



Review

Machine Learning Approaches in Diagnosis, Prognosis and Treatment Selection of Cardiac Amyloidosis

Alessandro Allegra ^{1,*}, Giuseppe Mirabile ¹, Alessandro Tonacci ², Sara Genovese ³, Giovanni Pioggia ³ and Sebastiano Gangemi ⁴

¹ Division of Hematology, Department of Human Pathology in Adulthood and Childhood “Gaetano Barresi”, University of Messina, 98125 Messina, Italy

² Clinical Physiology Institute, National Research Council of Italy (IFC-CNR), 56124 Pisa, Italy

³ Institute for Biomedical Research and Innovation (IRIB), National Research Council of Italy (CNR), 98164 Messina, Italy

⁴ Allergy and Clinical Immunology Unit, Department of Clinical and Experimental Medicine, University of Messina, 98125 Messina, Italy

* Correspondence: aallegra@unime.it

Abstract: Cardiac amyloidosis is an uncommon restrictive cardiomyopathy featuring an unregulated amyloid protein deposition that impairs organic function. Early cardiac amyloidosis diagnosis is generally delayed by indistinguishable clinical findings of more frequent hypertrophic diseases. Furthermore, amyloidosis is divided into various groups, according to a generally accepted taxonomy, based on the proteins that make up the amyloid deposits; a careful differentiation between the various forms of amyloidosis is necessary to undertake an adequate therapeutic treatment. Thus, cardiac amyloidosis is thought to be underdiagnosed, which delays necessary therapeutic procedures, diminishing quality of life and impairing clinical prognosis. The diagnostic work-up for cardiac amyloidosis begins with the identification of clinical features, electrocardiographic and imaging findings suggestive or compatible with cardiac amyloidosis, and often requires the histological demonstration of amyloid deposition. One approach to overcome the difficulty of an early diagnosis is the use of automated diagnostic algorithms. Machine learning enables the automatic extraction of salient information from “raw data” without the need for pre-processing methods based on the a priori knowledge of the human operator. This review attempts to assess the various diagnostic approaches and artificial intelligence computational techniques in the detection of cardiac amyloidosis.

Keywords: cardiac amyloidosis; artificial intelligence; machine learning; deep learning; ATTRwt amyloidosis; AL amyloidosis; hypertrophic cardiomyopathy; multiple myeloma; diagnosis



Citation: Allegra, A.; Mirabile, G.; Tonacci, A.; Genovese, S.; Pioggia, G.; Gangemi, S. Machine Learning Approaches in Diagnosis, Prognosis and Treatment Selection of Cardiac Amyloidosis. *Int. J. Mol. Sci.* **2023**, *24*, 5680. <https://doi.org/10.3390/ijms24065680>

Academic Editor: Olga V. Fedorova

Received: 22 February 2023

Revised: 12 March 2023

Accepted: 14 March 2023

Published: 16 March 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

General Considerations on Amyloidosis

A rare class of diseases known as amyloidoses is brought on by the extracellular deposition of amyloid, a fibrillar material created by the organism from several precursor proteins. These proteins harm essential organs by self-assembling into highly structured aberrant conformations and depositing as minute insoluble fibers. A variety of proteins can be used to create amyloid. Some varieties are passed down from one generation to the next. Others are brought on by external factors, such as chronic kidney disease or inflammatory disease, or they develop as a result of other disorders. Among the collection of diverse conditions, which includes Creutzfeldt–Jakob, Alzheimer’s, Huntington’s, and prion diseases, there are more than 20 amyloidosis diseases [1]. The affected organs determine the type and severity of amyloidosis symptoms, as some amyloidoses are localized, whereas others are systemic. As a result, some people only experience modest symptoms, while others experience serious diseases that could be deadly [2]. The clinical overlap with other illnesses is the greatest obstacle to distinguish the various amyloidoses.

Systemic amyloidosis is divided into various groups, according to a generally accepted taxonomy, based on the proteins that make up the amyloid deposits [3]. Primary amyloidosis (AL), often referred to as amyloidosis from light chains, is characterized by changes in plasma cells that produce a significant amount of abnormal antibody proteins. The most typical locations for primary amyloid deposits include the heart, liver, spleen, tongue, kidneys, gut, arteries, skin, and nerves.

Secondary amyloidosis (AA) is due to infections, persistent inflammation, certain types of tumors. In this case, the kidney is the organ that is most severely impacted, while other organs may also sustain damage.

A group of uncommon genetic disorders known as hereditary amyloidosis are brought on by abnormalities in specific blood proteins. These modified proteins are what lead to amyloid fibrils, which have an impact on the heart, kidneys, and nervous system.

A liver-produced protein called altered transthyretin (ATTRm) is the most frequently identified cause of amyloidosis. When ATTR-CM patients experience symptomatic heart failure (HF), their functional status and quality of life gradually deteriorate, and their morbidity and mortality are substantial, with high rates of hospitalization and death [4–7].

Fibrils that build up in the various organs generate ATTR-CM, a fatal disease that progresses over time [8]. In fact, the deposition of amyloid in patients with wild-type ATTR-CM is systemic and not restricted to the heart. As a result of multiorgan involvement, clinical symptoms include carpal tunnel syndrome, lumbar stenosis, renal illness, and neuropathy of variable severity [9,10]. However, to identify patients at high risk for ATTR-CM, no validated clinical decision support (CDS) or reporting methods that take into account the various comorbidities are currently available for use in healthcare system databases or at the point of care.

Wild-type ATTR-CM is frequently misdiagnosed or detected late in the illness course despite increased awareness because patients' HF is mistakenly assigned to more widespread causes, such as hypertension or renal disease [11,12].

AL amyloidosis is a separate type of amyloidosis, as previously mentioned. It is a rare, severe, progressive, crippling, systemic disease that is somewhat more prevalent in men and has a median age at diagnosis of roughly 65 years [13–16]. However, other less frequent forms of cardiac amyloidosis have been identified (Table 1).

Table 1. Some forms of cardiac amyloidosis.

Amyloid Type	Protein	Distribution	Etiology	Diseases Associations
AL	Immunoglobulin light chain	Systemic and localized	Acquired	Plasma cell dyscrasia
AH	Immunoglobulin heavy chain	Systemic	Acquired	Plasma cell dyscrasia
ATTR	Transthyretin	Systemic	Acquired and hereditary	
ATTRwt	Wild-type transthyretin	Systemic	Acquired	Aging
ATTRv	TTR gene variant	Systemic	Hereditary	
AA	Serum amyloid A	Systemic	Acquired	Chronic inflammation
A β 2M	β 2-microglobulin	Systemic	Acquired and hereditary	Chronic hemodialysis
AANF	ANF	Localized	Acquired	Atrial fibrillation

More than 61% of fatalities are attributable to cardiac amyloid infiltration, which is also the main factor in determining survival in individuals with AL amyloidosis. Arrhythmias or progressive heart failure are cardiovascular events that can result in mortality; however, the exact reason is frequently unknown [17]. Atrial or ventricular arrhythmia, heart failure, embolism, stroke, and conduction delays, including severe atrioventricular block, are common cardiovascular problems.

The cardiac stage of the illness at diagnosis affects median survival [18]. In modern cohorts, the median survival is close to five years [19]. Patients with less severe cardiac involvement have a median lifespan of roughly two years compared to only four months for those with highly advanced cardiac illness [20]. The overall survival of patients has increased because of the introduction of new therapeutic drugs and regimens as well as early diagnosis [21–24]. Clonal plasma cell growth is the underlying cause of AL amyloi-

dosis, which results in amyloidogenic immunoglobulin (Ig) light chains that aggregate to form insoluble fibrils that deposit in tissues and lead to organ failure [25–28]. More than 75% of patients with AL amyloidosis experience symptoms in the heart, making it the most frequently affected organ.

