates with the successful separation of  
281 racemates based on isotopic chirality reported by Tanaka and co-workers [40]. On the other  
282 hand, this observation suggests that not only mass-dependent forces are responsible for the  
283 separation of isotopologues.

284 In the present study, differences between isotopomers were also examined (Fig. 7 and  
285 Supporting Information, Fig. S8). In this regard, it is evident that a minor difference observed  
286 in the retention of isotopomers is (as expected) insufficient to observe their separation under  
287 the used experimental conditions. However, interesting trends could be observed by  
288 examining the variations in the retention times of the various isotopologues (or isotopomers)  
289 as the chromatographic conditions changed:

290 1. Slightly stronger isotope effect was observed for the isotopologue/isotopomer  
291 containing deuterium atoms in the side chain compared to that on the aromatic ring.

292 2. In several cases, the retention of AMP-d<sub>8</sub>, not containing deuterium atoms at C<sub>α</sub> and  
293 C<sub>β</sub> with respect to the NH<sub>2</sub> group, was lower than expected, deviating from the linear-like  
294 trend exhibited by the other isotopologues of the series.

295 3. On the Lux AMP column, the retention of AMP-d<sub>5(ring)</sub> was slightly lower compared  
296 to AMP-d<sub>6</sub> and AMP-d<sub>11</sub> with ACN containing 0.1 % ammonium hydroxide, whereas it was  
297 higher with a mobile phase containing high percentages of water (Supporting Information,  
298 Fig. S9).

299

### 300 3.5. *Possible recognition mechanism underlying isotope effects*

301 In spite of the importance of isotope effects in analytical and bioanalytical separations,  
302 studying the mechanisms and noncovalent interactions underlying these effects by  
303 computational analysis remains rather challenging. The main question arises from the fact that,  
304 under the Born-Oppenheimer approximation, the potential energy surfaces of different isotopes  
305 are the same. Consequently, the difference in the thermodynamic stability between two  
306 isotopomers (or isotopologues) can arise exclusively from the vibrational frequency of C–H  
307 and C–D bonds [41]. Indeed, the C–D bond presents smaller vibrational frequency, shorter bond  
308 length, and lower polarizability compared to the C–H bond [42]. Given these subtle differences,  
309 a very small or non-detectable difference between isotopologues was observed so far [43], and  
310 various methodological approaches often provided opposite results. For instance, whereas  
311 studies on the encapsulation of protic and deuterated guest molecules in molecular capsules led  
312 to the conclusion that the C–D···π interaction is stronger than the C–H···π interaction [44],  
313 others based on chromatographic analysis led to the opposite conclusion. In this field, Tanaka  
314 and co-authors studied the hydrogen/deuterium isotope effects on hydrophobic binding in the  
315 reversed-phase chromatographic separation of protiated and deuterated isotopologues,  
316 observing that protiated compounds bind to nonpolar moieties attached to silica more strongly

317 than the deuterated ones [21]. Later, by the comparison of the free energies of isotopologues  
318 derived in RPLC, Kubo and co-authors hypothesized that the C–H $\cdots\pi$  interaction was slightly  
319 stronger than C–D $\cdots\pi$  interaction providing an inverse isotope effect in the separation of  
320 aromatic hydrocarbons [22]. Therefore, the contribution of dispersion forces to H/D isotope  
321 effects was not conclusively confirmed so far.

322 On this basis, we studied molecular and electronic properties of AMP and MET  
323 conformers optimized at DFT level of theory (Supporting Information, Table S10). The  
324 calculations were performed by using the implicit SMD as solvation model in three different  
325 solvents, i.e. acetonitrile, methanol, and water, in agreement with the experimental mobile  
326 phases. In all cases, two types of lowest-energy conformers were found. The first conformer  
327 type (conformer **a**) featured an intramolecular C–H $\cdots\pi$  interaction ( $2.817 \text{ \AA} \leq d_{\text{CH}\cdots\pi} \leq 2.835 \text{ \AA}$ )  
328 underlying the structure, with the amine hydrogen(s) available as HB donor(s). In the other  
329 conformer type (conformer **b**), the amine hydrogen was found involved in the intramolecular  
330 NH $\cdots\pi$  interaction ( $2.551 \text{ \AA} \leq d_{\text{NH}\cdots\pi} \leq 2.697 \text{ \AA}$ ), thus it was not available for intermolecular HBs  
331 due to stereoelectronic reasons. In particular, conformer **b** of MET exposes an extended  
332 hydrophobic surface on the opposite side from the NH group. The calculated properties showed  
333 that MET presents a higher contribution of dispersion to the total energy and higher  
334 polarizability compared to AMP. To the contrary, this latter shows, in general, lower electron  
335 charge density, and more positive electrostatic potential maxima on the amine hydrogens that,  
336 consequently, exhibited higher ability as HB donors. Furthermore, the amine nitrogen of AMP  
337 featured a more negative value of electrostatic potential, thus higher electron charge density,  
338 compared to the amine nitrogen of MET. On this basis, the higher retention of MET observed  
339 in all cases compared to AMP could be ascribed to dispersion-type forces rather than to HB-  
340 type forces. On this basis, to evaluate the possible contribution of dispersion forces in the  
341 studied analytical (enantio)separations, we calculated the energy of the complexes between

342 conformer **b** of MET and benzene, 1,3-dimethylbenzene, and 1-chloro-3-methylbenzene to  
343 explore the impact of the electronic properties of the substituents on benzene (methyl and/or  
344 chlorine) on the contribution of the dispersion energy to the overall binding energy. As shown  
345 in Fig. 8, in all cases a CH $\cdots\pi$  interaction between one methylene hydrogen of MET and the  
346 aromatic counterpart was observed, as well as a calculated contribution of the dispersion to the  
347 total energy increasing following the order benzene (-0.0750 au) < 1-chloro-3-methylbenzene  
348 (-0.0881 au) < 1,3-dimethylbenzene (-0.0912 au). In this regard, it is worth mentioning that  
349 noncovalent interactions like C–H $\cdots\pi$  present a high character as dispersive forces [45].

350 Based on these computed results and on the structural features of AMP and MET, our  
351 mechanistic hypotheses were the following:

352 1. The medium determines the equilibrium between the conformers of the analyte in  
353 solution, whereas the structure of the (chiral) selector determines how the system of the two  
354 conformers at the equilibrium interacts with (chiral) selector. Indeed, the relative stability of  
355 conformers **a** and **b** depended on the medium. On the other hand, the energy differences were  
356 very low [ $\Delta E = E_{\text{conformer b}} - E_{\text{conformer a}}$ : 0.19 kcal/mol (ACN), 0.15 kcal/mol (MeOH), and 0.06  
357 kcal/mol (water) for AMP and 0.53 (ACN and MeOH), and 0.51 (water) for MET]. Thus, it was  
358 expected that the two conformers could interconvert rather easily.

359 2. Separation and enantioseparation outcomes depend on a subtle balance between HB  
360 and dispersive forces.

361 3. Depending on the location of the deuterium atoms, deuteration favored HB and  
362 disfavored dispersive forces. Indeed, as reported before [26], deuterium is more electronegative  
363 compared to protium when bound to a Csp<sup>3</sup>. Thus, deuterated groups close to the N-H moiety  
364 in the analyte structure can cause its stronger (increased) deshielding (meaning higher  
365 hydrogen-bonding donor capacity) compared to the N-H group in the nondeuterated analyte.



366 On the other hand, the frequency of vibration is inversely proportional to the mass of the atoms,  
367 so heavier atoms vibrate at lower wavenumbers. The C–H bond has a higher oscillation  
368 frequency than the C–D bond (the wavenumbers are 3300 vs. 2334 cm<sup>-1</sup>, respectively). Thus, it  
369 may induce stronger dispersion attraction. This could result in higher retention for the  
370 undeuterated compound compared to its deuterated analogue due to the difference in dispersive  
371 analyte-adsorbent interactions.

