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## ***In vitro* and *in vivo* MR evaluation of internal gradient to assess trabecular bone density**

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### **Abstract**

Here we propose a new magnetic resonance (MR) strategy based on the evaluation of internal gradient ( $G_i$ ) to assess the trabecular bone (TB) density in spongy bone. Spongy bone is a porous system characterized by a solid trabecular network immersed in bone marrow and characterized by a different relative percentage of water and fats. Using a 9.4 T MR micro-imaging system, we first evaluated the relative water and fat  $G_i$  as extracted from the Spin-Echo decay function *in vitro* of femoral head samples from calves. Indeed, the differential effects of fat and water diffusion result in different types of  $G_i$  behavior. Using a clinical MR 3T scanner, we then investigated *in vivo* the calcanei of individuals characterized by different known TB densities. We demonstrate, on these samples, that water is more prevalent in the boundary zone, while fats are rearranged primarily in the central zone of each pore. *In vitro* experiments showed that water  $G_i$  magnitude from the samples was directly proportional to their TB density. Similar behavior was also observed in the clinical measures. Conversely, fat  $G_i$  did not provide any information on spongy-bone density. Our results suggest that water  $G_i$  may be a reliable marker to assess the status of spongy bone.

### **1. Introduction**

Bone tissue is a complex biomaterial composed of a solid mineral matrix filled by bone marrow (the liquid interstitial phase). The solid matrix is composed mainly of mineral components, while bone marrow is composed mainly of hematopoietic marrow and fatty acid triglycerides at different relative percentages. The relative concentration of each bone marrow component is dependent on both, its anatomical skeletal location and the age of the individual (Liney *et al*

2007). Mammalian and human bone may be classified as cortical or trabecular. The former is mainly present in the shaft of long bones and is much denser (with a porosity ranging from 5% to 10%) than trabecular bone (TB). Conversely, TB, or spongy bone, is much more porous (with a porosity ranging from 50% to 90%) and is metabolically more active. From a physical point of view, spongy bone is a porous system that can be described as a solid with holes and cavities (i.e. presenting as connected void spaces randomly distributed within a solid matrix).

Osteoporosis is a highly diffuse disease that typically affects elderly individuals. It is a systemic skeletal disease characterized by low bone density, micro-architectural deterioration of the bone tissue and by an increase in bone porosity (Bousson *et al* 2000, 2004), leading to bone fragility and increased susceptibility to fracture (WHO Scientific Group Report 2000). From an epidemiological viewpoint, in elderly individuals the occurrence of bone fractures mainly affects those skeletal locations which are particularly rich in TB, such as the vertebrae and femoral head. The evaluation of bone mineral density (BMD) based on dual-energy x-ray absorptiometry (DXA) is currently considered the gold standard for clinical diagnosis of osteoporosis (Kanis and Gluer 2000). However, only a poor correlation between BMD assessments and the relative risk of bone fracture has been reported (Kanis 2002, Wehrli *et al* 2002a), suggesting that other factors besides low BMD likely contribute to determine bone fragility. This lack of information on the risk of bone fracture, which is critical for clinical purposes, has prompted intense research to identify new parameters with the ability to assess spongy-bone status and to provide reliable measures of bone's resistance. A new putative technique has to be first identified theoretically, then tested on *ex vivo* samples, and finally verified *in vivo*, to ascertain its potential utility for clinical purposes. Magnetic resonance (MR) is a non-invasive technique which is able to provide information on the relaxation and diffusion properties of water and fat protons from soft biological tissues such as bone marrow. Therefore, when applied to the investigation of spongy bone, MR provides information on bone marrow which is strictly linked to the surrounding trabecular architecture of the bone due to the mutual interaction between the solid matrix and liquid phase (Wehrli *et al* 2006). In this context, the apparent transverse relaxation time  $T_2^*$  was the first MR parameter to be considered as a new surrogate marker for osteoporosis. Indeed,  $T_2^*$  probes the micro-structure of TB due to its sensitivity to the microscopic field in-homogeneities caused by magnetic susceptibility differences between the solid bone structure and the liquid bone marrow (Majumdar *et al* 1991, Chung *et al* 1993). Several studies have consistently reported that an increase in inter-trabecular space, which typically occurs in patients with osteoporosis, prolongs the corresponding  $T_2^*$  values (Wehrli *et al* 1991, 2002a, Link *et al* 1998). However, the large standard deviation (SD) associated with average  $T_2^*$  values assessed in bone tissue from healthy, osteopenic and osteoporotic individuals does not allow the clinical use of  $T_2^*$  on a single subject basis (Capuani *et al* 2008).

In this paper we propose an alternative way to quantify the influence of susceptibility changes in spongy bone based on a new MR parameter (the internal gradient  $G_i$  that we demonstrate to be strictly associated with TB density and structural rearrangements). Estimation of  $G_i$  can be obtained by extraction of this parameter from the Spin-Echo (SE) decay, which is traditionally used for the investigation of porous systems (Watson and Chang 1997, Wilson and Hurlimann 2006, Kuntz *et al* 2008). In a SE sequence, signal refocusing is obtained by reversing the de-coherence effects of spins caused by the distribution of local fields. This method is valid only when molecular diffusion is not taken into account. Diffusion of liquid state protons in local magnetic field gradients (internal gradients  $G_i$ ) results in a more than expected rapid loss of coherence from the usual spin-spin interaction and surface effects (Watson and Chang 1997, Capuani *et al* 2005). In SE measurements, the additional decay of the echo amplitude is due to the diffusion of fluid molecules in the local magnetic field gradient

$G_i$  generated by the difference in magnetic susceptibility between bone marrow and solid bone plates. As a consequence, in systems like spongy bone, it is possible to assess the intensity of  $G_i$  from the attenuation of SE decay. In spongy bone,  $G_i$  values are not only affected by differences in magnetic susceptibility between bone marrow and solid bone plates, but also between fat and water diffusion in TB marrow. Moreover,  $G_i$  depends on the orientation of the trabecular plates with respect to the external magnetic field (Chung *et al* 1993, Hwang and Wehrli 1995). Some previous works have focused on these  $G_i$  features in order to assess the preferential orientation of trabeculae due to loading optimization (Chung *et al* 1993, Mertens *et al* 2008). Recently, a new MR approach based on the detection of spin diffusion through an internal magnetic field gradient has been proposed to characterize TB (Sigmund *et al* 2008, 2009). This approach is based on measuring the  $G_i$  to encode diffusion weighting to extract structural information (e.g., surface-to-volume ratio of spongy-bone pores) from the resulting signal decay.

In all these situations,  $G_i$  seems to be a potential surrogate marker reflecting different structural features of TB. They include not only the solid phase density, but also other characteristics related to spongy-bone quality (as expressed by water/fat concentrations within the bone marrow).

The aim of the present work was to assess the potential ability of  $G_i$  to describe spongy-bone status as related to its TB density and quality. For this purpose, using a micro-imaging probe we investigated *in vitro* at high magnetic field (9.4 T)  $G_i$  behavior inside each single pore of calf spongy-bone samples (extracted from a femur head) as a function of both TB density and relative fat and water–bone marrow concentrations. To test the methodological feasibility of this approach, we first quantified mean  $G_i$  in calf samples characterized by different TB densities. We then, investigated the potential suitability of  $G_i$  as a measure able to assess the TB status *in vivo*. For this purpose, using a 3T clinical scanner, we examined the calcanei of subjects with different TB densities.