Because of its insidious onset, the rate of misdiagnosis is high, the missed diagnosis rate is still very high, and it usually progresses to a late stage by the time it is diagnosed. Instead, early diagnosis is important for a number of reasons: overall survival is poor once cardiac involvement occurs; chemotherapy followed by stem cell transplantation has significantly improved prognosis for AL amyloidosis; antibody-mediated fibril phagocytosis as well as TTR gene silencers and protein stabilizers are emerging; and recent successful trials on all-cause mortality and hospitalization are evidence of these developments. In order to improve patient outcomes with this fatal condition, awareness must grow [29].

2. Machine Learning and Deep Learning

It is obvious from what has been discussed so far that the differential diagnosis of the various kinds of amyloidosis is difficult. This is why the potential application of artificial intelligence (AI) methods seems intriguing. In order to generate predictions or choices without being explicitly taught to do so, machine learning (ML) algorithms construct a mathematical model from sample data, which is also referred to as “training data”. The most accurate human procedures are comparable in speed and precision to automated ML analysis [30–32].

A subclass of AI, known as deep learning (DL), uses “raw data” to automatically discover important features utilizing a hierarchy of representation levels that are not explicitly created by humans, unlike in ML. Through the use of a number of levels of representation, DL enables the automatic extraction of salient information from “raw data” without the strict need for pre-processing methods based on the a priori knowledge of the human operator. The integration of DL-based predictive analytics into clinical imaging in the medical field follows a logical evolution, with advances in cardiovascular imaging now producing high-fidelity datasets with more data than those obtained from older scanner generations [33,34]. Clinical imaging integration with DL-based algorithms has the potential to automate repetitive operations, enhance disease diagnosis and prognosis, and reveal novel biomarkers linked to certain disease processes [35–37].

The overall structure of AI-based methods employed in the field of amyloidosis, with information about the most common methods retrieved, is displayed in Figure 1.

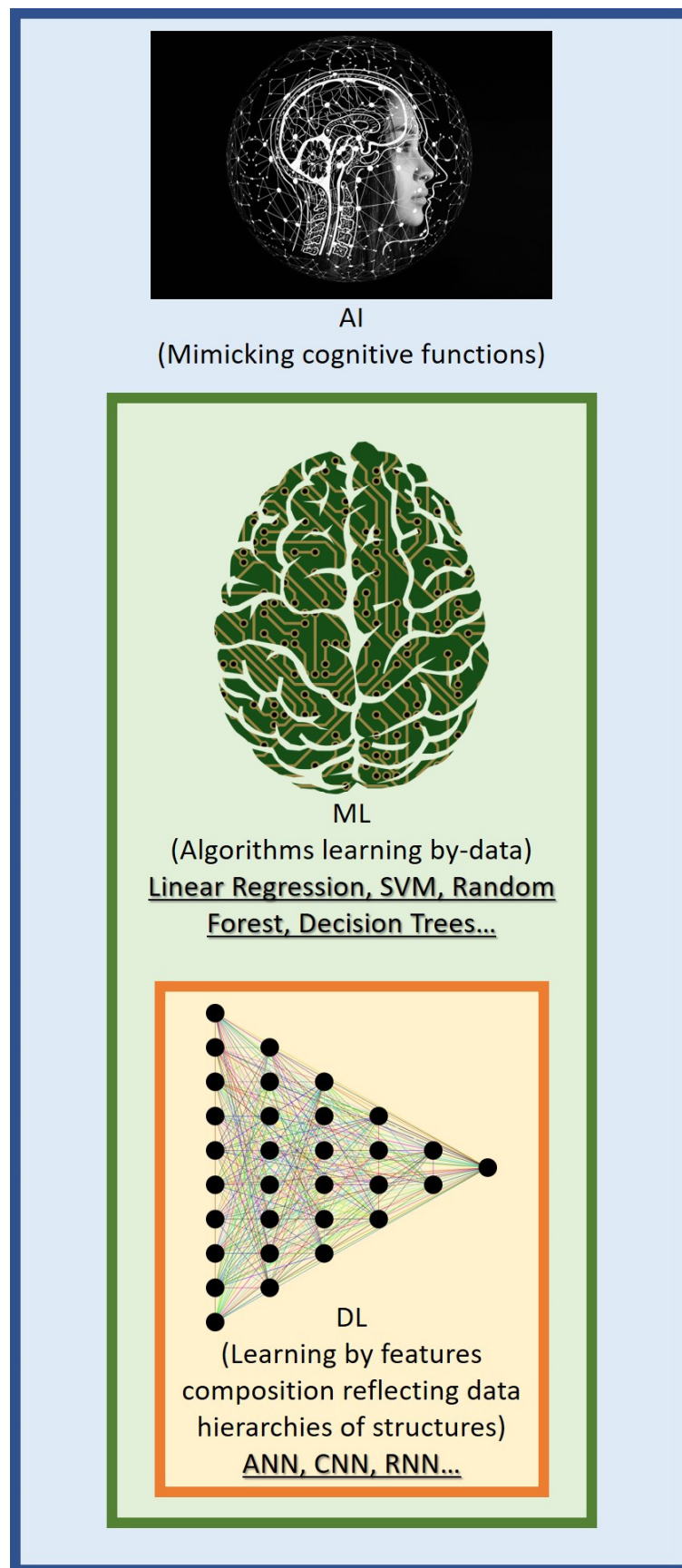


Figure 1. Main AI-based approaches employed in the field of amyloidosis.

3. Diagnosis of Amyloidosis

Once amyloidosis is suspected, a number of tests can be performed to confirm the diagnosis, to look for an underlying cardiopathy, and to identify the affected organs. Ig light chain abnormalities in serum can be found using free light chain (FLC) tests [38–40]. Immunofixation electrophoresis assays on serum and urine are used to measure and categorize aberrant Ig light chains or monoclonal proteins [41]. Positive outcomes from these assays would point to the presence of multiple myeloma, indolent B cell lymphoma, Waldenström macroglobulinemia, or AL amyloidosis [42]. In the majority of individuals with AL amyloidosis, the combination of these assays can be quite successful in detecting amyloidogenic light chains [42].

Confirmation of the diagnosis requires Congo red staining of a tissue biopsy sample. Amyloid deposits can be found quickly and safely with abdominal fat pad biopsies [43]. However, although the diagnosis of amyloidosis is confirmed by the presence of amyloid deposits by Congo red staining in fat pads, their absence does not rule out illness. A different key step in the diagnosis of AL amyloidosis is a bone marrow examination. About 85% of patients with AL amyloidosis will be detected using a combination of fat and bone marrow biopsies [44].

As for other forms of amyloidosis, in a significant, prospective, observational analysis of patients with ATTRwt-CM, the median diagnostic delay was 39 months, and 42% of patients experienced a diagnosis delay of more than 4 years [45]. Delayed ATTR-CM diagnosis can cause more advanced disease at diagnosis and result in needless consultations, testing, and treatment before diagnosis [46,47].

Cardiac involvement is frequent in both AL and ATTR amyloidosis, is associated with a worse prognosis, and has significant therapeutic implications [48]. CA is more likely to occur in patients with hypertrophic cardiomyopathy (HCM), and these two disorders can be misdiagnosed in a patient [49]. An estimated 1 in 500 to 1 in 5000 people in the general population have HCM.

Diagnoses for patients older than 60 years of age grew from 9.2% in 2000 to 31.8% after 2010, and the prevalence of diagnoses for patients older than 70 years of age climbed to 10.7% after 2010. Older patients were more frequently genotype-negative and more frequently affected by sporadic manifestations of the disease [50].

Due to major improvements in diagnostic and treatment approaches over the past few years, interest in an early identification of the disease—traditionally regarded as rare and incurable—has drastically transformed [51]. The most important diagnostic advancements are in the area of ATTR-CA, whose diagnostic process has been significantly changed by the ability to obtain a non-biopsy diagnosis by cardiac scintigraphy with bone tracers. This method is the cornerstone of an algorithm that has been tested on a sizable cohort of CA patients, with results showing 99% sensitivity, 86% specificity, and a positive predictive value that is very close to 100% when used in conjunction with serum/urine immunofixation and the detection of free light chains, which rule out the presence of monoclonal proteins [52]. As a result, diagnoses of ATTR-CA have dramatically increased, and patients are now identified earlier in the course of the disease.