372 Given these hypotheses, we calculated (Supporting information, Tables S11-S18) and  
373 compared (Supporting information, Tables S19-S24) in detail the vibration wavenumbers of the  
374 two conformers of AMP and MEP in the three solvents. Indeed, the amount of energy required  
375 to stretch a bond depends on the strength of the bond and the masses of the bound atoms, and  
376 the stronger is the bond, the greater is the energy required to stretch it. Thus, by comparing the  
377 stretching wavenumbers of N-H groups part of different isotopologues, we could evaluate how  
378 the strength of the HB involving the N-H groups changes. In terms of dispersion forces, we  
379 considered the zero-point vibrational energy (ZPVE) as descriptor to compare the dispersion  
380 capability of the different isotopologues. Indeed, the ZPVE results from the vibrational motion  
381 of molecular systems at 0 K, and it is calculated for a harmonic oscillator model as a sum of  
382 contributions for all vibrational degrees of freedom of the system.

$$383 \quad ZPVE = \sum_i 0.5hc\tilde{\nu}_i \quad (1)$$

384 where  $h$  is the Plank constant,  $c$  the light speed,  $\tilde{\nu}$  the vibrational wavenumber, and  $i$   
385 refers to the vibrational degrees of freedom.

386 Interesting correlations between the calculated vibrational wavenumbers and the  
387 experimental retention times were observed:

- 388 1. In all cases, lower  $\tilde{\nu}_{\text{N-H}}$  were calculated for the isotopologues containing  
389 deuterium atoms at C $\alpha$  and C $\beta$  with respect to the NH<sub>2</sub> group, i.e. AMP-d<sub>5(side chain)</sub>,

390 AMP-d<sub>6</sub>, an AMP-d<sub>11</sub> compared to AMP, AMP-d<sub>8</sub>, and AMP-d<sub>5(ring)</sub>. Thus, for the  
391 first series stronger HB donor ability could be expected.

392 2. The ZPVE increased as the deuteration degree and solvent polarity (ACN  
393 < MeOH < water) increased.

394 3. For both conformers of AMP, higher ZPVE was calculated for AMP-  
395 d<sub>5(ring)</sub> compared to AMP-d<sub>5(side chain)</sub>.

396 These results confirmed a close relationship between the retention times observed in the  
397 chromatographic experiments and the descriptors used to quantify the contribution of HB and  
398 dispersive forces to retention and in enantioseparation mechanisms, and in almost all cases, the  
399 lines describing the dependence of retention on the deuteration degree correlated satisfactorily  
400 with the lines describing the dependence of the vibrational wavenumbers on the deuteration  
401 degree. In Figs. 9 and 10, some representative correlations are reported for the separation of  
402 AMP isotopologues with the Biphenyl column and MeOH/water 90:10, 20:80, and 70:30 as  
403 mobile phases. At lower concentration of water (10%) (Fig. 9A), the HB dominates the retention  
404 mechanism and retention time of AMP\_*conformer a* (SMD, MeOH) increases or decreases as  
405 the  $\tilde{\nu}_{\text{N-H}}$  decreases (HB ability increases) or increases (HB ability decreases). In the opposite  
406 situation, at higher concentration of water (80%) (Fig. 9B), the retention time clearly depended  
407 on the ZPVE value, thus in this case the contribution of dispersive forces to the mechanism was  
408 dominant. At intermediate percentages of water (30%) (Fig. 10), retention time depended on  
409 the balance between HB and dispersive forces ( $r^2 = 0.9525$ ). In particular, the decrease in  
410 retention time moving from AMP-d<sub>6</sub> to AMP-d<sub>8</sub> could be explained by the corresponding  
411 increase in the  $\tilde{\nu}_{\text{N-H}}$  (decrease in HB donor ability).

412 This model could explain other experimental behaviors:

413 1. The explanation of the differences observed between AMP and MET on both  
414 Biphenyl and Lux AMP column with ACN, i.e. almost no isotope effect for AMP and a stronger

415 positive isotope effect for MET, could be found in the difference in the  $\tilde{\nu}_{\text{N-H}}$  of the protiated and  
416 the  $d_{5(\text{side chain})}$  isotopologues calculated in the two cases, higher for MET [ $\Delta\tilde{\nu}_{\text{N-H}} = \tilde{\nu}_{\text{N-H}}(\text{MET})$   
417  $- \tilde{\nu}_{\text{N-H}}(\text{MET-}d_{5(\text{side chain})}) = 0.19 \text{ cm}^{-1}$ ] compared to AMP [ $\Delta\tilde{\nu}_{\text{N-H}} = \tilde{\nu}_{\text{N-H}}(\text{AMP}) - \tilde{\nu}_{\text{N-H}}(\text{AMP-}d_{5(\text{side}}$   
418  $\text{chain}))$ ,  $0.01 \text{ cm}^{-1} \leq \Delta\tilde{\nu}_{\text{N-H}} \leq 0.09 \text{ cm}^{-1}$ ].

419 2. For the (enantio)separation of AMP- $d_{5(\text{side chain})}$ , AMP- $d_6$ , AMP- $d_{11}$ , and AMP- $d_{5(\text{ring})}$   
420 series with the Lux AMP column in ACN-containing mixture, the retention time order ( $d_{5(\text{ring})} <$   
421  $d_{11}$ ,  $d_6$ ,  $d_{5(\text{side chain})}$ ) in pure ACN correlated well with the change in  $\tilde{\nu}_{\text{N-H}}$  ( $d_{5(\text{ring})} > d_{11}$ ,  $d_6$ ,  $d_{5(\text{side}}$   
422  $\text{chain})$ ), whereas retention time changed (Fig. S9) as the ZPVE values at the highest percentages  
423 of water (80%) [RT(1) (min), ZPVE AMP\_a (SMD, water) (H/particle):  $d_{5(\text{ring})}$  (26.85,  
424  $0.186800) > d_{5(\text{side chain})}$  (26.66,  $0.186654) > d_6$  (26.50,  $0.183278) > d_{11}$  (25.27,  $0.166848)$ ].

425

#### 426 4. Conclusions

427 A strong isotope effect was observed for partially deuterated AMP derivatives under  
428 some experimental conditions enabling their baseline separation on achiral and  
429 polysaccharide-based chiral columns in HPLC. The nature (positive or negative) of the  
430 isotope effect and its extent strongly depends on the nature of selector, medium and structure  
431 of the studied compounds. On chiral columns some correlations were observed between the  
432 strength of the isotope effect and selectivity of enantioseparation. The isotope effect increased  
433 with increasing number of deuterium atoms in the molecule. Some differences in the retention  
434 of isotopomers were observed but this was not sufficient for their separation. Analysis of the  
435 vibrational wavenumbers and related ZPVE calculated for both AMP and MET isotopologues  
436 by quantum mechanics allowed to disclose some essential factors contributing to retention  
437 and enantioselectivity, profiling a model based on the interplay between HB- and dispersive-  
438 type interactions. The number and location of the deuterium atoms in the AMP and the MET  
439 impact the strength of these interactions. Increasing deuteration degree, in particular at

440 positions close to the amino group, was found to favor hydrogen bonding-type forces,  
441 whereas it was detrimental to dispersive forces.

442 This study reporting on concomitant significant differences observed in the isotope  
443 effect and enantioseparation based on the structure of the chiral selector, mobile phase and the  
444 structure of analytes may provide important material for better understanding of  
445 enantioselective recognition mechanisms.

446

#### 447 **CRedit authorship contribution statement**

448 **Giorgia Sprega, Giorgi Kobidze, Barbara Sechi:** Formal analysis, Investigation. **Alfredo**  
449 **Fabrizio Lo Faro:** Formal analysis, Investigation, Supervision. **Paola Peluso:**  
450 Conceptualization, Methodology, Formal analysis, Investigation, Writing. **Tivadar Farkas:**  
451 Writing – review & editing, Resources. **Francesco Paolo Busardò:** Conceptualization,  
452 Methodology, Formal analysis, Investigation, Supervision, Resources. **Bezhan**  
453 **Chankvetadze:** Conceptualization, Methodology, Formal analysis, Investigation,  
454 Supervision, Validation, Writing – original draft, Writing – review & editing.

455

#### 456 **Declaration of Competing Interest**

457 The authors declare that they have no known competing financial interests or personal  
458 relationships that could have appeared to influence the work reported in this paper.