## 2. Theory and methods

### 2.1. Theory

The signal decay of a SE sequence is usually described by the following equation (Hahn 1950):

$$S(TE) = S_0 \exp \left[ -\frac{TE}{T_2} \right], \quad (1)$$

where  $S_0$  is the signal intensity at  $TE = 0$ ,  $TE$  is the echo time and  $T_2$  is the transverse spin–spin relaxation time. However, in systems which are strongly influenced by the presence of internal magnetic field gradients  $G_i$ , the SE decay is no longer described as a simple exponential decay. In the linear gradient approximation, the signal decay can indeed be expressed by the following equation (Carr and Purcell 1954, Majumdar and Gore 1988):

$$S(TE) = S_0 \exp \left[ -\frac{TE}{T_2^{\text{true}}} - \frac{(\gamma G_i)^2 D (TE)^3}{12} \right], \quad (2)$$

where  $\gamma$  is the gyromagnetic ratio,  $D$  is the diffusion coefficient and  $T_2^{\text{true}}$  is the spin–spin relaxation time in which the effects of diffusion and field inhomogeneities are averaged out.

In systems like spongy bone, it is possible to assess the intensity of mean  $G_i$  from the attenuation of the SE signal as a function of different  $TE$ s. Furthermore, a measure of the diffusion coefficient  $D$  (i.e. a measure of the apparent diffusion coefficient (ADC)) and/or a measure of  $T_2^{\text{true}}$  are needed to reduce the number of unknown parameters and improve the fit

quality. In all *in vitro* and *in vivo* experiments reported in this paper, the ADC was measured and inserted in equation (2) to extract  $G_i$  and  $T_2^{\text{true}}$  using a fitting procedure.

## 2.2. *In vitro* experiments

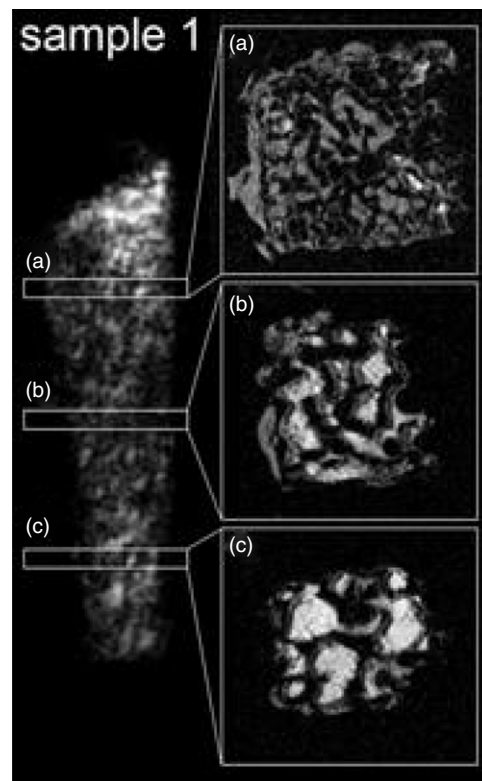
Experiments were performed using a Bruker Avance-400 high-resolution spectrometer operating at 9.4 T with a micro-imaging probe (10 mm internal diameter), equipped with a gradient unit characterized by a maximum gradient strength of  $1200 \text{ mT m}^{-1}$  and a rise time of  $100 \mu\text{s}$ . XWINNMR<sup>®</sup> and ParaVision<sup>®</sup> 3.0 software were employed for data acquisition and analysis.

Five *ex vivo* spongy-bone samples, excised from calf femur head, were cut into pieces of approximately 20 mm high and 7 mm deep in order to fit into the micro-imaging probe bore. The long axis of each sample was located parallel to the main direction of the static magnetic field (the  $z$ -axis). The temperature of each sample was fixed to 291 K.  $T_2^*$ -weighted (Gradient Echo, GE) and  $T_2$ -weighted (SE) images were obtained for initial evaluation of TB density. All samples were characterized by a non-uniform trabecular density. As shown in figure 1, TB density varies continuously along the height of each sample, moving from the upper to the lower zone. The upper zone, which is close to the cortical bone, is characterized by higher trabecular density and smaller spongy-bone pores (figure 1(a)). Conversely, the lower zone is characterized by a larger inter-trabecular space (figure 1(c)). The middle zone (located between upper and lower zones) shows intermediate characteristics of TB density (figure 1(b)). Throughout the paper, we will refer to these three anatomical locations as follows: (a) HTD (high trabecular density; upper zone), (b) ITD (intermediate trabecular density; middle zone) and (c) LTD (lower trabecular density; lower zone). In each sample, for each considered location, we identified three 0.2 mm thickness slices to measure quantitative MR parameters. Data extracted from each of the three slices were then averaged to obtain mean values of each MR parameter in the three locations (HTD, ITD and LTD). Five analyzed samples were selected such that they were characterized by similar TB densities and similar fat-to-water ratio quantities in bone marrow. TB density and bone marrow water content (together with the relative content of different types of fat in bone marrow) are known to be dependent on age, race and skeletal site. To correct for these potential biases, we obtained all our samples from 10- to 12-month-old calves of German race, bred in Germany and butchered in Rome (Italy). In all fresh samples (analyzed immediately after slaughter) we identified the same area of the femur head for MR investigation.

**2.2.1. Spectroscopic experiments.** In the first step of our experiments, a validation experiment was performed under ideal circumstances where molecular diffusion and  $T_2$  information were obtained for each of the two individual spectral components (water and fat molecules).

A spectroscopic Carr–Purcell–Meiboom–Gill (CPMG, repetition time  $TR = 1 \text{ s}$ , number of averaged scans  $NS = 8$ ,  $N = 64$  data points, (corresponding to 64 echoes refocusing every 2 ms from 1 to 127 ms) and a spectroscopic pulsed field gradient stimulated echo (PGSE,  $TE/TR = 18/3000 \text{ ms}$ , diffusion gradient pulses delay  $\Delta = 400 \text{ ms}$ , diffusion gradient pulses duration  $\delta = 4.6 \text{ ms}$  and diffusion gradient strength  $g$  applied along the  $x$ -axis using 32 gradient amplitude steps from 6 to  $100 \text{ mT m}^{-1}$ ) were used to recognize and discriminate  $T_2$  and ADC of water and fat molecules in the samples.

**2.2.2. Spatially resolved experiments.** A MSME (multi-slice multi-echo) imaging sequence ( $TR = 2000 \text{ ms}$ , field of view  $FOV = 7 \text{ mm}$ , matrix  $256 \times 256$ , slice thickness,  $STH = 0.2 \text{ mm}$ , pixel dimensions  $27 \times 27 \mu\text{m}^2$ ,  $NS = 8$ ) at various  $TE$ s (4.8, 6, 8, 10, 12, 14, 16, 18, 20, 24,



**Figure 1.** An example of the calf spongy-bone sample used for experiments *in vitro*. GE localizer and SE images ( $TE = 4.3$  ms) selected perpendicularly to the static magnetic field at three different depths in the sample show a non-uniform TB density. In each experimental sample, trabeculae are closely spaced in the upper zone (a). Conversely, they show an intermediate density in the middle zone (b) and a larger inter-trabecular space in the lower zone (c).