The identification of clinical features, electrocardiographic (ECG), and imaging findings suggestive or compatible with CA is the first step in the diagnostic process. Amyloid deposition must frequently be shown histologically, with the exception of cases where diphosphonate scintigraphy reveals a significant increase in myocardial uptake without the presence of a monoclonal gammopathy (Perugini scores 2–3) [2]. Cardiovascular magnetic resonance (CMR) has a special ability that makes it possible to characterize cardiac tissue, although its function in this diagnostic flowchart is not entirely clear [52]. A pattern of fluctuating biventricular pseudohypertrophy with widespread subendocardial-to-transmural late gadolinium enhancement (LGE) is one of several CMR findings that are quite distinctive for CA. The extent of LGE and the degree of wall thickness growth are related to the level of amyloid fiber infiltration in the myocardium. In fact, in individuals with early disease stages, amyloid deposition is restricted to the sub-endocardium of a small number of cardiac segments and becomes increasingly diffuse [53]. The myocardial and blood-pool

gadolinium kinetics are entirely disrupted in the final phases of amyloid infiltration, with diffuse gadolinium retention in the myocardium and a faster gadolinium washout from the blood pool. This could make it difficult to determine the ideal myocardial inversion time (TI) in post-contrast pictures in order to get acceptable LGE images. For these situations, TI-scout sequences [54], early-to-late enhancement acquisitions [55], phase-sensitive inversion recovery (PSIR) LGE sequences [56], native T1 mapping [57], and extracellular volume fraction (ECV) [58] may be useful in making the diagnosis and identifying the stage of the disease.

The following sections will attempt to assess the various diagnostic approaches and their applicability in the detection of cardiac amyloidosis.

3.1. *Electrocardiographic Evaluation in the Diagnostic Work-Up for CA*

Even though the left ventricular wall width is thicker, the presence of a low-voltage electrocardiographic signal strongly suggests CA and can distinguish it from hypertensive or hypertrophic cardiomyopathy [59]. However, only 25% to 40% of ATTR-CA patients match the low-voltage criteria and may even meet the requirements for left ventricular hypertrophy [60]. Unknown factors may have caused the low-voltage ECG. Myocyte shrinkage and cardiac toxicity brought on by circulating light chains are potential contributing factors in AL amyloidosis [61]. In descriptive studies, more than half of the affected patients had an absence of R progression, sometimes known as a pseudo-infarction pattern, in the anterior precordial leads. However, only around 25% of patients had both a low ECG voltage and a pseudo-infarct pattern [62].

Artificial intelligence methods have been effective in deciphering electrocardiographic data to diagnose CA. A study obtained electrocardiographic maps of CA patients using ECG imaging. The complicated data sets collected from ECG data were then decoded using an ML approach, which also assisted in the creation of a straightforward surface ECG-based diagnostic algorithm for the identification of CA [63]. Two distinctive patterns were visible in the respective surface ECG leads. An ECG-based tool was created using thorough electroanatomic mapping of CA patients and an ML methodology. The ECG algorithm is straightforward and has been shown to be useful in detecting CA without the assistance of cutting-edge imaging modalities.

3.2. *Echocardiography Evaluation in the Diagnostic Work-Up for CA*

As reported above, CA is fatal, has a poor prognosis, and is frequently misdiagnosed as hypertrophic cardiomyopathy, which delays diagnosis [64]. The automated differentiation of two conditions using speckle tracking echocardiography was proposed. A recent study reported the findings of a great multicenter study assessing the diagnostic precision of a combination of traditional (non-deformation) and strain-derived echocardiographic variables in subjects with suspected CA. Authors derived two simple multiparametric scores to either identify or exclude CA in two different clinical settings: among subjects with established systemic AL amyloidosis or in subjects with a hypertrophic cardiac phenotype. Finally, they ascertained the functional and structural changes across the spectrum of gravity of amyloid deposition [65].

In order to assess the diagnostic benefit produced by AI techniques in the echocardiographic diagnosis of CA, patients with pathologically confirmed monoclonal immunoglobulin light chain cardiac amyloidosis and patients with hypertrophic cardiomyopathy were enrolled in a study. The most important indicators were found using ML models built using both conventional and cutting-edge technologies. The receiver operating characteristic curve (ROC) and area under the curve (AUC) were used to assess the performance. All models demonstrated excellent discriminative performance using clinical and echocardiography data. Support vector machine (AUC 0.95, $p = 0.477$), random forest (AUC 0.97, $p = 0.31$), and gradient boosting machine (AUC 0.98) showed comparable capacity to differentiate cardiac amyloidosis and hypertrophic cardiomyopathy when compared to logistic regression (AUC 0.91) [66]. The ability of data-driven machine learning to distinguish be-

tween two circumstances has been impressive. It can also automatically integrate a wealth of information to find the most discriminating predictors without making any assumptions. Automated ML can help identify patients with cardiac amyloidosis and enable prompt and efficient intervention, improving the prognosis in the big data era.

A different study sought to create machine learning models with robust generalizability across numerous cohorts to detect HCM and distinguish it from other heart diseases using ECGs and echocardiograms [67]. On a data set with a realistic HCM prevalence, automated detection and manual interpretation were tested. The models distinguished between HCM and aortic stenosis, hypertension, and cardiac amyloidosis. Compared to cardiologists, automated HCM identification had a greater sensitivity in electrocardiography–echocardiography paired data analysis [67].

Finally, several other studies have attempted to automatically identify patients with unexplained left ventricular hypertrophy in electronic health record (EHR) data using computer methods such as text mining and ML [68–70].

3.3. Magnetic Resonance Evaluation in the Diagnostic Work-Up for CA

A common method for imaging the cardiovascular system for diagnosis and intervention is cardiac MRI (CMR). In addition to visualizing the anatomy, CMR has improved to the point that it can now be utilized to provide detailed quantitative measurements of the myocardium. These include physiologic measurements for mapping myocardial perfusion and blood volume, as well as relaxometry T1, T2, and T2* measures for the assessment of fibrosis, oedema, and iron as well as for assessment of tissue composition for the fat fraction [71]. A variety of disease states, such as diffuse interstitial or replacement fibrosis, oedema, or infiltrative diseases, can benefit from myocardial T1 mapping [72].

While in-patient regional or even temporal variations in native T1 may be informative of disease development or progression, cross-patient comparisons are less relevant due to the considerable overlap in myocardial native T1 levels between health and disease. The diagnostic value of T1 mapping for the quick identification of myocardial illness without the use of contrast agents would be improved by a method that could make use of the wealth of information included in myocardial T1 maps. The segmented anatomical volumes are converted into quantitative radiomic characteristics that, by examining the spatial relationship between (dis-)similar voxels (akin to terrain mapping), reveal details about tissue volume, shape, and texture [73–79].

The left ventricle is used as the main chamber for validating diagnoses. Recent research, however, has demonstrated that cardiac strain has discriminative value for all cardiac chambers and may help distinguish CA from HCM and healthy controls (CTRL) [80]. An experiment tested the hypothesis that strain measurements of the diagnostically less common cardiac chambers combined with general cardiac functional parameters may achieve a high CA diagnostic accuracy. This hypothesis was based on prior observations [81–84] and the desire to optimize CMR diagnostic accuracy for CA detection. The right ventricle and atria would therefore be the only sources of cardiac strain metrics.

By putting cardiac function and multi-chamber strain into supervised ML algorithms, the work challenges CA diagnoses [85]. CMR imaging was performed on CA patients, HCM participants, and healthy volunteers. Using the strain values for the left, right, and right ventricular atriums as well as heart function, the decision trees (DT), k-nearest neighbor (KNN), SVM linear, and SVM radial basis function (RBF) kernel algorithms developed a 41-feature matrix. Using linear SVM and RBF, a 10-feature principal component analysis (PCA) was carried out [85]. As a result, under supervised conditions, the SVM RBF kernel obtained competitive diagnostic accuracies. For non-contrast clinical decision-support systems in CA diagnostics, ML of multi-chamber cardiac strain and function may present unique perspectives.