459

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466

467 **5. References**

- 468 [1] J. Atzrodt, V. Derdau, W.J. Kerr, M. Reid, Deuterium-and tritium-labelled compounds:  
469 applications in the life sciences, *Angew. Chem. Int. Ed. Engl.* 57 (2018) 1758-1784.  
470 doi:10.1002/anie.201704146.
- 471 [2] C. Schmidt, First deuterated drug approved, *Nat. Biotechnol.* 35 (2017) 493-494.  
472 doi:10.1038/nbt0617-493.
- 473 [3] S.J. Keam, S. Duggan, Donafenib: first approval. *Drugs* 81 (2021) 1915-1920.  
474 doi:10.1007/s40265-021-01603-0.
- 475 [4] S.T. Wroblewski, R. Moslin, S. Lin, Y. Zhang, S. Spengel, J. Kempson, J.S. Tokarski, J.  
476 Strnad, A. Zupa-Fernandez, L. Cheng, D. Shuster, K. Gillooly, X. Yang, E. Heimrich,  
477 K.W. McIntyre, C. Chaudhry, J. Khan, M. Ruzanov, J. Tredup, D. Mulligan, D. Xie, H.  
478 Sun, C. Huang, C. D'Arienzo, N. Aranibar, M. Chiney, A. Chimalakonda, W.J. Pitts, L.  
479 Lombardo, P.H. Carter, J.R. Burke, D.S. Weinstein, Highly selective inhibition of  
480 tyrosine kinase 2 (TYK2) for the treatment of autoimmune diseases: discovery of the  
481 allosteric inhibitor BMS-986165, *J. Med. Chem.* 62 (2019) 8973-8995.  
482 doi:10.1021/acs.jmedchem.9b00444.
- 483 [5] R.M.C. Di Martino, B.D. Maxwell, T. Pirali, Deuterium in drug discovery: progress,  
484 opportunities and challenges, *Nat. Rev. Drug Discov.* 22 (2023) 562-584.  
485 doi:10.1038/s41573-023-00703-8.
- 486 [8] J. Bigeleisen, M.G. Mayer, Calculation of equilibrium constants for isotopic exchange  
487 reactions, *Int. J. Chem. Phys.* 261 (1947) 261-267. <https://doi.org/10.1063/1.1746492>.
- 488 [6] Y. Fu, D. Barkley, W. Li, F. Picard, J. Flarakos, Evaluation, identification and impact  
489 assessment of abnormal internal standard response variability in regulated LC-MS  
490 bioanalysis, *Bioanalysis* 12 (2020) 545-559. doi:10.4155/bio-2020-0058.
- 491 [7] L. Heinle, K. Sulaiman, A. Olson, K. Ruterbories, A homologous series of internal  
492 standards for near universal application in the discovery LC-MS/MS bioanalytical  
493 laboratory, *J. Pharm. Biomed. Anal.* 190 (2020) 113578.  
494 doi:10.1016/j.jpba.2020.113578.
- 495 [9] K.E. Wilzbach, P. Riesz, Isotope Effects in Gas-Liquid Chromatography, *Science* 126  
496 (1957) 748-749. doi:10.1126/science.126.3277.748.
- 497 [10] S. Ohkoshi, Y. Fujita, T. Kwan, Gas chromatographic separation of hydrogen isotopes  
498 D<sub>2</sub> and HD, *Bull. Chem. Soc. Jpn.* 31 (1958) 770-771.  
499 <https://doi.org/10.1246/bcsj.31.770>.
- 500 [11] F. Bruner, G. Cartoni, A. Liberti, Gas chromatography of isotopic molecules on open  
501 tubular columns, *Anal. Chem.* 38 (1966) 298-303.  
502 <https://doi.org/10.1021/ac60234a035>.
- 503 [12] B. Shi, B.H. Davis, Gas chromatographic separation of pairs of isotopic molecules. *J.*  
504 *Chromatogr. A* 654 (1993) 319-325. [https://doi.org/10.1016/0021-9673\(93\)83377-5](https://doi.org/10.1016/0021-9673(93)83377-5).
- 505 [13] H.G. Schmarr, M. Wacker, M. Mathes, Isotopic separation of acetaldehyde and  
506 methanol from their deuterated isotopologues on a porous layer open tubular column

- 507 allows quantification by stable isotope dilution without mass spectrometric detection,  
508 *J. Chromatogr. A.* 1481 (2017) 111-115. doi:10.1016/j.chroma.2016.12.023.
- 509 [14] N. Thakur, S. Aslani, D.W. Armstrong, Evaluation of gas chromatography for the  
510 separation of a broad range of isotopic compounds, *Anal. Chim. Acta* 1165 (2021)  
511 338490. doi:10.1016/j.aca.2021.338490.
- 512 [15] S. Aslani, D.W. Armstrong, Effect of position of deuterium atoms on gas  
513 chromatographic isotope effects, *Talanta* 265 (2023) 124857.  
514 doi:10.1016/j.talanta.2023.124857.
- 515 [16] M.D. Chermá, G.H. Nilsson, A. Johansson, A.K. Jönsson, J. Ahlner, Use of  
516 Lisdexamphetamine or Amphetamine? Interpretation of Chiral Amphetamine Analyses,  
517 *J. Anal. Toxicol.* 46 (2022) 10-16. doi:10.1093/jat/bkaa170.
- 518 [17] H. Gottschling, E. Freese, A tritium isotope effect on ion exchange chromatography,  
519 *Nature* 196 (1962) 829-831. doi:10.1038/196829a0.
- 520 [18] S.S Iyer, K.P. Zhang, G.E. Kellogg, H.T. Karnes, Evaluation of deuterium isotope  
521 effects in normal-phase LC-MS-MS separations using a molecular modeling approach,  
522 *J. Chromatogr. Sci.* 42 (2004) 383-387. doi:10.1093/chromsci/42.7.383.
- 523 [19] J.J. Pratt, Isotope dilution analysis using chromatographic separation of isotopic forms  
524 of the compound to be measured, *Ann. Clin. Biochem.* 23 (1986) 251-276.  
525 doi:10.1177/000456328602300305.
- 526 [20] C.F. Masters, S.P. Markey, I.N. Mefford, M.W. Duncan, Separation of deuteriated  
527 isotopomers of dopamine by ion-pair reversed-phase high-performance liquid  
528 chromatography, *Anal. Chem.* 60 (1988) 2131-2134. doi:10.1021/ac00170a029.
- 529 [21] M. Turowski, N. Yamakawa, J. Meller, K. Kimata, T. Ikegami, K. Hosoya, N. Tanaka,  
530 E.R. Thornton, Deuterium isotope effects on hydrophobic interactions: the importance  
531 of dispersion interactions in the hydrophobic phase, *J. Am. Chem. Soc.* 125 (2003)  
532 13836-13849. doi:10.1021/ja036006g.
- 533 [22] E. Kanao, T. Kubo, T. Naito, T. Sano, M. Yan, N. Tanaka, K. Otsuka, Tunable Liquid  
534 Chromatographic Separation of H/D Isotopologues Enabled by Aromatic  $\pi$   
535 Interactions, *Anal. Chem.* 92 (2020) 4065-4072. doi:10.1021/acs.analchem.9b05672
- 536 [23] S. Szarka, K. Prokai-Tatrai, L. Prokai, Application of screening experimental designs  
537 to assess chromatographic isotope effect upon isotope-coded derivatization for  
538 quantitative liquid chromatography-mass spectrometry, *Anal. Chem.* 86 (2014) 7033-  
539 7040. doi:10.1021/ac501309s
- 540 [24] S. Souverain, C. Eap, J.L. Veuthey, S. Rudaz, Automated LC-MS method for the fast  
541 stereoselective determination of methadone in plasma, *Clin. Chem. Lab. Med.* 41  
542 (2003) 1615-1621. doi:10.1515/CCLM.2003.245.
- 543 [25] A. Valleix, S. Carrat, C. Caussignac, E. Léonce, A. Tchaplá, Secondary isotope effects  
544 in liquid chromatography behaviour of 2H and 3H labelled solutes and solvents, *J*  
545 *Chromatogr A* 1116 (2006) 109-126. doi:10.1016/j.chroma.2006.03.078.