30, 40, 50, 60, 70, 80, 100, 120 ms) was used to obtain the SE decay in the three regions of each sample characterized by a different TB density. A PGSTE imaging sequence was also employed ( $TE/TR = 21.9/3000$  ms,  $FOV = 7$  mm, matrix  $256 \times 256$ ,  $\Delta = 40$  ms,  $\delta = 4$  ms, using eight  $b$  values ranging from 400 to 40 000  $\text{s mm}^{-2}$ ,  $STH = 0.2$  mm,  $NS = 8$ ) in order to measure the ADC along the  $x$ -axis for each of the three selected regions of the considered samples. The  $b$  value is related to the aforementioned parameters through the known relation:  $b = \gamma^2 g^2 (\Delta - \delta/3)$ , where  $g$  is the diffusion gradient strength. The  $x$ -axis was arbitrarily chosen to assess molecular diffusion perpendicular to trabeculae surfaces (see figure 1).

After the micro-imaging investigation of the bone marrow's water and fat behavior within pores of spongy bone, we set up an equivalent acquisition protocol at lower  $TE$ s (4.8, 10, 20, 40, 60, 80, 100 ms) and  $b$  values (ranging from 200 to 10 000  $\text{s mm}^{-2}$ ). This additional protocol was added to obtain a protocol potentially suitable for clinical application (i.e. lower scan time, lower  $b$  values). However, due to a reduced number of experimental points, this acquisition provides SE decays and ADCs that do not clearly discriminate between water and fat.

### 2.3. Data processing and statistical analysis

$T_2$  and ADC of water and fat components were obtained using a Levenberg–Marquardt (L–M) fit taking into account the peak area decays respectively centered at 4.7 ppm (large from 4 to 5.4 ppm) for water and at 1.3 ppm (large from 0.1 to 2.5 ppm) for fat. Due to the low resolution of bone marrow spectra in the spongy-bone samples, the olefinic fat peak at 5.3 ppm overlaps the broad water peak centered at 4.7 ppm. For this reason, we fitted the attenuation of this broad peak (from 6 to 4 ppm) versus the diffusion gradient strength or the  $TE$  value using a bi-exponential function. This took into account the olefinic and water component separately. In figure 2 an example of spectroscopic investigation is shown.

Two different *in vitro* analyzes were performed using the data extracted from the imaging experiments. In both cases we considered only the central zone of each image (i.e. excluding the image border), in order to exclude or limit the presence of air in bone marrow cavities. Indeed, this represents one of the major sources of artifacts when performing imaging on excised samples.

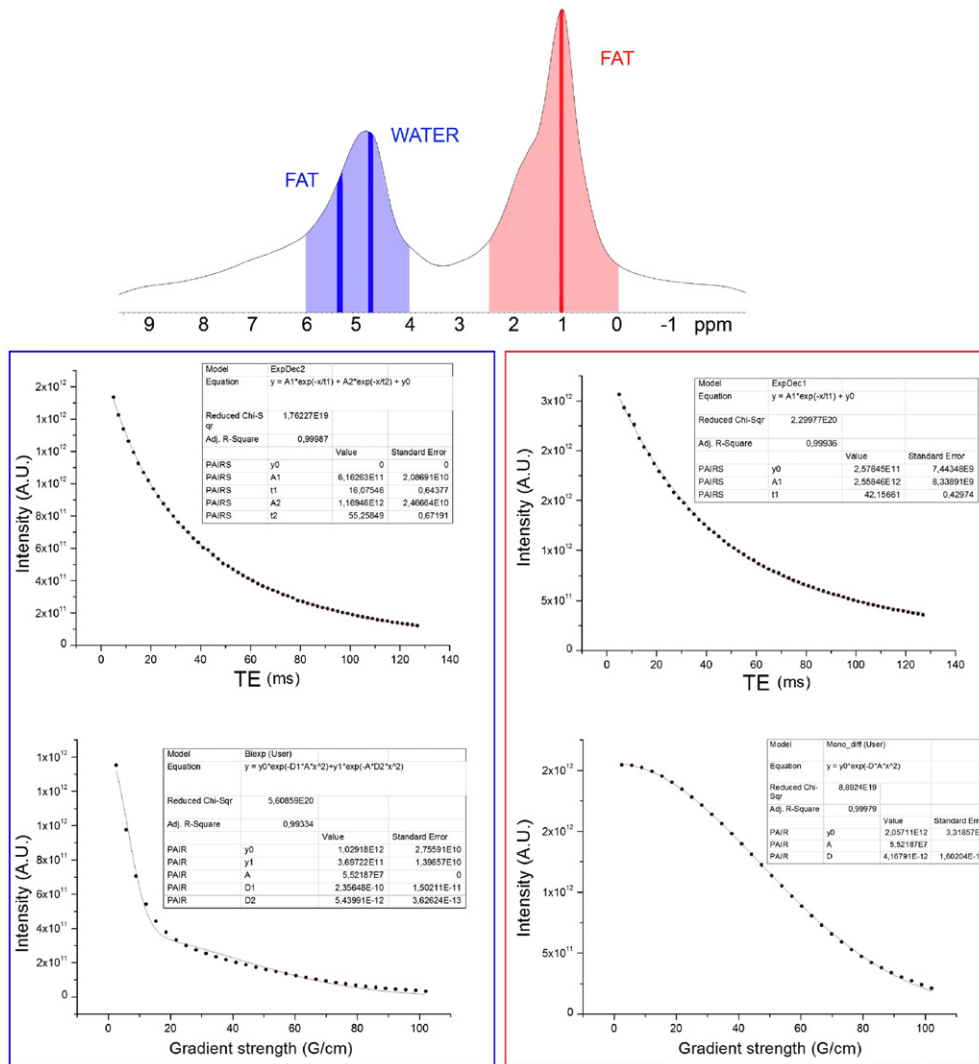
In the first analysis, the signal deriving from water and fat protons in  $T_2$ -weighted images was investigated as a function of each voxel location with respect to the bone–bone marrow interface. In each slice of the three considered locations, the signal nearby the boundaries between bone marrow and solid bone was identified using a software (written in Matlab) (figure 3) written in-house.

SE signal properties and  $G_i$  behavior were then assessed in the boundary and inner regions of the trabecular pores. The inner regions of the pores were identified by subtracting the boundary regions from the whole pore area. An L–M fit was performed using the signal as a superimposition of two components: one belonging to fats and another belonging to water molecules. Each component contributes to the total signal as one exponential term of formula (2). The signal intensity at  $TE = 0$ ,  $T_2^{\text{true}}$ ,  $G_i$  and the ADC of water and of fat components were obtained. Using this procedure, mean and SD values of  $G_i$  from water and fat were respectively derived. Water ADC and fat ADC were measured from diffusion-weighted images, while  $T_2^{\text{true}}$  values were considered a free parameter in the fit.

Conventional  $T_2$  relaxation times of water and fat components were also evaluated from the same images, executing a bi-exponential L–M fit of water and fat components, in accordance with formula (1). This analysis was repeated at different widths (measured in voxels) from the boundary region of each pore. The mean values of water and fat  $G_i$  were calculated by averaging the results obtained in all pores contained in three image slices for each specific location (HTD, ITD and LTD).