However, other new demands are simultaneously placed on image analysis and reporting by CMR capabilities, opening up new prospects [86–91]. AI methods could speed up analysis times in addition to improving diagnostic accuracy. Higher patient

throughputs, better objectivity, and reproducibility are all benefits of a fully automated solution. Cardiologists still primarily use manual delineation in clinical settings to evaluate heart function, viability, and tissue characteristics [92]. An expert's thorough manual analysis can take between 9 and 19 minutes, according to recent research [93]. Convolutional neural networks (CNNs) in particular have been developed as DL models to automate CMR analysis and greatly shorten analysis times. CNN measurements have shown better reliability in multicenter studies [94], and they can be used to automatically interpret cardiac cine images to quantify the ejection fraction and other parameters with a performance level comparable to experienced readers. CNNs have been used to interpret and report on cardiac perfusion pictures obtained from MRI scanners [95]. In tests involving multiple centers, CNNs have also been constructed to measure LV function [96]. In a study, the LV blood volume and myocardium were segmented using a completely automated ML algorithm that was trained using 1923 images [97]. Performance of an ML algorithm was compared to that of three physicians (the "human") and a commercial tool (cvi42, Circle Cardiovascular Imaging). Human analysis took longer (20 s per patient) than machine analysis (13 min). The combined dataset's overall machine mis-segmentation rate was 1 in 479 pictures, which largely occurred in rare diseases that were not observed during training. Without correcting these segmentation errors, machine analysis outperformed three physicians in terms of precision, which resulted in a 46% decrease in the trial sample size needed to use an LVEF objective [97]. Consequently, a fully automated approach that outperforms human performance in terms of speed and accuracy can be used to measure LV structure and global systolic function.

The superiority of AI techniques was supported by additional investigations [98]. Using non-contrast cine CMR images, a study was conducted to see if texture analysis (TA) and ML-based classifications might be used in the differential diagnosis of CA [99]. Cine pictures were used to extract texture features. Nine ML models were created based on regression analysis with the least absolute shrinkage and selection operator (LASSO), and their diagnostic performance was assessed. The combined model including traditional MR metrics and radiomics texture characteristics performed better in terms of discrimination than the traditional MR metrics model. Additionally, the findings demonstrated that gray level non-uniformity (GLenNonU) levels in HCM patients were considerably greater than levels in CA patients and control groups. It has been proven that GLenNonU 25 can distinguish between CA and HCM patients. The discriminatory performance can be enhanced using the radiomics-MR combined model, and TA can be used to evaluate the changes in myocardial microstructure that take place across various phases of cardiomyopathies.

Another analysis produced similar outcomes. It was investigated in a study [100] whether radiomic characteristics from T1 maps obtained by CMR may improve the diagnostic utility of T1 mapping in separating health from disease and categorizing cardiac disease phenotypes. The type of heart illness was clearly associated with the first three major T1 radiomic components. The population's unsupervised hierarchical grouping by myocardial T1 radiomics was substantially correlated with the type of cardiac illness [100].

In a subsequent investigation, CMR was carried out on participants who had suspected CA [101]. Three networks analyzed two-chamber, four-chamber, and short-axis LGE pictures (DL algorithms). When the average probability of CA from the three networks was 50% or less, the tags "amyloidosis present" or "absence" were assigned accordingly. The DL approach was contrasted with an ML algorithm taking into account all manually derived features to mimic an expert operator reading an exam. The DL approach demonstrated appropriate diagnostic precision. The diagnostic yield of an ML algorithm that took into account every CMR feature was comparable to a DL technique [101]. An ML-based strategy that simulates CMR reading by skilled operators and a DL approach evaluating LGE acquisitions both demonstrated comparable diagnostic performance for CA. However, there are a number of limitations to this study that must be acknowledged, including the fact that CA prevalence was very high. The study would also have benefited from an external validation cohort with a good representation of patients with hypertensive heart

disease, hypertrophic cardiomyopathy, cardiac sarcoidosis, and other pathologies that could be mistaken for CA, as well as other limitations that are important to note. Additionally, because there were so few patients in the study, AL and ATTR cardiomyopathies were combined into a single diagnostic entity (CA).

Finally, three-dimensional myocardial deformation analysis, for example, has been incorporated into the computational analysis for the detection of the CA (3D-MDA). This parameter may provide suitable features for the training of ML-based models because it has been validated to provide uniform descriptors of cardiac architecture and deformation. Using automated disease identification to distinguish HCM from states that mirror it, such as CA, Anderson–Fabry disease, and hypertensive cardiomyopathy, was the goal of one study (HTNcm). To distinguish between HCM and other situations, a fully connected layer feed-forward neural network was developed [102]. This study revealed that it is possible to diagnose cardiomyopathy states using only 3D-MDA and that it is capable of differentiating between HCM and similar disease states. These results point to a promising future for computer-assisted diagnosis in clinical settings, and this finding that regional deformation factors add to global architectural and global functional aspects in determining illness etiology is in line with new data showing that these measures are closely related to underlying tissue characteristics [103]. In patients with HCM [104–107] and CA [108], as well as in non-hypertrophic conditions such as ischemic and dilated cardiomyopathy, strain-based markers have demonstrated a strong regional connection with underlying signs of fibrosis (i.e., interstitial expansion). As a result, there is potential for regionally encoded deformation indicators to offer novel insights into underlying myocardial tissue health and, in turn, provide pertinent data for the classification of illness genesis.

3.4. Mass Spectrometry Evaluation in the Diagnostic Work-Up for CA

Immunohistochemical examination of biopsies from the affected organ or tissue has historically been used to determine the sub-type of the amyloidogenic protein [109,110]. However, because of its low sensitivity and low specificity [111–113], the latter of which is probably related to unspecific staining, this approach has been abandoned by many clinical pathology departments. However, utilizing immunohistochemistry (IHC), certain diagnostic facilities consistently attain a sensitivity higher than 90% [114]. Other recent techniques for classifying amyloidosis include mass-spectrometry-based shotgun proteome analyses of biopsies [115] and laser microdissection (LMD) of amyloid deposits visualized by Congo red staining combined with MS or immune electron microscopy (IEM) for the classification of localized amyloidosis [115–117]. In addition to measuring the amyloidogenic protein in question, mass-spectrometry-based approaches for amyloidosis subtyping also assess an amyloid protein signature that is common to all amyloidosis subtypes in distinct tissues in a very specific and quantitative manner.

A method for the objective detection of biopsies containing amyloid and amyloidosis subtyping was created by a study with the use of statistical models [118]. Clusterin, fibulin-1, vitronectin, complement component C9, three collagen proteins, as well as the well-known amyloid signature proteins apolipoprotein E, apolipoprotein A4, and serum amyloid P were discovered as novel “amyloid signature” proteins using a Boruta method applied on a random forest classifier to proteomics data obtained from the mass spectrometric analysis of Congo-red-positive amyloid-containing biopsies and Congo-red-negative biopsies in order to train an SVM learning system. When used on a blinded mass spectrometry validation data set of amyloid-containing biopsies that had been obtained, the trained algorithm outperformed controls in the distinction of amyloid-containing biopsies from controls, with an accuracy of 1.0. Additionally, in 102 out of 103 blinded cases, this technique successfully diagnosed amyloidosis patients according to the subtype [118]. With the objective and trustworthy classification of amyloid deposits and of the particular amyloid subtype, this model-assisted technique found novel amyloid-related proteins and proved the application of mass-spectrometry-based data in clinical diagnostics of disease (Table 2).

Table 2. Possible ML approaches in the diagnosis of cardiac amyloidosis.