- 546 [26] G. Kobidze, G. Sprega, G. Daziani, A. Balloni, A.F. Lo Faro, T. Farkas, P. Peluso, G.  
547 Basile, F.P. Busardò, B. Chankvetadze, Separation of undeuterated and partially  
548 deuterated enantioisotopologues of some amphetamine derivatives on achiral and  
549 polysaccharide-based chiral columns in high-performance liquid chrom (atography, *J.*  
550 *Chromatogr. A.* 1718 (2024) 464709. doi:10.1016/j.chroma.2024.464709.
- 551 [27] M.M. Bushey, J.W. Jorgenson, Separation of Dansylated Methylamine and Dansylated  
552 Methyl-d3-amine by Micellar Electrokinetic Capillary Chromatography with  
553 Methanol-Modified Mobile Phase, *Anal. Chem.* 61 (1989) 491-493.  
554 doi:10.1021/ac00180a022.
- 555 [28] M.M. Bushey, J.W. Jorgenson, Effects of methanol-modified mobile phase on the  
556 separation of isotopically substituted compounds by micellar electrokinetic capillary  
557 chromatography, *J. Microcol. Sep.* 1 (1989) 125-130. doi:10.1002/mcs.1220010304.
- 558 [29] Y. Shao, L.F. Molnar, Y. Jung, J. Kussmann, C. Ochsenfeld, S.T. Brown, A.T. Gilbert,  
559 L.V. Slipchenko, S.V. Levchenko, D.P. O'Neill, R.A. Jr DiStasio, R.C. Lochan, T.  
560 Wang, G.J. Beran, N.A. Besley, J.M. Herbert, C.Y. Lin, T. Van Voorhis, S.H. Chien, A.  
561 Sodt, R.P. Steele, V.A. Rassolov, P.E. Maslen, P.P. Korambath, R.D. Adamson, B.  
562 Austin, J. Baker, E.F. Byrd, H. Dachsel, R.J. Doerksen, A. Dreuw, B.D. Dunietz, A.D.  
563 Dutoi, T.R. Furlani, S.R. Gwaltney, A. Heyden, S. Hirata, C.P. Hsu, G. Kedziora, R.Z.  
564 Khallilulin, P. Klunzinger, A.M. Lee, M.S. Lee, W. Liang, I. Lotan, N. Nair, B. Peters,  
565 E.I. Proynov, P.A. Pieniazek, Y.M. Rhee, J. Ritchie, E. Rosta, C.D. Sherrill, A.C.  
566 Simmonett, J.E. Subotnik, H.L. 3rd Woodcock, W. Zhang, A.T. Bell, A.K. Chakraborty,  
567 D.M. Chipman, F.J. Keil, A. Warshel, W.J. Hehre, H.F. 3rd Schaefer, J. Kong, A.I.  
568 Krylov, P.M. Gill, M. Head-Gordon, Advances in methods and algorithms in a modern  
569 quantum chemistry program package, *Phys. Chem. Chem. Phys.* 8 (2006) 3172-3191.  
570 doi: 10.1039/B517914A.
- 571 [30] Gaussian 16, Revision C.01, M.J. Frisch, G.W. Trucks, H.B. Schlegel, G.E. Scuseria,  
572 M.A. Robb, J.R. Cheeseman, G. Scalmani, V. Barone, G.A. Petersson, H. Nakatsuji,  
573 X. Li, M. Caricato, A.V. Marenich, J. Bloino, B.G. Janesko, R. Gomperts, B.  
574 Mennucci, H.P. Hratchian, J.V. Ortiz, A.F. Izmaylov, J.L. Sonnenberg, D. Williams-  
575 Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson,  
576 D. Ranasinghe, V.G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M.  
577 Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O.  
578 Kitao, H. Nakai, T. Vreven, K. Throssell, J.A. Montgomery Jr., J.E. Peralta, F. Ogliaro,  
579 M.J. Bearpark, J.J. Heyd, E.N. Brothers, K.N. Kudin, V.N. Staroverov, T.A. Keith, R.  
580 Kobayashi, J. Normand, K. Raghavachari, A.P. Rendell, J.C. Burant, S.S. Iyengar, J.  
581 Tomasi, M. Cossi, J.M. Millam, M. Klene, C. Adamo, R. Cammi, J.W. Ochterski, R.L.  
582 Martin, K. Morokuma, O. Farkas, J.B. Foresman, D.J. Fox, Gaussian, Inc.,  
583 Wallingford CT, (2016).
- 584 [31] S. Grimme, J. Antony, S. Ehrlich, H. Krieg, A consistent and accurate ab initio  
585 parameterization of density functional dispersion correction (DFT-D) for the 94  
586 elements H-Pu, *J. Chem. Phys.* 132 (2010) 154104. doi: 10.1063/1.3382344.
- 587 [32] A.V. Marenich, C.J. Cramer, D.G. Truhlar, Universal solvation model based on solute  
588 electron density and a continuum model of the solvent defined by the bulk dielectric



- 589 constant and atomic surface tensions, *J. Phys Chem. B.* 113 (2009) 6378-6396. doi:  
590 10.1021/jp810292n]
- 591 [33] T. Lu, F. Chen, Multiwfn: a multifunctional wavefunction analyser, *J. Comput. Chem.*  
592 33 (2012) 580-592. doi: 10.1002/jcc.22885.
- 593 [34] T. Lu, F. Chen, Quantitative analysis of molecular surface based on improved  
594 Marching Tetrahedra algorithm, *J. Mol. Graph. Model.* 38 (2012) 314-323. doi:  
595 10.1016/j.jmgm.2012.07.004.
- 596 [35] S. Grimme, S. Ehrlich, L. Goerigk, Effect of the damping function in dispersion  
597 corrected density functional theory, *J. Comp. Chem.* 32 (2011) 1456-1465. doi:  
598 10.1002/jcc.21759.
- 599 [36] B. Chankvetadze, C. Yamamoto, Y. Okamoto, Enantioseparation of selected chiral  
600 sulfoxides using polysaccharide-type chiral stationary phases and polar organic, polar  
601 aqueous-organic and normal-phase eluents, *J. Chromatogr. A* 922 (2001) 127-137.  
602 doi:10.1016/s0021-9673(01)00958-x.
- 603 [37] G. Jibuti, A. Mskhiladze, N. Takaishvili, M. Karchkhadze, L. Chankvetadze, T. Farkas,  
604 B. Chankvetadze, HPLC separation of dihydropyridine derivatives enantiomers with  
605 emphasis on elution order using polysaccharide-based chiral columns, *J. Sep. Sci.* 35  
606 (2012) 2529-2537. doi:10.1002/jssc.201200443.
- 607 [38] I. Matarashvili, D. Ghughunishvili, L. Chankvetadze, N. Takaishvili, T. Khatiashvili,  
608 M. Tsintsadze, T. Farkas, B. Chankvetadze, Separation of enantiomers of chiral weak  
609 acids with polysaccharide-based chiral columns and aqueous-organic mobile phases in  
610 high-performance liquid chromatography: Typical reversed-phase behavior? *J.*  
611 *Chromatogr. A.* 1483 (2017) 86-92. doi:10.1016/j.chroma.2016.12.064.
- 612 [39] S. Materazzo, S. Carradori, R. Ferretti, B. Gallinella, D. Secci, R. Cirilli, Effect of the  
613 water content on the retention and enantioselectivity of albendazole and fenbendazole  
614 sulfoxides using amylose-based chiral stationary phases in organic-aqueous  
615 conditions, *J. Chromatogr. A.* 1327 (2014) 73-79. doi:10.1016/j.chroma.2013.12.051.
- 616 [40] K. Kimata, K. Hosoya, T. Araki, N. Tanaka, Direct chromatographic separation of  
617 racemates on the basis of isotopic chirality, *Anal. Chem.* 69 (1997) 2610-2612.  
618 doi:10.1021/ac970338k.
- 619 [41] S. Puricelli, G. Bruno, C. Gatti, A. Ponti, M. Mella, Viability of hydrogen isotopes  
620 separation via heterolytic dissociation-driven Chemical Affinity Quantum Sieving on  
621 inexpensive alkali-earth oxides, *Appl. Surf. Chem.* 657 (2024) 159596.  
622 <https://doi.org/10.1016/j.apsusc.2024.159596>.
- 623 [42] Y.Y. Zhan, Q.C. Jiang, K. Ishii, T. Koide, O. Kobayashi, T. Kojima, S. Takahashi, M.  
624 Tachikawa, S. Uchiyama, S. Hiraoka, Polarizability and isotope effects on dispersion  
625 interactions in water, *Commun. Chem.* 2 (2019) 141. doi: 10.1038/s42004-019-0242-  
626 0.
- 627 [43] C. Zhao, R.M. Parrish, M.D. Smith, P.J. Pellechia, C.D. Sherrill, K.D. Shimizu, Do  
628 deuteriums form stronger CH- $\pi$  interactions? *J. Am. Chem. Soc.* 134 (2012) 14306-  
629 14309. doi: 10.1021/ja305788p.