In order to have a more objective quantification of TB density, we calculated the ratio between the perimeter and the area of each pore in the slices. As a result, the ratio ( $N_p/N_a$ ) between the number of voxels defining each pore perimeter ( $N_p$ ) and the number of voxels constituting the corresponding area ( $N_a$ ) was obtained from all pores included in each of the slices. Then, the mean values of  $N_p/N_a$  and  $G_i$  in different bone locations (characterized by different TB densities as defined by  $N_p/N_a$ ) were obtained from each sample. Moreover, mean values of both magnetization associated with water and fat extracted from the bi-exponential fit as a function of the boundary region width were evaluated. The averages were calculated in all five samples. Pearson's correlation coefficient ( $r$ ) and a paired Student's  $t$ -test were used to assess the linear correlation between  $G_i$  and TB densities and differences between water and fat magnetization as a function of boundary distances measured in voxels.

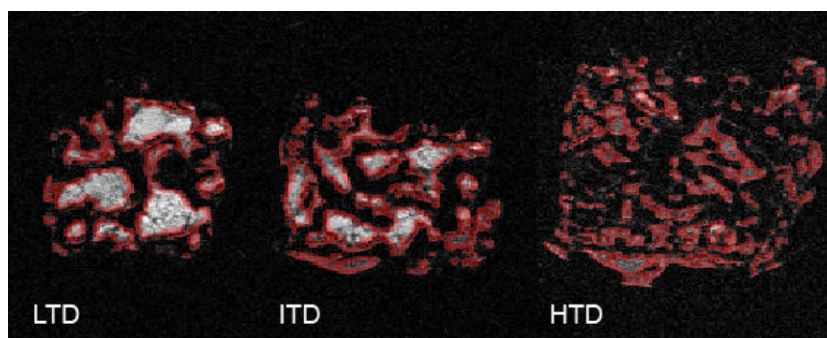
In a second experiment,  $G_i$  was derived using the signal arising from each entire slice. In this case, we aimed to identify an acquisition protocol at high magnetic field which might also be suitable for clinical application (using relatively short acquisition times,



**Figure 2.** An example of the fitting procedure used in spectroscopic validation experiments to obtain  $T_2$  and ADC of water and fat components. The broad peak on the left side of the spectrum is characterized by two different components: water centered at 4.7 ppm and olefinic fat centered at 5.3 ppm. Peak attenuation versus gradient strength or  $TE$  value was fitted using a bi-exponential function which took into account olefinic and water components separately. Conversely, the attenuation of fat peak at 1.3 ppm (on the right side of the spectrum) was fitted using monoexponential functions. See the text for further details.

conventional sequences and a lower image resolution compared to that obtained by micro-imaging apparatus). Distinguishing water from fat components was not a priority in this case. Therefore, a monoexponential L–M fit was employed to obtain the mean ADC along the  $x$ -axis. A monoexponential L–M fit was performed using formula (2) and included a constant which took into account the deviation from monoexponential behavior. Mean and SD values of  $G_i$  were obtained in the three bone locations (HTD, ITD and LTD) performing an average of





**Figure 3.** Example of selection of a three pixel wide boundary region in three considered locations, respectively, characterized by lower (LTD), intermediate (ITD), and higher trabecular density (HTD). The red mask, which represents the boundary region, is superimposed over SE images obtained at  $TE = 4.3$  ms.

$G_i$  values across the five samples together with their corresponding values of  $T_2^{\text{true}}$ . Moreover, conventional  $T_2$  relaxation time was estimated from each slice using a monoexponential L–M fit as described by formula (1) plus a constant. Again, mean and SD values of  $T_2$  were obtained in the three zones (HTD, ITD and LTD) by performing an average of  $T_2$  values across the five samples.

#### 2.4. *In vivo* experiments

Six subjects were recruited for this study, three healthy females (24, 42, 52 years old, respectively), one osteopenic and one osteoporotic female (both subjects 62 years old), and one male (33 years old).

Healthy, osteopenic or osteoporotic status of the 52- and 62-year-old females was established on the basis of their last (within 6 months from our MR investigation) DXA medical report. The mean  $T$ -score in L1 to L4 vertebrae of the 52- and of the two 62-year-old females were  $-1.30$ ,  $-1.81$  and  $-3.51$ , respectively.

The study was approved by the Local Ethics Committee. Each subject gave her/his informed written consent to participate in this study.

A progressive reduction of TB density across aging has been well described in the previous literature (Wehrli *et al* 1995, Thomsen *et al* 2002). In humans, bone mass reaches a maximum peak at the approximate age of 25 years, and then progressively decreases over the following decades. As a consequence, it is reasonable to assume that both the 42- and the 52-year-old females had a lower TB density when compared to the 24-year-old female. Gender-related differences are also well reported (Mosekilde 2000). Men are known to have a higher bone mass peak than age-matched women, thus suggesting that our male subject had the highest TB density. Moreover, the osteoporotic female ( $T$ -score =  $-3.51$ ) included in this study had a lower TB density than the age matched osteopenic ( $T$ -score =  $-1.81$ ) female. As expected, both 62-year-old females (with osteopenia and osteoporosis) had a lower TB density than the 52-year-old healthy woman ( $T$ -score =  $-1.30$ ).

Each subject underwent an MRI examination of the right foot (figure 4), using a 3T MR system (Siemens Allegra, Erlangen, Germany). A FLASH (Fast Low-Angle SHot) sequence ( $TR = 1500$  ms, field of view FOV = 192 mm, matrix  $128 \times 128$ , slice thickness 5 mm, flip angle  $30^\circ$ ) was acquired using various  $TE$ s (5, 7, 10, 20 ms) to estimate  $T_2^*$ . Sagittal



**Figure 4.** MR sagittal view of the foot examined in a 24-year-old female. The region outlined in black shows the calcaneal area on the SE image used to assess the mean value of  $G_i$  after fitting based on the L–M procedure.

SE images ( $TR = 1500$  ms,  $FOV = 192$  mm, matrix  $256 \times 256$ , slice thickness 5 mm) were also collected using a SEMC (spin echo multi contrast) sequence at various  $TE$ s (20, 30, 40, 50, 80, 100 ms). The ADC was evaluated from sagittal diffusion-weighted images using a SE segmented-EPI (echo planar image) sequence at two different  $b$  values (0 and  $8000 \text{ s mm}^{-2}$ ) with diffusion gradient applied along the anterior–posterior direction and  $TE/TR = 89/2500$  ms.  $G_i$  was obtained by the same mono-exponential L–M fitting procedure (using equation (2) without discriminating between water and fat components) described in the previous section by selecting the entire calcaneal area of each subject as shown in figure 3. Single voxel spectroscopy (PRESS sequence) with  $TE = 22$  ms,  $TR = 5$  s,  $NS = 32$ , was used to obtain bone marrow proton spectra to extract fat and water content percentage. A single voxel (size of  $15 \times 15 \times 15 \text{ mm}^3$ ) was positioned in the center of the calcaneus of each subject.