Technique	Parameters	Population	IA Method	Ref.
Electrocardiographic evaluation	R-peak time	Treatment-naïve patients with CA	Unsupervised machine learning approach	[63]
Echocardiography data	Speckle tracking echocardiography	Monoclonal immunoglobulin light chain cardiac amyloidosis and patients with hypertrophic cardiomyopathy	Support vector machine, random forest, and gradient boosting machine	[66]
	Time series of voltages recorded for 10 seconds at 250 Hz Echo videos standardized to 30 frames with 30 frames per second	Patients with hypertrophic cardiomyopathy	A federated learning approach	[67]
Magnetic resonance	Multi-chamber strain and heart function	Patients with hypertrophic cardiomyopathy	Decision tree, k-nearest neighbor, SVM linear, and SVM radial basis function kernel algorithm processing	[85]
	Left ventricular cavity and myocardium right ventricle	Adenosine stress and rest perfusion scans	A convolutional neural network	[95]
	Cine MRI images of left ventricle	Data from three major MR vendors	Convolutional neural networks	[96]
	Left ventricle blood volume and myocardium	Images from 1932 patients with multiple diseases from multiple centers	Convolutional neural networks	[97]
	Left ventricular volume in the end-systolic images	Patients with CA, HCM, and normal subjects	K-nearest neighbor, random forest, naïve Bayes, support vector machine, logistic regression, and artificial neural networks	[99]
	T1 mapping	Normal subjects, patients with left ventricular hypertrophy of various causes, patients with HCM, and patients with known cardiac amyloidosis	A random forest machine learning algorithm	[100]
	Biventricular systolic function	Subjects with suspected CA, unexplained left ventricular hypertrophy with blood dyscrasia and suspected light-chain amyloidosis	Three base convolutional neural networks	[101]
	Volumetric and strain markers	Patients from the Cardiovascular Imaging Registry of Canada	A neural-network-based model	[102]
Mass spectrometry	Clusterin1, fibulin-1, vitronectin, complement, apolipoprotein E	Congo-red-positive and -negative amyloid-containing biopsies	A Boruta method applied on a random forest classifier	[118]

Finally, immunoelectron microscopy is a specific and extremely sensitive method for amyloid typing; it employs gold-labelled antibodies binding to amyloid fibrils. It can be utilized on formalin-fixed, paraffin-embedded material and is believed to be a new gold standard for amyloid typing next to mass spectrometry [119].

3.5. Medical Data and Radiomics in the Diagnostic Work-Up for CA

Beyond adopting a specific diagnostic method to identify CA, AI may be advantageous in using patient data for accurate and speedy diagnosis. Inpatient and outpatient hospital data, demographics, diagnoses, patient interactions and medical practices, prescriptions, laboratory tests, and charges are all included in the medical database, which is based in hospitals [120–123]. According to a study, using information from medical claims, a random forest machine learning model may spot prospective cases of wild-type transthyretin amyloid cardiomyopathy. In 1071 cases and 1071 non-amyloid heart failure controls,

authors developed an ML model. Three nationally representative cohorts (9412 cases and 9412 matched controls) and a sizable single-center electronic health record-based cohort (261 cases and 39,393 controls) were used to validate the model [124]. The authors demonstrated that the ML model had good performance for detecting individuals with cCA in the derivation cohort and all four validation cohorts, giving a structured framework to raise suspicion of transthyretin CA in patients with heart failure.

An alternative study was conducted to create a routinely calculated set of laboratory-parameters-based expert-independent ML prediction model for CA [125]. The authors first created logistic-regression-based baseline linear models. In a subsequent phase, they performed non-linear prediction and enhanced their linear prediction model using an ML technique based on gradient tree boosting. The effectiveness of each diagnostic algorithm was then compared. A training cohort of patients with established CA and HF patients unrelated to amyloidosis served as the basis for all prediction models. The ROC AUC score for this model was 0.86, with sensitivity and specificity of 89.2% and 78.2%, respectively. This study reveals how ML enables the generation of a differentiated CA-related HF profile in comparison to HF patients without a CA connection using standard laboratory measures. This proof-of-concept study offers a potential new direction in the investigation of CA's diagnosis and could support doctors' clinical judgment.

4. Computational Methods for Amyloid Fibril Identification

The identification of the amyloid molecule is a critical step in the diagnostic procedure of a suspected amyloidosis. Immunohistochemistry and laser capture are the two most popular techniques for classifying amyloid fibrils, and mass spectrometry is then used to determine the amyloid subtype [114,115].

The amyloid fibrils appear to share a same structure regardless of the type of amyloid protein that makes them up. Therefore, it can be examined in vitro using X-ray diffraction techniques, electron microscopy, and certain chemical stains [126]. Due to the high volume of protein data generated by sequencing, computational approaches for detecting and forecasting fibrils have just recently evolved. However, these methods have cost and time constraints. Currently used protein sequencing methods run millions of reactions simultaneously while gathering enormous volumes of data.

The computational investigations have given us important atomic understanding. The "dock-and-lock" mechanism is a widely accepted explanation for fibril elongation that has been supported by extensive computational studies. In this mechanism, soluble monomers are quickly adsorbed on the fibril ends, primarily as a result of the entropy effect. This is followed by a slow structural rearrangement of the bound monomers, which is hampered by the need to break interactions between the monomers or misaligned monomer–fibril interactions. The process of the more recent discovery of fibril-dependent secondary nucleation is less well understood computationally. A proper interaction strength between the peptides and the fibril surface, the preference of the bound peptides for amyloidogenic conformations, the unrestricted movement of the peptides on the fibril surface, and their simple detachment for a quick turn-over are some of the key elements that simulation studies have so far identified as being crucial for an efficient secondary nucleation [127].

Doing more than just offering a molecular explanation for experimental findings regarding aggregation mechanisms is the ultimate goal of computational investigations.

Tian et al. [128] introduced the Prediction of the Amyloid Fibril-Forming Segment (Pafg) approach for finding fibril-forming segments in proteins based on a support vector machine (SVM). The Amino Acid Index database's 41 physicochemical attributes were employed in their model (AAindex) [129]. With a Matthews correlation coefficient of 0.63, Pafg achieved an overall accuracy of 81%, a specificity of 80%, and a sensitivity of 82% [128].

Employing a different approach, RFamyloid, a web server based on the SVMProt 188-D feature representation, the pse-in-one feature representation, and the random forest classifier, was presented as a method to discover amyloid proteins [130]. The Matthews

correlation coefficient, sensitivity, specificity, and F-measure were 0.739, 0.781, 0.927, and 0.891, respectively, while the accuracy was 89.19%.

Building an ML-based prediction model known as the PredAmylmultilayer perceptron led to similar outcomes. The authors integrated SVMProt-188D, tripeptide composition, and seven feature extraction techniques [131]. Other experiments used an accessible ReRF-Pred, a novel machine-learning approach based on a multi-feature encoding strategy to identify amyloidogenic regions utilizing the composition of tripeptides and pseudo amino acids [132–134]. A further attempt was performed using a linear SVM architecture on the Waltz database of 1415 total hexapeptides to classify amyloidogenicity patterns based on the short parts known as hot spots that might cause aggregation [135–137].

Because the datasets used in the literature are almost always unbalanced, a study was conducted to assess the classifier's behavior solely on balanced amyloid fibril datasets. The automated learning of the various ML algorithms is influenced by the performance on balanced or unbalanced datasets [138]. More than 4000 chemical descriptors were used in the development of this new classifier, dubbed ENTAIL. With an accuracy on the test set of 81.80%, a sensibility of 100%, and a specificity of 63.63% on a balanced dataset, ENTAIL was based on the Naive Bayes Classifier with Unbounded Support and Gaussian Kernel Type. The investigation that was carried out showed that performance is superior in terms of performance on a balanced dataset [139] despite the various configurations of the tests.

In addition, researchers have predicted the solubility and located the aggregation hotspots within amyloid-forming proteins using sequence-based aggregation-scoring algorithms such as GAP, TANGO, WALTZ, PASTA, Aggrescan, FoldAmyloid, ANuPP, etc. [140–148]. These algorithms have made use of sequence- and structure-based properties, including patterns of hydrophobic and polar residues, charges, the capacity to form cross-motifs, scales for aggregation propensity derived from experimental data, solvent-exposed hydrophobic patches on molecular surfaces, and others. These algorithms' benefits and drawbacks have been discussed elsewhere [149]. It has become widely accepted as a result of this research that the existence of an aggregation-prone region (APR) may be a prerequisite but not a necessary condition for protein aggregation. Other important parameters include the position of APRs in protein structure, the conformational stability of the native state, the conditions in the solution, and the kinetics of the aggregation process [150–156].