- 630 [44] T. Haino, K. Fukuta, H. Iwamoto, S. Iwata, Noncovalent isotope effect for guest  
631 encapsulation in self-assembled molecular capsules, *Chem. Eur. J.* 15 (2009) 13286-  
632 13290. doi: 10.1002/chem.200902526.
- 633 [45] E. Arunan, G.R. Desiraju, R.A. Klein, J. Sadlej, S. Scheiner, I. Alkorta, D.C. Clary,  
634 R.H. Crabtree, J.J. Dannenberg, P. Hobza, H.G. Kjaergaard, A.C. Legon, B. Mennucci,  
635 D.J. Nesbitt, Defining the hydrogen bond: An account (IUPAC Technical Report), *Pure*  
636 *Appl. Chem.* 83 (2011) 1619-1636. doi: 10.1351/PAC-REP-10-01-01.
- 637

638 **Legends:**

639 **Fig. 1** Structure of AMP and MET isotopologues part of this study.

640 **Fig. 2** Separation of AMP and AMP-d<sub>5</sub> on a Lux AMP (250 x 4.6 mm) column with the  
641 mobile phases made of MeOH with 0.1 % (v/v) ammonium hydroxide (a), 5 mM  
642 ammonium bicarbonate in H<sub>2</sub>O (pH-11.0) and MeOH in the ratio (v/v) 10 : 90 (b) and  
643 40 : 60 (c). Flow rate: 1 ml/min. MS-detection conditions were as described in Table 1.

644 **Fig. 3** Separations of MET and MET-d<sub>5</sub> on a Lux AMP (250 x 4.6 mm) column with the  
645 mobile phases made of MeOH with 0.1% (v/v) ammonium hydroxide (a), 5 mM  
646 ammonium bicarbonate in H<sub>2</sub>O (pH-11.0) and MeOH in the ratio (v/v) 10 : 90 (b) and  
647 40 : 60 (c). Flow rate: 1 ml/min. MS-detection conditions were as described in Table 1.

648 **Fig. 4** Separation of AMP and AMP-d<sub>5</sub> on a Lux i-Amylose-3 (250 x 4.6 mm) column with  
649 the mobile phases made of MeOH with 0.1 % (v/v) ammonium hydroxide (a), 5 mM  
650 ammonium bicarbonate in H<sub>2</sub>O (pH-11.0) and MeOH in the ratio (v/v) 5 : 95 (b) and  
651 50 : 50 (c). Flow rate: 1 ml/min. MS-detection conditions were as described in Table 1.

652 **Fig. 5** Separation of AMP, AMP-d<sub>5</sub> (a), MET and MET-d<sub>5</sub> (b) on a Biphenyl (100 x 2.1mm)  
653 column with the mobile phase acetonitrile + 0.1% NH<sub>4</sub>OH. Flow rate: 0.4 ml/min (a).  
654 MS-detection conditions were as described in Table 1.

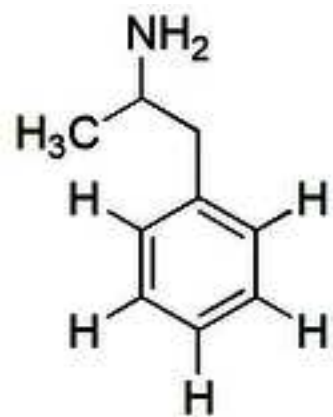
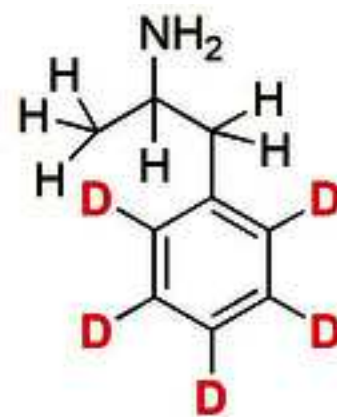
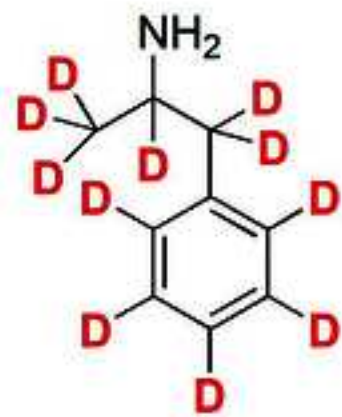
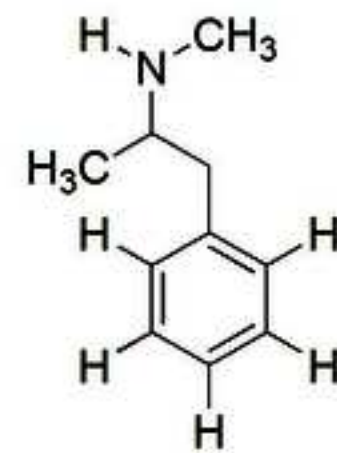
655 **Fig. 6** Separation of AMP, AMP-d<sub>5</sub> (a), MET and MET-d<sub>5</sub> (b) on a Lux i-Amylose-3 (250 x  
656 4.6 mm) column with the mobile phase acetonitrile + 0.1% NH<sub>4</sub>OH. Flow rate: 0.4  
657 ml/min (a). MS-detection conditions were as described in Table 1.

658 **Fig. 7** Separation of AMP and its isotopologues with various degree of deuteration on a Lux  
659 AMP (250 x 4.6 mm) column in the mobile phase: 5mM ammonium bicarbonate in  
660 H<sub>2</sub>O (pH-11.0)/MeOH 40:60, Flow rate: 1ml/min. MS-detection conditions were as  
661 described in Table 1.

662 **Fig. 8** DFT optimized structures for complexes between MET conformer **b** and benzene (A),  
663 1,3-dimethylbenzene (B) and 1-chloro-3-methylbenzene (C). Legend colour: C, grey;  
664 Cl, green; H, pale grey; N, blue.

665 **Fig. 9** Representative correlation between retention times of AMP isotopologues on the  
666 achiral Biphenyl column [MeOH/water 90:10 (A) and 20:80 (B)], quantum mechanics  
667 calculated vibrational frequencies ( $\nu$ ,  $\text{cm}^{-1}$ ), and zero-point vibrational energy (ZPVE,  
668 Hartree/particle): AMP conformer **a**; SMD, MeOH (A), AMP\_conformer **b**; SMD,  
669 water (B).

670 **Fig. 10** Representative correlation between retention times of AMP isotopologues on the  
671 achiral Biphenyl column (MeOH/water 70:30), quantum mechanics calculated  
672 vibrational frequencies ( $\nu$ ,  $\text{cm}^{-1}$ ) and zero-point vibrational energy (ZPVE,  
673 Hartree/particle): AMP\_conformer **a**; SMD, MeOH.