### 3. Results and discussion

#### 3.1. *In vitro* results

Spectroscopic  $T_2$  and ADC mean values and their SD of water and fat components in excised bone samples are reported in table 1. Data show that both  $T_2$  and ADC of the water component (centered at 4.7 ppm) are significantly lower than those of fat components (centered at 5.3 ppm and 1.3 ppm). This allows us to discriminate clearly between the two components (water and fat). Specifically,  $T_2$  values of water and fat are respectively consistent with those previously reported (Chabanova *et al* 2006). Similarly, ADCs of fat components, which are approximately two order of magnitude lower than those of water molecules, fit well with those recently reported by other authors (Ong *et al* 2009).

In table 2, mean values (and their SD) of  $T_2$ , ADC,  $G_i$  and  $T_2^{\text{true}}$  are reported for the water component obtained in LTD, ITD and HTD locations of one sample (performing an average of

**Table 1.** Spectroscopic evaluation of excised calf samples.

	$(T_2 \pm \text{SD})$ (ms)	$(\text{ADC} \pm \text{SD})$	
H <sub>2</sub> O at 4.7 ppm	19 ± 7	4.2 ± 1.8	10 <sup>-10</sup> m <sup>2</sup> s <sup>-1</sup>
Fats at 5.3 ppm	72 ± 36	5.0 ± 0.6	10 <sup>-12</sup> m <sup>2</sup> s <sup>-1</sup>
Fats at 1.3 ppm	65 ± 16	5.3 ± 0.3	10 <sup>-12</sup> m <sup>2</sup> s <sup>-1</sup>

**Table 2.** Results from one excised calf sample: (a) water component and (b) fat component.

(a) Water component					
	Water (%)	$(T_2 \pm \text{SD})$ (ms)	$(\text{ADC} \pm \text{SD})$ ( $\times 10^{-10}$ m <sup>2</sup> s <sup>-1</sup> )	$(G_i \pm \text{SD})$ (mT m <sup>-1</sup> )	$(T_2^{\text{true}} \pm \text{SD})$ (ms)
LTD	55	17.0 ± 3.0	19.0 ± 0.4	222 ± 131	17.9 ± 1.0
ITD	47	15.0 ± 2.3	7.0 ± 0.4	282 ± 134	15.2 ± 0.5
HTD	97	14.8 ± 0.4	1.6 ± 0.1	803 ± 181	16.0 ± 0.3
(b) Fat component					
	Fat (%)	$(T_2 \pm \text{SD})$ (ms)	$(\text{ADC} \pm \text{SD})$ ( $\times 10^{-12}$ m <sup>2</sup> s <sup>-1</sup> )	$(G_i \pm \text{SD})$ (mT m <sup>-1</sup> )	$(T_2^{\text{true}} \pm \text{SD})$ (ms)
LTD	45	42.3 ± 6.2	5.3 ± 0.3	760 ± 195	43.0 ± 4.8
ITD	53	39.7 ± 4.0	5.9 ± 0.3	727 ± 437	44.0 ± 5.3
HTD	3	31.6 ± 6.0	5.4 ± 0.2	904 ± 119	37.5 ± 9.8

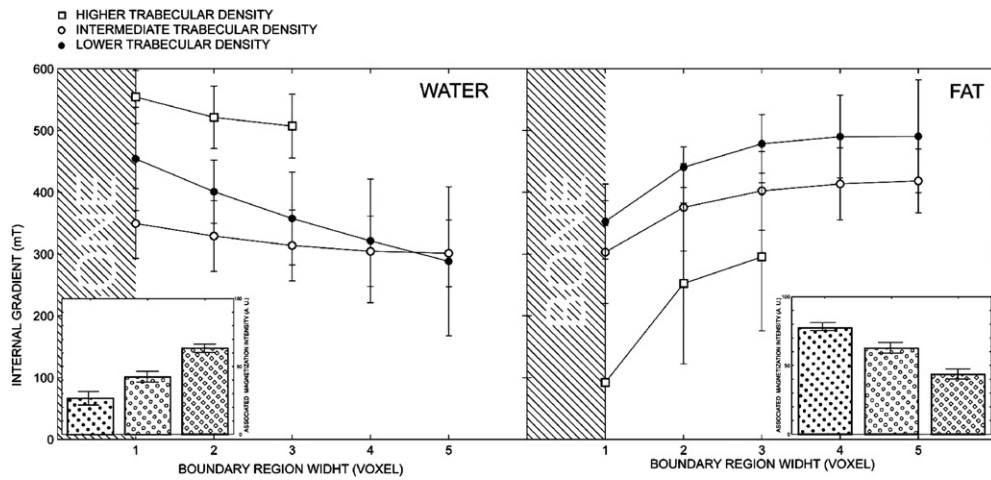
values extracted from each slice belonging to a single location). In table 2, the corresponding results for the fat component are reported.

The water ADC showed a decreasing trend proportional to the increase of trabecular density; thus, water mobility is more restricted in regions where the trabeculae are smaller. Conversely, it is evident from table 2 that the fat ADC is not sensitive to increase in trabecular density. Water  $G_i$  increases proportionally with the increase in TB density, while fat  $G_i$  is characterized by an opposite trend and does not allow discrimination between LTD and ITD.

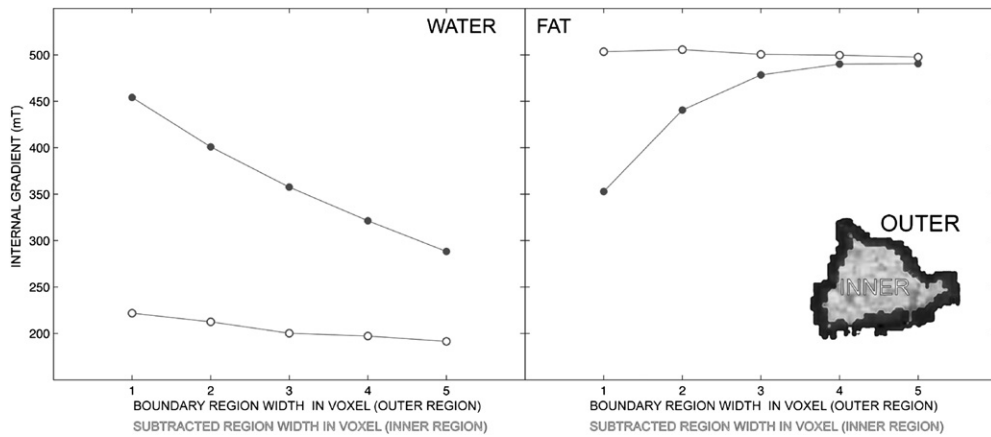
Figure 5 displays different types of behavior of water and fat  $G_i$  as a function of the distance (in voxels) between the TB marrow interface and the internal part of the trabecular pore.  $G_i$  values were derived using a fitting procedure, while ADC values were derived from diffusion-weighted images.

According to our findings, water  $G_i$  is characterized by a decreasing trend when moving from zones adjacent to the trabecula to zones located in the center of the inter-trabecular space. This behavior was observed in each of the three locations characterized by different TB densities. When comparing  $G_i$  magnitude between the three locations, it was otherwise evident that  $G_i$  in the HTD location is higher than that extracted from the other two locations (ITD and LTD). Conversely, fat compared to water  $G_i$  revealed an opposite behavior. In the latter case,  $G_i$  was characterized by an increasing trend when moving from voxels adjacent to trabeculae to those located in the center of the inter-trabecular space. In the insets of figure 5, mean magnetization intensities are reported for fat and water components as derived from the fitting procedure. Water concentration is higher in the HTD slices and decreases proportionally with the reduction of trabecular density (i.e. in ITD and in LTD).