High individual antibody diversity is a significant problem for AL amyloidosis prediction and treatment. Although there are techniques for high-throughput antibody repertoire sequencing, it is not practical to experimentally determine each antibody's amyloidogenicity. Therefore, it is essential to create computational techniques for quick and precise prediction of aggregating light chains. The scientific community needs to develop the computational algorithms now in use because they are not effective enough to estimate the solubility of antibodies and, in certain cases, only weakly correlate with conformational stability [157,158].

Common architecture and well-known binding locations in antibody light chains can offer crucial details for the prediction of amyloidogenicity under physiological settings. In a study, authors compared the hydrophobicity, presence of gatekeeper residues, disorderness, and other traditional sequence-based, aggregation-related features calculated for the CDR, FR, or VL regions of amyloidogenic and non-amyloidogenic antibody light chains. They then implemented the findings in a machine-learning-based webserver called "VLAmY-Pred" (<https://web.iitm.ac.in/bioinfo2/vlamy>, accessed on 21 February 2023). On a dataset of 1828 variable region sequences of the antibody light chains, the model exhibited a prediction accuracy of 79.7% (sensitivity of 78.7% and specificity of 79.9%) with a ROC value of 0.88 [159].

This model will be useful for a better understanding of the prognosis for patients who may likely have diseases brought on by light chain amyloidosis, for large-scale *in silico* analysis of antibody sequences produced by next generation sequencing, and, finally, for rational engineering of aggregation-resistant antibodies (Table 3).

Table 3. Computational methods for amyloid fibril identification.

Method	Target	IA	Results	Ref.
Pafig	Hexpeptides associated with amyloid fibrillar aggregates	Support vector machine	Accuracy of 81% and Matthews correlation coefficient of 0.63	[128]
Pse-in-One	DNA, RNA, and protein sequences	Support vector machine and neural network	Matthews correlation coefficient 0.739, sensitivity 0.781, specificity 0.927, F-measure 0.891, and accuracy 89.19%	[130]
PredAmylmultilayer	Protein sequences	Waikato environment for knowledge analysis	Accuracy 91.59%, specificity 0.950, and sensitivity 0.836	[131]
ReRF-Pred	Composition of tripeptides and pseudo amino acids	Amyloidogenic regions	Accuracy 0.828, specificity 0.921, and Matthew correlation coefficient 0.619	[132]
ENTAIL	Protein precursors	Machine learning	Accuracy 81.80%, sensibility 100%, and specificity 63.63%	[139]
VLAmy-Pred	Tripeptides composition	Random forest	Accuracy 79.7%, sensitivity 78.7%, and specificity 79.9%	[159]

5. Prognosis

Prognostic risk can also be determined by computational analysis carried out utilizing artificial intelligence tools. In order to identify patterns that correlate with amyloidosis type and prognosis, a study used an automated method to aggregate individual patients with cardiac amyloidosis as defined by 24 prespecified clinical characteristics [160]. The researchers gathered information from 1394 consecutive patients. The cohort included patients with AL, ATTRv, ATTRwt, and no definitive amyloidosis diagnosis. The 24 domains chosen for phenotyping the patients broadly defined their characteristics, including demographics (sex, age, and race), biometrics (body mass index and blood pressure), cardiovascular risk factors, New York Heart Association (NYHA) functional class, cardiac biomarkers, renal function, anemia, electrocardiographic features (low voltage and arrhythmia), echocardiographic variables (septal wall thickness, LV ejection fraction, and global longitudinal strain), and evidence of sarcoidosis. Diagnostic characteristics of ATTR amyloidosis and AL amyloidosis were also among the variables selected for characterization (scintigraphy uptake or cardiac fixation). Data were first transformed using principal component analysis to account for variations in qualitative and quantitative variables in order to produce continuous scores. Following this, the data were subjected to an unsupervised clustering analysis to produce self-organized maps from which a total of seven discrete clusters were defined. After being placed on these clusters, patients with known amyloid types had their prognosis (4-year mortality) reported. Low global longitudinal strain (mean 8.4%) and LV ejection percent (mean 41%), low voltage, and monoclonal gammopathy were the characteristics of Cluster 1, which also had the highest levels of cardiac biomarkers and the highest NYHA functional class. Unsurprisingly, Cluster 1 had AL amyloidosis in 88%, accounting for 67% of all patients with AL amyloidosis. Older males, soft tissue deposits, the thickest LV septum, and the highest rates of bone-seeking radiotracer uptake were all present in Clusters 2 to 4. Clusters 2, 3, and 4 received the majority of the mapping for ATTRwt patients. These clusters differed from one another, allowing for additional differentiation. While Cluster 3 patients had arrhythmia but fewer cardiovascular risk factors, Cluster 4 patients had a high proportion of risk factors but preserved biochemical and functional parameters, and Cluster 2 patients had a higher proportion of cardiovascular risk factors and impaired biochemical and functional parameters. When survival was taken into account, these variations became significant. Comparing Clusters 3 and 4 (about 55% survival) to Cluster 2, ATTRwt patients had a 20% survival rate. In actuality, the survival rate for ATTRwt patients in Cluster 2 was lower (42% survival) than that of AL patients in Cluster 1. Patients with ATTRv amyloidosis were more uniformly spread out among

Clusters 2 to 4 as well as Cluster 6, which reflects the variation in phenotypic manifestation (cardiomyopathy and neuropathy) between the various TTR variants and the ages of ATTRv patients. Patients with nonamyloid disease were discovered in Clusters 5 to 7, with Cluster 7 standing out because patients there had greater body mass indices, diabetes, and dyslipidemia than in the other clusters, which is consistent with the profile of metabolic heart disease. In Cluster 7, the four-year death rate was 31% (69% survival) [160]. It is important to note the study's shortcomings. First, the investigators admitted that a sizeable amount of the missing data (up to 33% for scintigraphy) was handled via imputation, which is formally correct but could eventually draw spurious results. Second, using the same dataset, the clustering analysis was created and then examined for relationships with amyloid type. Third, not all of the analyses were unsupervised. Fourth, no information about the medical care obtained is provided. Without understanding the medicines used and their results, it is challenging to assess survival data [161].

6. Conclusions

Although there has been significant progress in the diagnosis and management of cardiac amyloidosis, there are still significant knowledge and diagnostic process gaps. Nuclear scintigraphy utilizing technetium-labelled bone-seeking radiotracers for ATTR cardiac amyloidosis has become widely available, which has facilitated diagnoses, although improper patient selection and interpretation continue to happen, leading to erroneous and/or missing diagnoses [162,163]. Furthermore, there is significant phenotypic overlap in the echocardiographic, cardiac magnetic resonance, and biomarker profiles of the most prevalent cardiac amyloidosis types of AL, ATTRwt, and variant transthyretin [ATTRv], which makes proper adjudication difficult.

A conclusive diagnosis of HCM is made using an echocardiography as well as cardiac MRI [164]. Due to the underdiagnosis of HCM, typical workflows involving these diagnostic modalities are obviously insufficient. This is due to factors such as cost, the subtlety of early disease findings, the invasiveness of the approach, the availability of the modality, as well as the requirement that non-specialist providers either remember the necessary diagnostic steps or make a timely referral. HCM can also be identified by distinctive ECG alterations [165–169], a method that is more generally accessible.

Several studies have developed ML strategies to detect HCM from both electrocardiography and echocardiography [170–173], but to date, these approaches have only involved a single modality and have only been evaluated at single centers.

Unfortunately, CA lacks a large amount of data when compared to other cardiovascular conditions, making the development of reliable AI-based models difficult and somewhat poorly scalable and generalizable. However, new frontiers of AI are advancing in this sense, with the development of novel principles and the practical application of well-grounded methods that are yet neglected or poorly used in medicine, which can promote a full adoption of AI in CA. Such methods, which are of different natures, include, for example, transfer learning, commonly used in scientific and technological realms, or federated learning, which enables huge amounts of data to be collected from various sources and also features overall superior safety and security for patients and their sensitive data.