**AMP****AMP-d<sub>5</sub>(side chain)****AMP-d<sub>5</sub>(ring)****AMP-d<sub>6</sub>****AMP-d<sub>8</sub>****AMP-d<sub>11</sub>****MET****MET-d<sub>5</sub>**

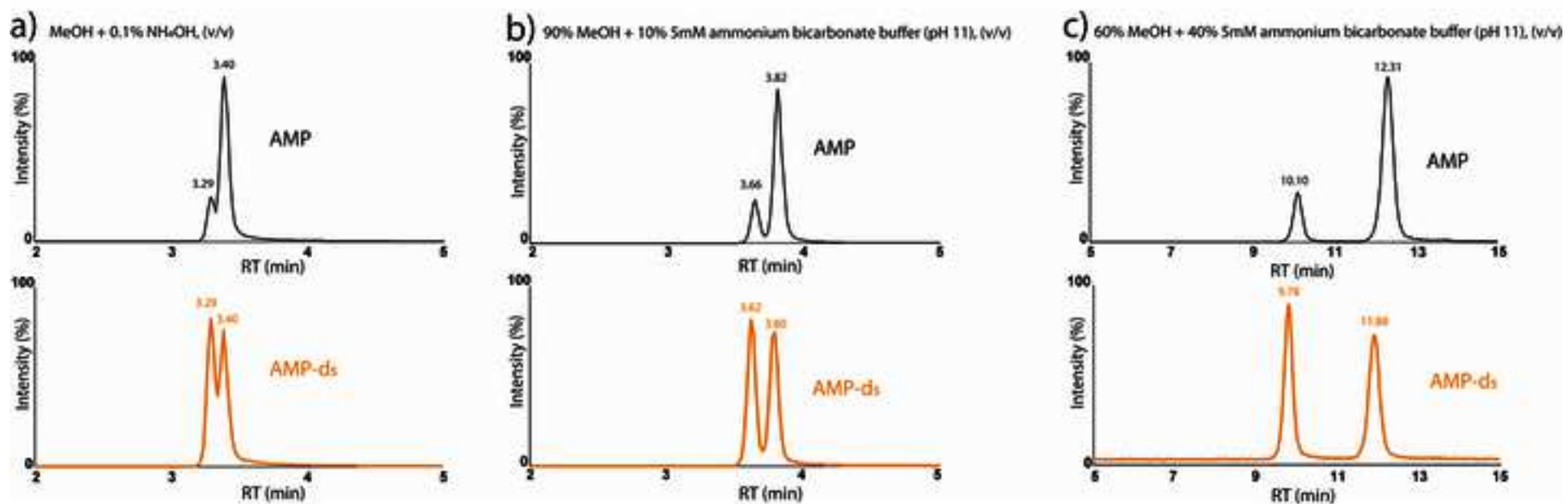


Fig. 2

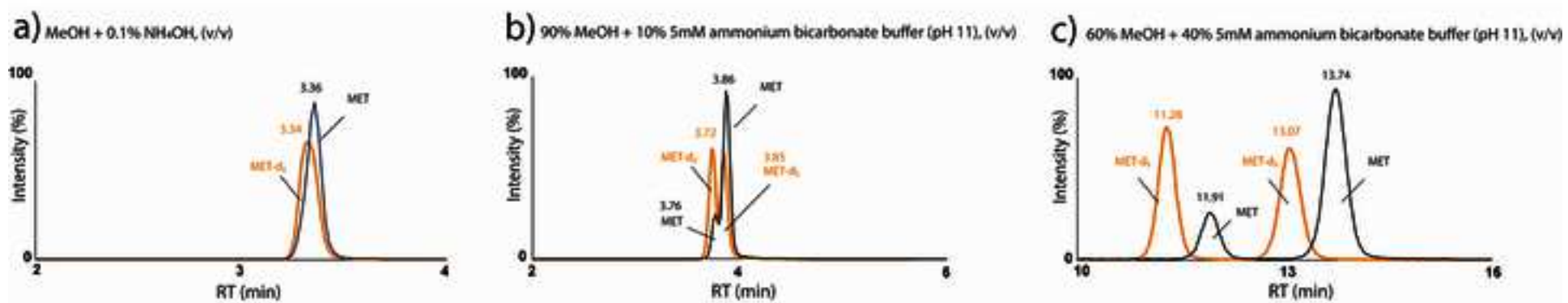


Fig. 3

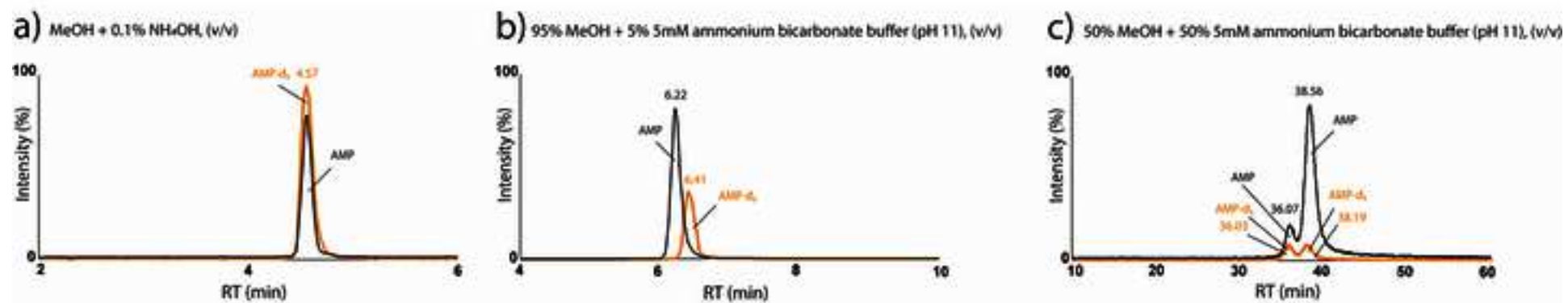


Fig. 4



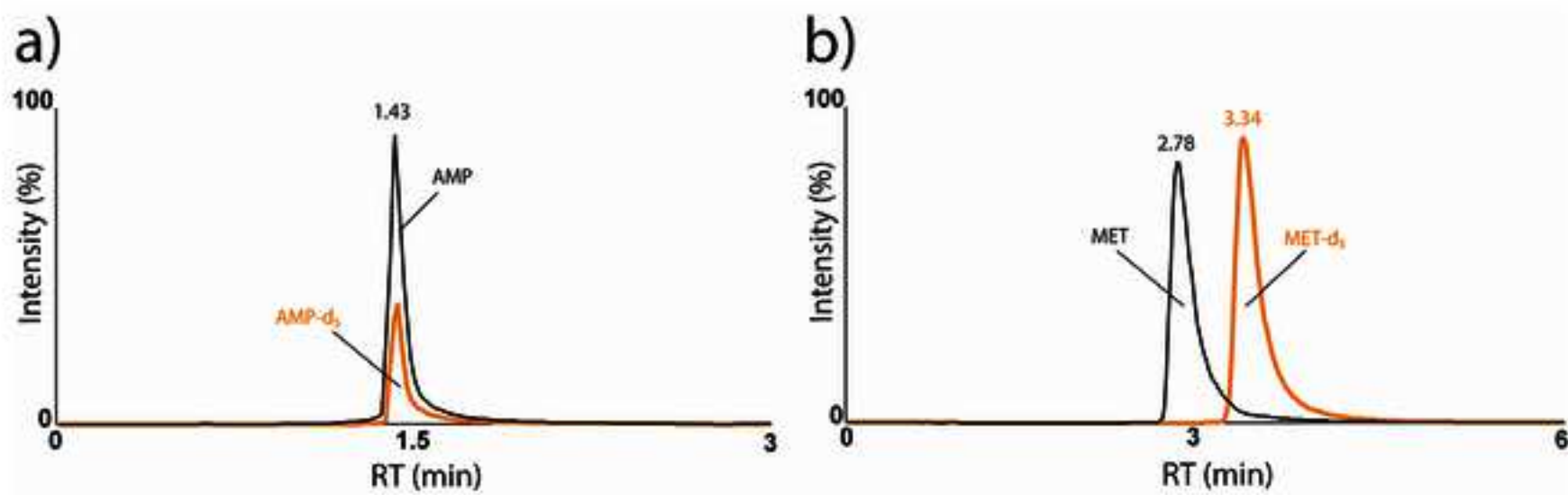


Fig. 5

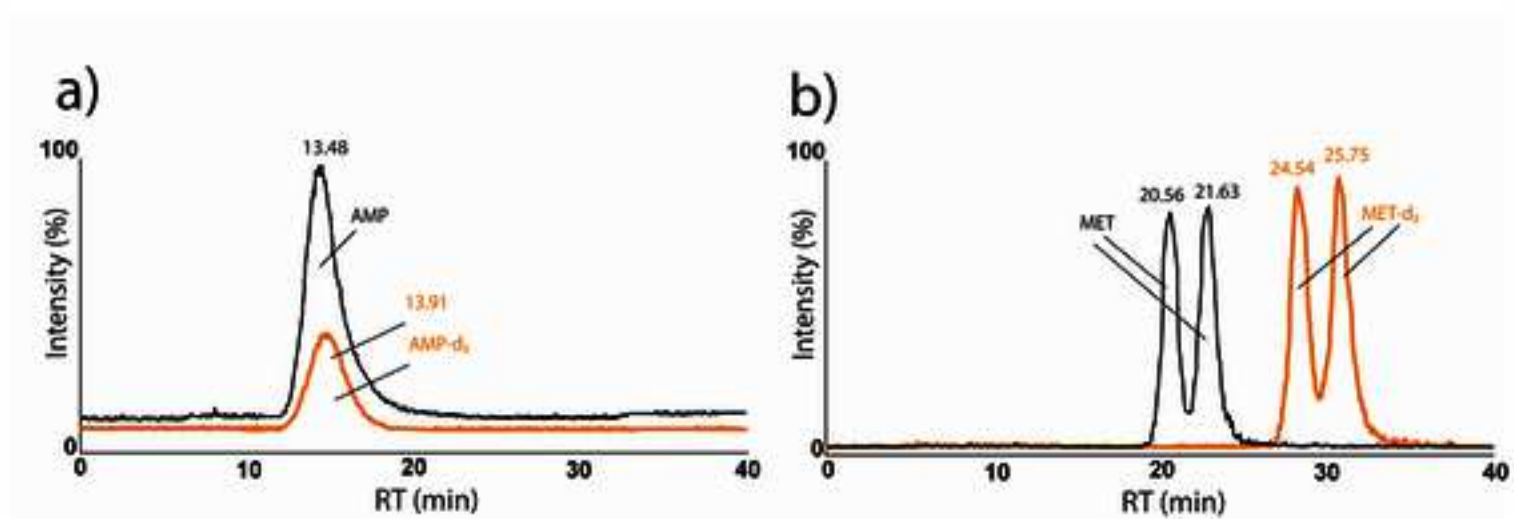


Fig. 6

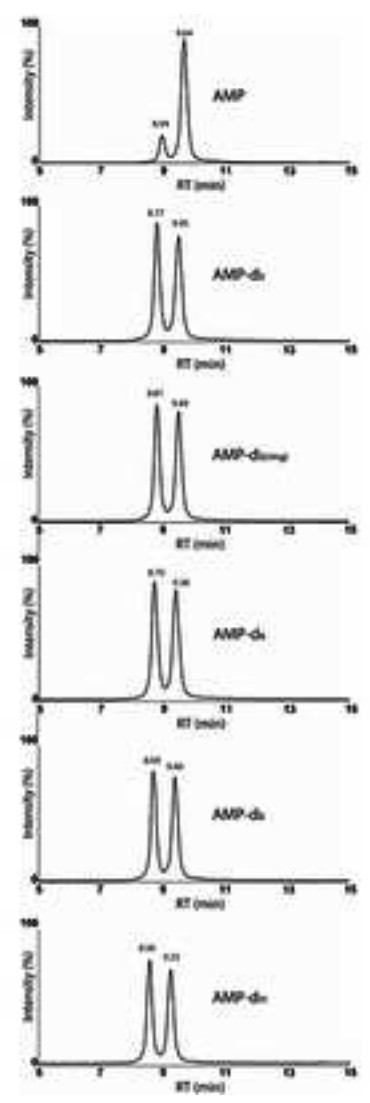
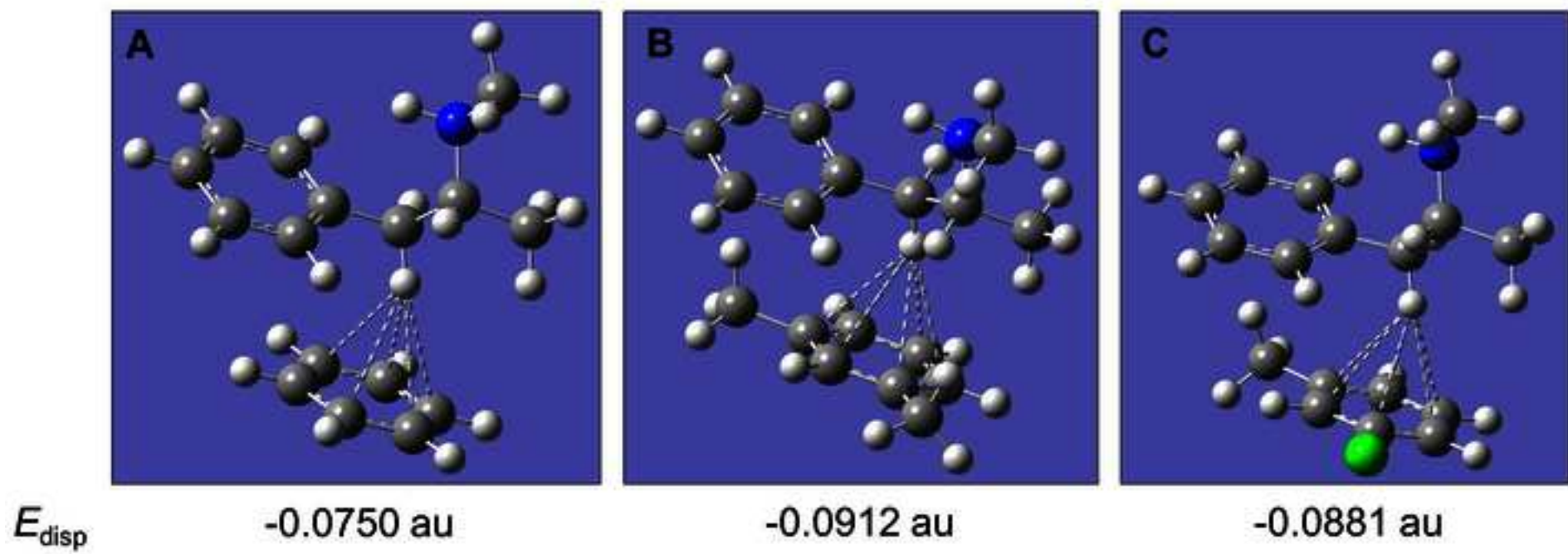