ADC values and their magnetic susceptibility are the key parameters accounting for the different types of behavior of water and fat  $G_i$  (figures 5 and 6). The water ADC is



**Figure 5.** Behavior of water (left) and fat  $G_i$  (right) as a function of increasing voxel number ranging from regions with one pixel (located close to the bone surface) to regions constituted by five pixels (from the bone surface to the pore center). Results are reported for HTD (empty squares), ITD (empty circles) and LTD (filled circles). In regions with HTD, the last two points are missing because the trabecular interspaces were not large enough to allow a boundary region (which is wider than three pixels) to be drawn.



**Figure 6.** Water  $G_i$  (left) and fat  $G_i$  (right) behavior for LTD in the boundary region (filled circles) and in the inner region (empty circles) as a function of increasing boundary region width (which corresponds to a decrease of the inner region width). The size of each pixel is  $27 \mu\text{m}$ .

approximately two orders of magnitude higher than the fat ADC. As a consequence, water molecules compared to fat molecules are characterized by a faster motion. According to Einstein's relation  $\langle r \rangle^2 = 2Dt$ , which links the mean square spatial range spanned by diffusing molecules (characterized by a certain diffusion coefficient  $D$  at a diffusion time  $t$ ), water spins travel a distance of the order of tenths of microns (calculated using the diffusion times  $\Delta$  employed in this study). Conversely, the displacement of protons in fat lies in the sub-micron range. Moreover, water and fat are characterized by different magnetic susceptibilities:

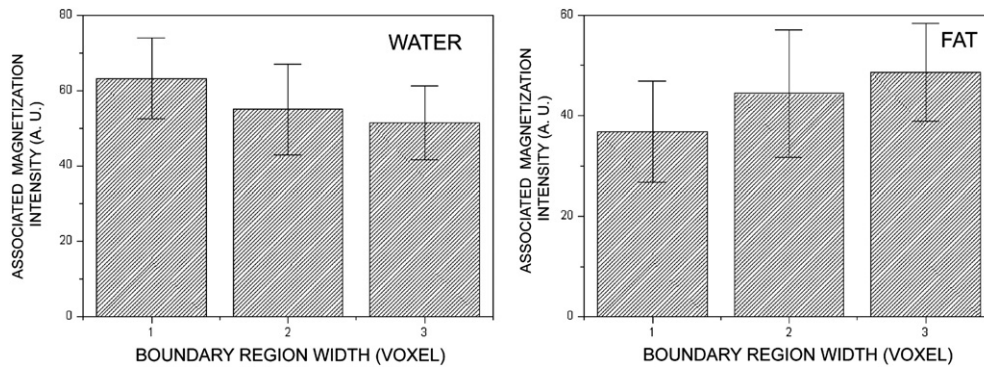
$\chi_{\text{water}} = -9.05 \times 10^{-6}$  and  $\chi_{\text{fat}} = -8.44 \times 10^{-6}$  (SI units) (Szczepaniak *et al* 2002), while bone tissue is believed to be characterized by  $\chi_{\text{bone}} = -11 \times 10^{-6}$  (Hwang and Wehrli 1999).

The  $G_i$  belonging to protons in water is sensitive to differences in magnetic susceptibility in the range of the pixel dimension. As expected,  $G_i$  shows a decreasing trend when moving from the bone–bone marrow interface to the pore center. Indeed, the difference in water–bone susceptibility represents the main source of water  $G_i$ . Conversely, protons in fat are sensitive to differences in magnetic susceptibility ranging at least from two orders of magnitude less than the pixel size. For this reason, fat  $G_i$  measured in this study cannot be attributed to the bone–bone marrow interface. Fat  $G_i$  magnitude is better explained by the difference in magnetic susceptibility between fat and water. The increasing trend of fat  $G_i$  values (when moving from the bone–bone marrow interface to the pore center) can thus be ascribed to a different rearrangement of fat molecules. In the central zone of each pore, there is a higher concentration of fat molecules characterized by a lower molecular motion than that of water molecules. As a consequence, fat  $G_i$  in the center of the pore assumes higher values than those observed nearby the bone surface. Indeed, in this latter location, there is a reduced concentration of fat molecules. This means that a correspondingly higher concentration of water molecules (characterized by faster motion) modulates fat  $G_i$  by reducing its values. This scenario fits well with the hypothesized distribution of bone marrow filling trabecular pores, for which we could say that ‘water wets’ the surface of bone pores. On the other hand, fat molecules, due to their hydrophobic nature, are mainly rearranged to lie in the central zone of the pore. As shown in figure 6, water and fat  $G_i$  behavior in the boundary and inner regions of LTD pores support this hypothesis. In the figure, water and fat  $G_i$  behavior (expressed as a function of increasing pixel width) of the boundary region are compared to those measured from the inner region.  $G_i$  values of both water and fat bone marrow components move toward an asymptotic value which is very close to that obtained in the central zone of the trabecular pore.

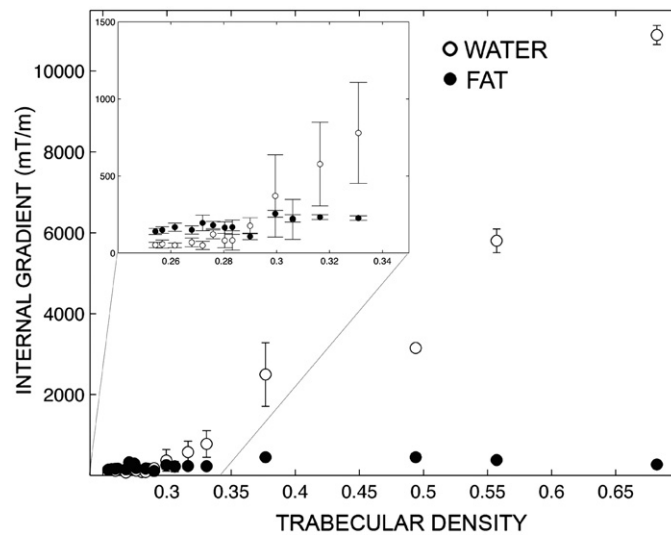
This observation confirms that water  $G_i$  variation is mainly due to bone–bone marrow interface susceptibility properties. Therefore, water  $G_i$  might be considered as a surrogate marker of the TB structure. The asymptotic  $G_i$  values reached by the fat component were higher when compared to those reached by the water component. Again, the difference in the diffusion coefficient between fat and water might explain these results. As water molecules, compared to fat molecules, are associated with a faster diffusive motion during the experimental diffusion time, water spins are more dependent on motional averaging. As a consequence, the susceptibility dephasing effect which affects the SE signal of water is partially averaged. This peculiar behavior of water/fat systems has already been investigated and verified elsewhere (Lammentausta *et al* 2008). To support the hypothesis that water concentration is higher in the boundary zone of pores, while fat is more concentrated in the central zone of pores, mean magnetization, extracted from the fit and associated with water (left) and fat (right) quantities in the first three voxels of the boundary regions, is reported in figure 7. Mean magnetization values and their corresponding SD values were obtained from all five samples belonging to different calves. Mean magnetization values associated with each voxel’s zone of the boundary region are statistically different ( $p = 0.05$ ).