Increased generalizability requires the use of vast amounts of data from various institutions. However, training data in the medical area could include personally identifiable information, making sharing difficult. An ML technique known as federated learning enables the training of an ML model using data sets from other institutions without sharing the raw data. Such an approach was never explored directly in CA, but the success demonstrated in HCM detection and differential diagnosis with respect to cardiac disorders featuring somewhat similar clinical characteristics, including CA, suggests its likely application in future studies dealing with CA [67]. This would benefit researchers and clinicians from several points of view, including greater data availability and the possibility to guarantee the patients whose data are collected with higher degrees of data protection and privacy regulations.

Accurate diagnosis is a prerequisite for individualized medicine, which necessitates analysis that goes beyond the data encoded in the systems utilized in clinical practice [174]. Clinical researchers have taken a keen interest in this area with the surge of electronic health records (EHRs), as the comprehensive and extensive use of clinical datasets holds enormous promise for revolutionizing the healthcare system. For instance, nursing records give information on patient conditions, and the data frequently reflects nurses' cumulative knowledge of how to evaluate physiological and psychological signs to identify patient deterioration.

The learning and training approaches of the algorithm are based on the inpatient clinical (medical and nursing) records employed for a study. The strategy involved treating the data produced by each admission and discharge occurrence as data vectors in order to facilitate data aggregation. The high dimensionality of the data was inferred by the vast number of clinical histories, and the absence of a diagnosis resulted in a significant class imbalance because the condition was not common. Despite the small number of patients with amyloidosis in this study, the suggested method shows that it is still possible to learn from clinical records in the absence of data. The algorithm originally used data from the overall research population during the validation phase. Then, it was used on a sample of people with heart failure. The findings showed that when data vectors profile each disease occurrence, the algorithm may detect disease. The prediction levels demonstrated that this method may be helpful in illness screening procedures on a particular group [175].

Given the trends noticed in medicine concerning the use of AI, and in particular in the cardiovascular field and in the CA diagnosis and detection, it is hypothesized that the number of scientific products related to this subject is probably destined to grow in the next few years. However, aside to the important scaling up of AI-related research, which is particularly useful to lead to new discoveries and important novelties and insights about physiopathological and etiopathological signs of the disease, important gaps are still present in the translation from bench to bedside of AI and technological advancements in general in every sector of medicine [176]. However, with the advent of new AI-based tools, which are pervasively entering everyday life (e.g., ChatGPT or similar programs), it is predictable that such novel approaches will be more often scaled into the clinical practice in the next few years, which will have significant benefits for the clinical community.

In conclusion, for the two most prevalent kinds of cardiac amyloidoses (ATTR and amyloidogenic light chain), disease-modifying therapies are now available, and earlier treatment in the course of the disease may be associated with a higher treatment response [177]. Therefore, it is crucial to diagnose cardiac amyloidosis early and more thoroughly. In any event, it is crucial to diagnose CA as soon as possible since, if left untreated, the median survival time after the onset of HF is just 6 months [178], even if contemporary treatments can prolong life by several years and put the disease into a protracted remission [179]. The prognosis of patients with AL-CA has significantly improved as a result of the availability of proteasome-inhibiting drugs, particularly bortezomib, which is frequently coupled with dexamethasone and low-dose cyclophosphamide. These factors make early identification of the illness a crucial clinical requirement. In the past few years, early diagnoses of the disease have resulted from the active screening for CA using new AI techniques [180]. This approach could guarantee an overall more adequate management of the pathology.

Author Contributions: Conceptualization, A.A., G.P., and S.G. (Sara Genoveseand); methodology, S.G. (Sebastiano Gangemi) and A.T.; G.M.; formal analysis, G.M., S.G. (Sebastiano Gangemi), and A.T.; data curation, A.T., S.G. (Sebastiano Gangemi), and G.M.; writing—original draft preparation, A.A.; writing—review and editing, A.A. and A.T.; supervision, A.A., G.P., and S.G. (Sara Genoveseand). All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. DeArmond, S.J. Alzheimer's disease and Creutzfeldt-Jakob disease: Overlap of pathogenic mechanisms. *Curr. Opin. Neurol.* **1993**, *6*, 872–881. [[CrossRef](#)] [[PubMed](#)]
2. Wechalekar, A.D.; Gillmore, J.D.; Hawkins, P.N. Systemic amyloidosis. *Lancet* **2016**, *387*, 2641–2654. [[CrossRef](#)] [[PubMed](#)]
3. Scott, P.P.; Scott, W.W., Jr.; Siegelman, S.S. Amyloidosis: An overview. In *Seminars in Roentgenology*; WB Saunders: Philadelphia, PA, USA, 1986; Volume 21, pp. 103–112.
4. Ruberg, F.L.; Berk, J.L. Transthyretin (TTR) cardiac amyloidosis. *Circulation* **2012**, *126*, 1286–1300. [[CrossRef](#)] [[PubMed](#)]
5. Kittleson, M.M.; Maurer, M.S.; Ambardekar, A.V.; Bullock-Palmer, R.P.; Chang, P.P.; Eisen, H.J.; Nair, A.P.; Nativi-Nicolau, J.; Ruberg, F.L.; American Heart Association Heart Failure and Transplantation Committee of the Council on Clinical Cardiology. Cardiac Amyloidosis: Evolving Diagnosis and Management: A Scientific Statement From the American Heart Association. *Circulation* **2020**, *142*, e7–e22. [[CrossRef](#)] [[PubMed](#)]
6. González-López, E.; Gallego-Delgado, M.; Guzzo-Merello, G.; de Haro-Del Moral, F.J.; Cobo-Marcos, M.; Robles, C.; Bornstein, B.; Salas, C.; Lara-Pezzi, E.; Alonso-Pulpon, L.; et al. Wild-type transthyretin amyloidosis as a cause of heart failure with preserved ejection fraction. *Eur. Heart J.* **2015**, *36*, 2585–2594. [[CrossRef](#)]
7. Castaño, A.; Narotsky, D.L.; Hamid, N.; Khalique, O.K.; Morgenstern, R.; DeLuca, A.; Rubin, J.; Chiuzan, C.; Nazif, T.; Vahl, T.; et al. Unveiling transthyretin cardiac amyloidosis and its predictors among elderly patients with severe aortic stenosis undergoing transcatheter aortic valve replacement. *Eur. Heart J.* **2017**, *38*, 2879–2887. [[CrossRef](#)]
8. Rapezzi, C.; Quarta, C.C.; Riva, L.; Longhi, S.; Gallelli, I.; Lorenzini, M.; Ciliberti, P.; Biagini, E.; Salvi, F.; Branzi, A. Transthyretin-related amyloidoses and the heart: A clinical overview. *Nat. Rev. Cardiol.* **2010**, *7*, 398–408. [[CrossRef](#)]
9. Fosbøl, E.L.; Rørth, R.; Leicht, B.P.; Schou, M.; Maurer, M.S.; Kristensen, S.L.; Kober, L.; Gustafsson, F. Association of Carpal Tunnel Syndrome With Amyloidosis, Heart Failure, and Adverse Cardiovascular Outcomes. *J. Am. Coll. Cardiol.* **2019**, *74*, 15–23. [[CrossRef](#)]
10. Witteles, R.M.; Bokhari, S.; Damy, T.; Elliott, P.M.; Falk, R.H.; Fine, N.M.; Gospodinova, M.; Obici, L.; Rapezzi, C.; Garcia-Pavia, P. Screening for Transthyretin Amyloid Cardiomyopathy in Everyday Practice. *JACC Heart Fail.* **2019**, *7*, 709–716. [[CrossRef](#)]
11. Abou Ezzeddine, O.F.; Davies, D.R.; Scott, C.G.; Fayyaz, A.U.; Askew, J.W.; McKie, P.M.; Noseworthy, P.A.; Johnson, G.B.; Dunlay, S.M.; Borlaug, B.A.; et al. Prevalence of Transthyretin Amyloid Cardiomyopathy in Heart Failure With Preserved Ejection Fraction. *JAMA Cardiol.* **2021**, *6*, 1267–1274. [[CrossRef](#)]
12. Bishop, E.; Brown, E.E.; Fajardo, J.; Barouch, L.A.; Judge, D.P.; Halushka, M.K. Seven factors predict a delayed diagnosis of cardiac amyloidosis. *Amyloid* **2018**, *25*, 174–179. [[CrossRef](#)] [[PubMed](#)]
13. Lousada, I. The amyloidosis forum: A public private partnership to advance drug development in AL amyloidosis. *Orphanet J. Rare Dis.* **2020**, *15*, 268. [[PubMed](#)]
14. Desport, E.; Bridoux, F.; Sirac, C.; Delbes, S.; Bender, S.; Fernandez, B.; Quillard, N.; Lacombe, C.; Goujon, J.M.; Lavergne, D.; et al. AL amyloidosis. *Orphanet J. Rare Dis.* **2012**, *7*, 54. [[CrossRef](#)]
15. Quock, T.P.; Yan, T.; Chang, E.; Guthrie, S.; Broder, M.S. Epidemiology of AL amyloidosis: A real-world study using US claims data. *Blood Adv.* **2018**, *2*, 1046–1053. [[CrossRef](#)] [[PubMed](#)]
16. Vaxman, I.; Gertz, M. Recent advances in the diagnosis, risk stratification, and management of systemic light-chain amyloidosis. *Acta Haematol.* **2019**, *141*, 93–106. [[CrossRef](#)]
17. Escher, F.; Senoner, M.; Doerler, J.; Zaruba, M.M.; Messner, M.; Mussner-Seeber, C.; Ebert, M.; Ensinger, C.; Mair, A.; Kroiss, A.; et al. When and how do patients with cardiac amyloidosis die? *Clin. Res. Cardiol.* **2020**, *109*, 78–88. [[CrossRef](#)]
18. Kumar, S.; Dispenzieri, A.; Lacy, M.Q.; Hayman, S.R.; Buadi, F.K.; Colby, C.; Laumann, K.; Zeldenrust, S.R.; Leung, N.; Dingli, D.; et al. Revised prognostic staging system for light chain amyloidosis incorporating cardiac biomarkers and serum free light chain measurements. *J. Clin. Oncol.* **2012**, *30*, 989–995. [[CrossRef](#)] [[PubMed](#)]
19. Kumar, S.K.; Gertz, M.A.; Lacy, M.Q.; Dingli, D.; Hayman, S.R.; Buadi, F.K.; Short-Detweiler, K.; Zeldenrust, S.R.; Leung, N.; Greipp, P.R.; et al. Recent improvements in survival in primary systemic amyloidosis and the importance of an early mortality risk score. In *Mayo Clinic Proceedings*; Elsevier: Amsterdam, The Netherlands, 2011; Volume 86, pp. 12–18.
20. Manwani, R.; Cohen, O.; Sharpley, F.; Mahmood, S.; Sachchithanatham, S.; Foard, D.; Lachmann, H.J.; Quarta, C.; Fontana, M.; Gillmore, J.D.; et al. A prospective observational study of 915 patients with systemic AL amyloidosis treated with upfront bortezomib. *Blood* **2019**, *134*, 2271–2280. [[CrossRef](#)]
21. Barrett, C.D.; Dobos, K.; Liedtke, M.; Tuzovic, M.; Haddad, F.; Kobayashi, Y.; Lafayette, R.; Fowler, M.B.; Arai, S.; Schrier, S.; et al. A changing landscape of mortality for systemic light chain amyloidosis. *J. Am. Coll. Cardiol. HF* **2019**, *7*, 958–966. [[CrossRef](#)]
22. Muchtar, E.; Gertz, M.A.; Kumar, S.K.; Lacy, M.Q.; Dingli, D.; Buadi, F.K.; Grogan, M.; Hayman, S.R.; Kapoor, P.; Leung, N.; et al. Improved outcomes for newly diagnosed AL amyloidosis between 2000 and 2014: Cracking the glass ceiling of early death. *Blood* **2017**, *129*, 2111–2119. [[CrossRef](#)]