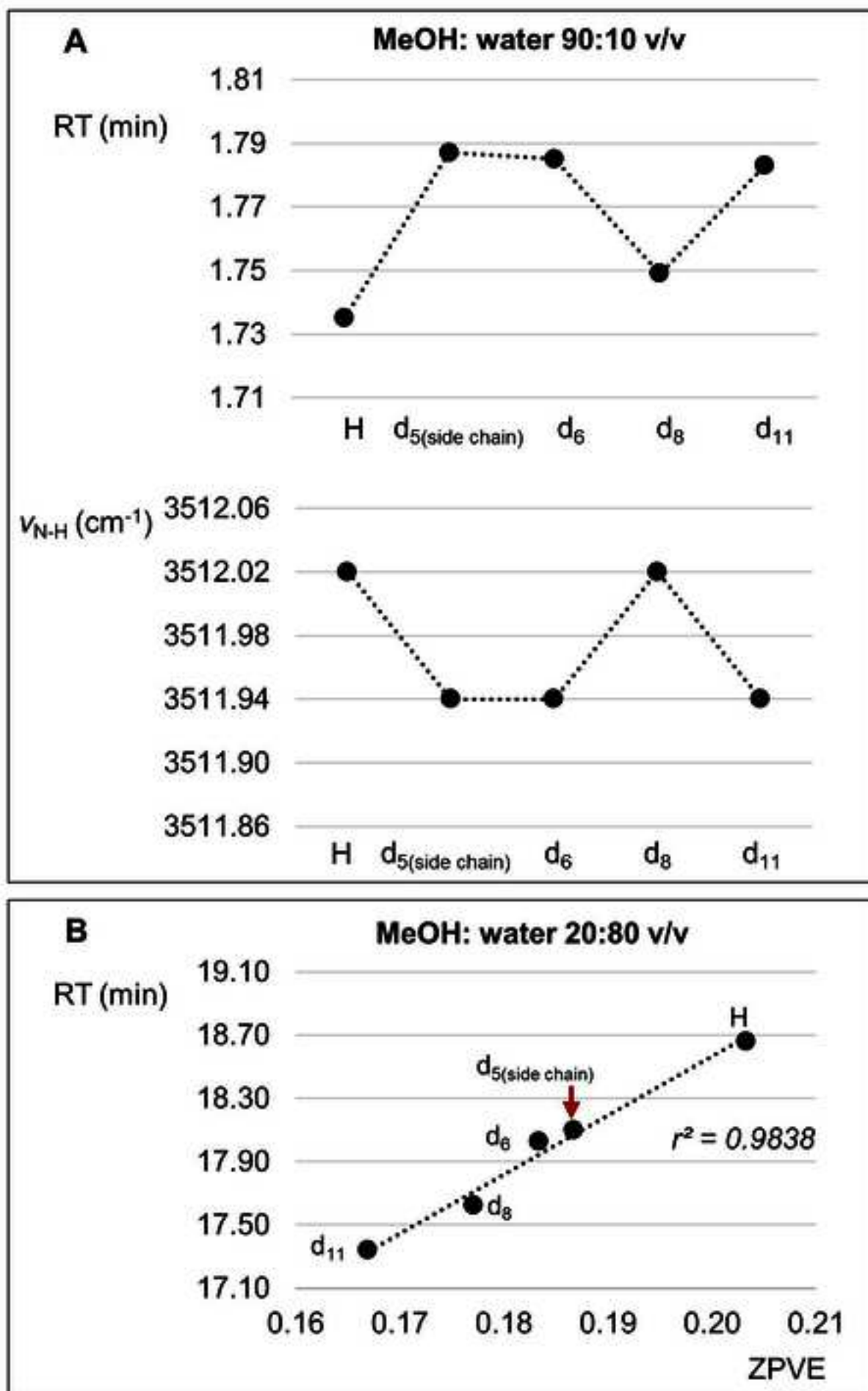
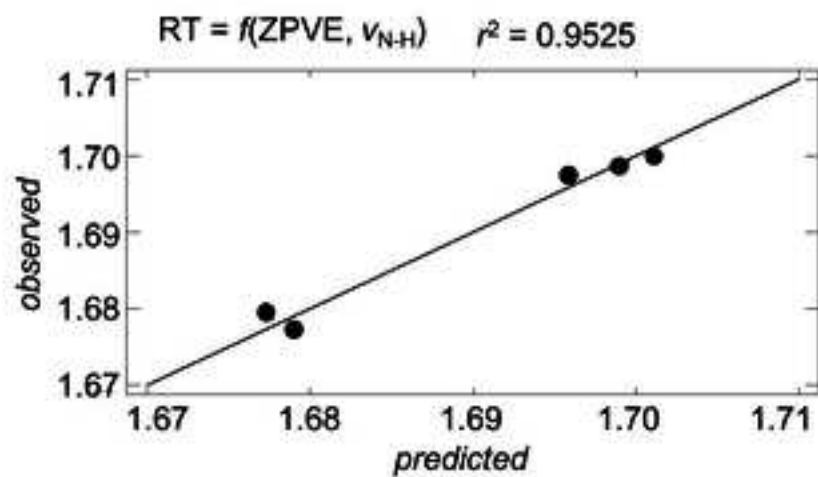
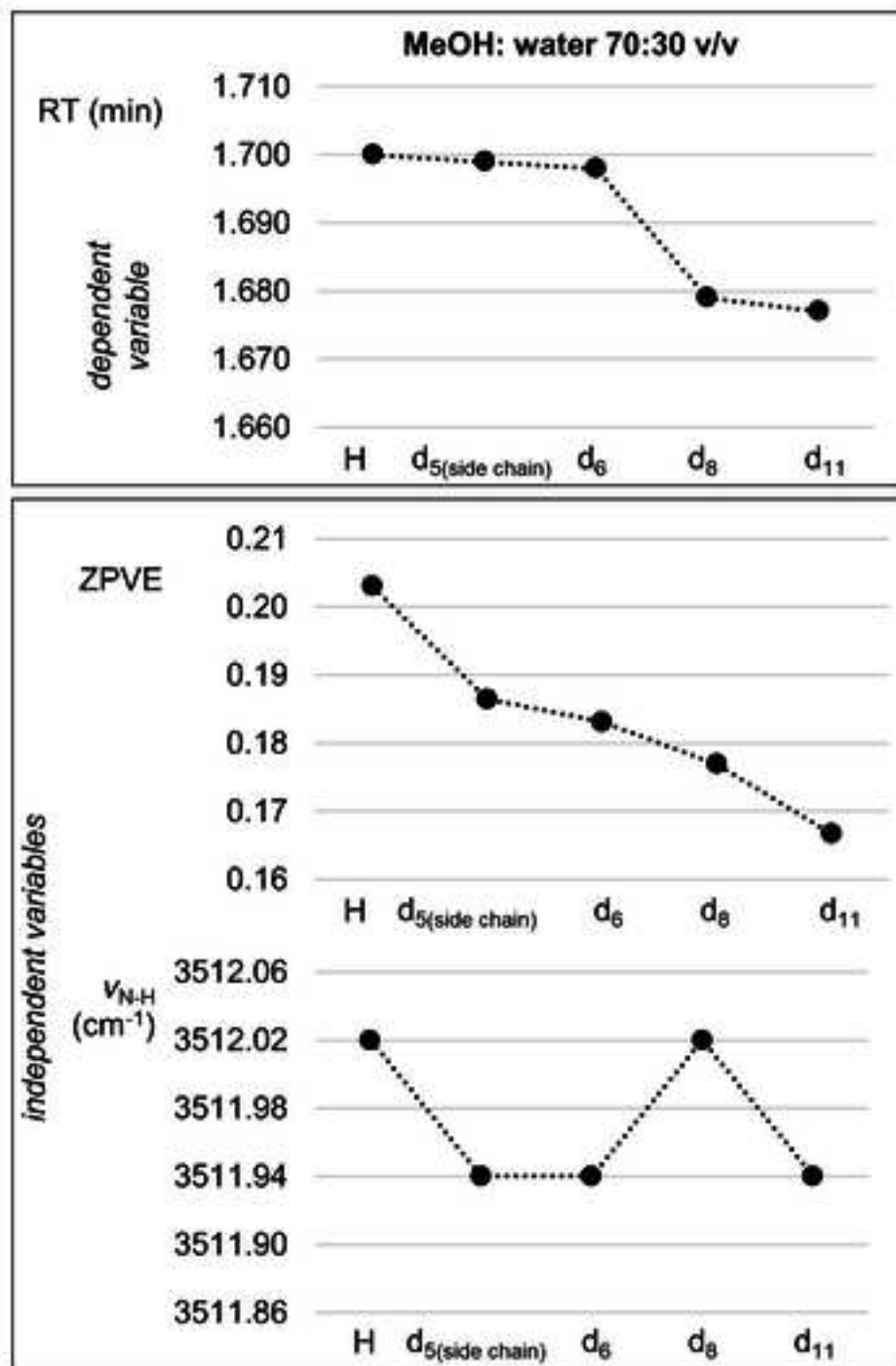


Fig. 7







**Table 1** Mass spectrometry parameters for analytes and internal standards in the positive ionization mode.

Analytes	Molecular mass, g/mol	Precursor ion, m/z	Product ion, m/z	CE, eV
AMPHETAMINE	135.1	136.1	119.1	5
			91.1	13
AMPHETAMINE-d <sub>5</sub> (side chain)	140.2	141.25	124.1	8
			93.1	24
AMPHETAMINE-d <sub>5</sub> (ring)	140.2	141.25	96.1	16
			68.1	44
AMPHETAMINE-d <sub>6</sub>	141.2	142.25	125.1	8
			93.1	16
AMPHETAMINE-d <sub>8</sub>	143.2	144.25	127.1	8
			97.1	16
AMPHETAMINE-d <sub>11</sub>	146.28	147.28	130.1	8
			98.1	24
METHAMPHETAMINE	149.2	150.2	119.1	8
			91.1	20
METHAMPHETAMINE-d <sub>5</sub>	154.1	155.1	121.1	8
			92.1	20

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## **Declaration of Interest Statement**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.