Data reported in figure 7 demonstrate that in each pore of spongy bone, the quantities of water molecules at the bone–bone marrow interface are higher than that of fat molecules. As a consequence, in spongy bone (filled with bone marrow), the measurements of water MR parameters (such as  $G_i$ ,  $T_2^*$  or ADC) best provide information on TB density.

In figure 8 mean  $G_i$  values and their SD, obtained from the five samples as a function of their TB density, are reported. The graph in the inset of figure 8 shows a positive linear correlation between water  $G_i$  and TB densities (Pearson’s correlation coefficient  $r = 0.71$ ).



**Figure 7.** Mean magnetization values arising from the bi-exponential fit associated with water (left) and fat (right) versus boundary region width, expressed as a distance in voxels from the bone/marrow boundary. The average is performed across all samples. LTD location was considered in all experimental samples.



**Figure 8.** Water  $G_i$  behavior (empty circles) and fat  $G_i$  behavior (filled circles) as a function of trabecular density obtained as the ratio between the number of voxels belonging to the boundary perimeter ( $N_p$ ) and the number of voxels of the whole area ( $N_a$ ). The mean  $G_i$  values were calculated in a boundary region of three voxels wide. In the figure inset a zoom is displayed for TB densities ranging from 0.25 to 0.34.

The large SD associated with water  $G_i$  mean values is mainly due to differences in water content between the five samples. The strong variation of water ADC is likely due to the small differences in water–fat content percentages across samples. The larger the ADC SDs, the larger are the SDs of  $G_i$ . Conversely, the fat ADC changes less as a function of water–fat percentage variation.

Finally,  $T_2$ , ADC,  $G_i$  and  $T_2^{\text{true}}$  mean values and their SDs obtained from the entire slice area from each of the selected locations (averaging on the five samples) are reported in table 3.  $G_i$  and  $T_2^{\text{true}}$  were extracted from the fit by function (2), while  $T_2$  and ADC represent values

**Table 3.** Results obtained from excised calf samples without discriminating between water and fat.

	$(T_2 \pm \text{SD})$ (ms)	$(\text{ADC} \pm \text{SD})$ ( $\times 10^{-10} \text{ m}^2 \text{ s}^{-1}$ )	$(G_i \pm \text{SD})$ ( $\text{mT m}^{-1}$ )	$(T_2^{\text{true}} \pm \text{SD})$ (ms)
LTD	$22.6 \pm 9.7$	$4.0 \pm 0.7$	$263 \pm 68$	$30.7 \pm 3.3$
ITD	$22.0 \pm 5.7$	$3.8 \pm 1.2$	$411 \pm 102$	$28.1 \pm 2.9$
HTD	$8.7 \pm 2.8$	$5.3 \pm 1.6$	$675 \pm 183$	$8.3 \pm 1.2$

**Table 4.** Calcaneus volunteer results.

Subject			Fat	$(T_2^* \pm \text{SD})$	$(T_2 \pm \text{SD})$	$(\text{ADC} \pm \text{SD})$	$(G_i \pm \text{SD})$	$(T_2^{\text{true}} \pm \text{SD})$
Sex	Age	Status	(%)	(ms)	(ms)	( $\times 10^{-11} \text{ m}^2 \text{ s}^{-1}$ )	( $\text{mT m}^{-1}$ )	(ms)
M	33	H	93	$12.5 \pm 3.1$	$34.6 \pm 6.0$	$4.5 \pm 0.9$	$894 \pm 113$	$55.3 \pm 7.1$
F	24	H	87	$9.8 \pm 1.7$	$41.0 \pm 3.7$	$4.3 \pm 0.5$	$477 \pm 40$	$41.3 \pm 1.4$
F	42	H	87	$12.7 \pm 2.3$	$35.3 \pm 2.8$	$3.7 \pm 0.7$	$415 \pm 56$	$39.5 \pm 1.9$
F	52	H	88	$9.5 \pm 1.8$	$41.2 \pm 3.2$	$5.0 \pm 0.5$	$399 \pm 44$	$40.6 \pm 1.1$
F	62	OPE	87	$15.1 \pm 2.3$	$42.0 \pm 2.7$	$7.3 \pm 0.5$	$311 \pm 29$	$41.3 \pm 0.9$
F	62	OPO	90	$21.1 \pm 2.5$	$38.0 \pm 2.5$	$6.6 \pm 0.6$	$240 \pm 47$	$47.0 \pm 2.3$

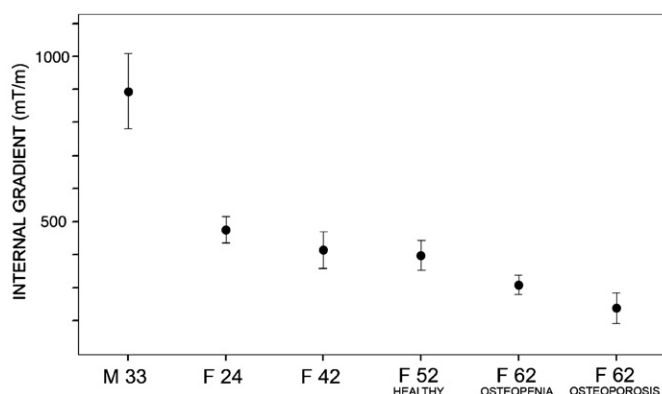
obtained by direct measures. Results reported in table 3 are from an additional analysis in which we did not discriminate between fat and water components. This means that both components contribute, with different modalities, to the total fitted signal. However, the acquisition parameters chosen for this experiment were intentionally optimized to focus on the water signal (for example,  $b$  values from 0 to  $10\,000 \text{ s mm}^{-2}$  were chosen to detect the typical water ADC but not the fat ADC). This means that there is only a minimal weight of the fat component in the determined results reported in table 3.

$T_2$  and ADC values did not discriminate between LTD, ITD and HTD. Conversely,  $T_2^{\text{true}}$  was characterized by a decreasing trend when moving from LTD to HTD, while  $G_i$  showed different values in the three locations with different TB densities. In bone marrow,  $T_2$  is mainly dependent on the relative proportion of fat and water protons (which are characterized by different transverse relaxation times), as well as from the TB density. The ADC depends on the interstitial spaces between bone and fat where water diffuses.  $G_i$  assumes a higher value for higher trabecular densities, as expected by water  $G_i$  values reported in table 3, which are due to differences in magnetic susceptibility at the bone–water interface.

Results reported in table 3 represent a preliminary test at high field to investigate the feasibility of including  $G_i$  measurements within clinical protocols (i.e. collection of  $T_2$ -weighted images and ADC maps).  $G_i$  would add useful information by discriminating between different TB densities.

### 3.2. In vivo results

Mean  $T_2^*$ ,  $T_2$ , ADC,  $G_i$  and  $T_2^{\text{true}}$  with their corresponding SD values are summarized in table 4, together with the bone marrow fat content percentage extracted from the calcanei of six subjects. These findings indicate that  $T_2^*$ ,  $T_2$  and ADC do not discriminate between the



**Figure 9.** Clinical measures of  $G_i$  obtained from human calcanei as a function of individual 'supposed' TB densities. See the text for further details.

six different subjects presenting with different TB densities, while  $T_2^*$  discriminates between a healthy female of 52 years, and subjects with osteopenia and osteoporosis. Conversely,  $G_i$  discriminates between calcanei with different TB densities as shown in figure 9. Moreover,  $T_2^{\text{true}}$  values seem to be more associated with fat percentage than with TB densities.

According to the previous literature, the only male included in the current study showed a higher fat fraction when compared to females (Kugel *et al* 2001). The two youngest females (24 and 42 years old respectively) and the subject with osteopenia, showed a similar fat bone marrow percentage content with no evident differences related to aging.  $T_2^*$  was lower in the youngest female, characterized by a higher TB density when compared with the 42-year-old female. However, since  $T_2^*$  depends on both TB density and fat bone marrow concentrations, the male subject showed a higher  $T_2^*$ . Moreover, younger subjects were characterized by  $T_2^*$  values equal to those from a 52-year-old healthy subject characterized by a higher fat percentage value when compared to the 24-year-old female. Conversely,  $G_i$  values were remarkably different among subjects with different TB densities, independently from their bone marrow fat content.

The relationship between ADC *in vivo* and age/gender is less clear: a decreasing ADC for reduced BMD was reported in the vertebrae (Yeung *et al* 2004, Hatipoglu *et al* 2007), while other works reported a lack of correlation between diffusion indexes and BMD (Griffith *et al* 2006). The presence or absence of this relationship seems, therefore, to be related to different anatomical locations. Since the fat content in the calcaneus is higher than that observed in the spine (Liney *et al* 2007), the two sites are likely to be affected by BMD modifications in a different way. In the specific case of calcaneus spongy bone, restricted diffusion experienced by water confined between fat and bone could be hypothesized, as suggested by the ADC values reported in table 4, which are of the order of  $10^{-11} \text{ m}^2 \text{ s}^{-1}$ . Moreover, a lower restricted diffusion due to increasing age or decreasing TB density could also be expected. However, it is not possible to conclude any general trend for all parameters listed in table 4 due to the small sample size.

Several studies reported an increase of  $T_2^*$  values for decreasing trabecular densities. However, the high inter-subject variability observed across studies is likely to be due to differences in water/fat ratios which affect  $T_2^*$  decay. Remarkably, higher values of  $T_2^*$  have been reported for fat as compared to the water component, either *in vitro* (Hussain *et al* 2005, O'Regan *et al* 2008) or *in vivo* (Weis *et al* 2008). Furthermore, several kinds of



fat molecules are present in bone tissue (Ren *et al* 2008), with relative concentration again varying across subjects. In table 4,  $T_2^*$  values of healthy subjects are very close to each other and overlap when considering their SDs. Moreover, the large  $T_2^*$  value reported for the male subject is more likely to be linked to a higher fat concentration rather than to a reduced TB density, which is not supported by any evidence from young healthy men. According to our experimental results,  $T_2^*$  turned out to be affected by the water/fat ratio, a parameter that shows a huge spread across different subjects. Some authors have suggested introducing corrections to account for variations in marrow composition (Wehrli *et al* 2000). However, to our knowledge, no-clinical studies involving these corrections have been published so far.

TB density is believed to be higher in male than in female subjects and to be lower in young than in elderly females. Moreover, TB density is lower in osteopenic subjects than in healthy subjects with the lowest values in females with osteoporosis. In our experiments,  $G_i$  has shown the ability to clearly discriminate between six subjects with different ages and different TB densities. Furthermore,  $G_i$  does not seem to suffer from the aforementioned limitation, which does not currently allow the clinical use of MR parameters on a single subject basis for diagnostic purposes. Clearly, our discussion about the potential clinical use of  $T_2^*$  and  $G_i$  parameters requires caution due to the small sample size recruited for the current study. In figure 9,  $G_i$  values presented are obtained from the central slice of the calcanei in six subjects, who presumably had different TB densities as expected in individuals with different sex, age and  $T$ -score values.

These results illustrate a smooth decrease of  $G_i$  values in the five females from the youngest to the oldest (linear correlation coefficient  $r = 0.81$ ). Considering that age is a well-known factor affecting TB density, we can conclude that our preliminary data obtained *in vivo* demonstrate a progressive reduction of  $G_i$  across aging. With this perspective, we can speculate that  $G_i$  decreases proportionally with the physiological reduction of TB density. Future studies, including both large populations of healthy subjects and subjects with osteopenia and osteoporosis, are needed to clarify the potential diagnostic role of  $G_i$ . Moreover, it should be noted that our results, obtained *in vivo* by conventional  $T_2$  and ADC measurement performed on a clinical scanner, fit well with those obtained *in vitro* (see figure 8).

The great potential of  $G_i$  as a novel marker for *in vivo* assessment of TB density relies on two different measurements, SE decay and ADC evaluation, which are needed to obtain  $G_i$  as a more specific parameter to reflect bone properties.

Since the fitting procedure involves the evaluation of the coefficient multiplying  $TE^3$  in the second term of equation (2), the SD associated with  $G_i$  also depends on the uncertainty with which the ADC is known. From the statistical error propagation theory, the diffusion coefficient uncertainty affects the total variance with a term proportional to the square of the  $G_i$  times and the relative error associated with the ADC:

$$\frac{\sigma_{G_i}}{G_i} = \frac{\sigma_B}{B} + \frac{\sigma_D}{D}, \quad (3)$$

where  $B = \frac{1}{12} \gamma^2 G_i^2 D$ ,  $\sigma_{G_i}$ ,  $\sigma_B$  and  $\sigma_D$  are the SDs associated with  $G_i$ ,  $B$  and ADC respectively. In a common experimental fit,  $B$  can be derived with a relative uncertainty of less than 10%. Also, an uncertainty of approximately 10% on the ADC measurement can be afforded using common clinical scanners. As a consequence,  $G_i$  can be estimated with a relative error that should not exceed 20%. From the perspective of a clinical application, this may guarantee the precision needed for  $G_i$  to discriminate among subjects characterized by different TB densities.

#### 4. Conclusions

In this study, using high field microimaging to investigate spongy-bone samples, we demonstrate that water  $G_i$  is directly proportional to TB densities, while fat  $G_i$  does not provide specific information on bone's structural features. Moreover, we show that the concentration of water molecules at the bone–bone marrow interface is higher than that of fat molecules. It is further confirmed that measuring water MR parameters in spongy bone provides information on TB densities. To test  $G_i$  as a potential surrogate marker of TB density, we measured (using a clinical MR scanner) this parameter in the calcaneus of human subjects at different ages and with different DXA  $T$ -scores. In this experimental setting, we were able to detect only water  $G_i$  due to software and hardware limitations of clinical scanners. Data obtained *in vivo* demonstrated a progressive reduction of  $G_i$  across aging, thus suggesting that  $G_i$  decreases proportionally to the physiological reduction of TB density. Future studies, including both large populations of healthy subjects and patients with osteopenia and osteoporosis, are needed to clarify the potential diagnostic role of  $G_i$  in clinical protocols.

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