23. Schulman, A.; Connors, L.H.; Weinberg, J.; Mendelson, L.M.; Joshi, T.; Shelton, A.C.; Sanchorawala, V. Patient outcomes in light chain (AL) amyloidosis: The clock is ticking from symptoms to diagnosis. *Eur. J. Haematol.* **2020**, *105*, 495–501. [[CrossRef](#)] [[PubMed](#)]
24. Vaxman, I.; Kumar, S.K.; Buadi, F.; Lacy, M.Q.; Dingli, D.; Hwa, Y.; Fonder, A.; Hobbs, M.; Hayman, S.; Kourelis, T.; et al. Outcomes among newly diagnosed AL amyloidosis patients with a very high NT-proBNP: Implications for trial design. *Leukemia* **2021**, *35*, 3604–3607. [[CrossRef](#)] [[PubMed](#)]
25. Baker, K.R.; Rice, L. The amyloidoses: Clinical features, diagnosis and treatment. *Methodist DeBakey Cardiovasc. J.* **2012**, *8*, 3–7. [[CrossRef](#)] [[PubMed](#)]
26. Merlini, G. AL amyloidosis: From molecular mechanisms to targeted therapies. *Hematol. Am. Soc. Hematol. Educ. Program* **2017**, *2017*, e009586. [[CrossRef](#)]
27. Imperlini, E.; Gnechi, M.; Rognoni, P.; Sabidò, E.; Ciuffreda, M.C.; Palladini, G.; Espadas, G.; Mancuso, F.M.; Bozzola, M.; Malpasso, G.; et al. Proteotoxicity in cardiac amyloidosis: Amyloidogenic light chains affect the levels of intracellular proteins in human heart cells. *Sci. Rep.* **2017**, *7*, 15661. [[CrossRef](#)]
28. Lavatelli, F.; Imperiini, E.; Orrù, S.; Rognoni, P.; Sarnataro, D.; Palladini, G.; Malpasso, G.; Soriano, M.E.; Di Fonzo, A.; Valentini, V.; et al. Novel mitochondrial protein interactors of immunoglobulin light chains causing heart amyloidosis. *FASEB J.* **2015**, *29*, 4614–4628. [[CrossRef](#)]
29. Oerlemans, M.I.F.J.; Rutten, K.H.G.; Minnema, M.C.; Raymakers, R.A.P.; Asselbergs, F.W.; de Jonge, N. Cardiac amyloidosis: The need for early diagnosis. *Neth. Heart J.* **2019**, *27*, 525–536. [[CrossRef](#)]
30. Gertz, M.A.; Dispenzieri, A. Systemic amyloidosis recognition, prognosis, and therapy: A systematic review. *JAMA* **2020**, *324*, 79–89. [[CrossRef](#)]
31. Gillmore, J.D.; Hawkins, P.N. Pathophysiology and treatment of systemic amyloidosis. *Nat. Rev. Nephrol.* **2013**, *9*, 574–586. [[CrossRef](#)]
32. Bhuva, A.N.; Bai, W.; Lau, C.; Davies, R.H.; Ye, Y.; Bulluck, H.; McAlindon, E.; Culotta, V.; Swoboda, P.P.; Captur, G.; et al. A Multicenter, Scan-Rescan, Human and Machine Learning CMR Study to Test Generalizability and Precision in Imaging Biomarker Analysis. *Circ. Cardiovasc. Imaging* **2019**, *12*, e009214. [[CrossRef](#)]
33. Henglin, M.; Stein, G.; Hushcha, P.V.; Snoek, J.; Wiltschko, A.B.; Cheng, S. Machine Learning Approaches in Cardiovascular Imaging. *Circ. Cardiovasc. Imaging* **2017**, *10*, e005614. [[CrossRef](#)]
34. Leiner, T.; Rueckert, D.; Suinesiaputra, A.; Baeßler, B.; Nezafat, R.; Išgum, I.; Young, A.A. Machine learning in cardiovascular magnetic resonance: Basic concepts and applications. *J. Cardiovasc. Magn. Reson.* **2019**, *21*, 61. [[CrossRef](#)]
35. Martin-Isla, C.; Campello, V.M.; Izquierdo, C.; Raisi-Estabragh, Z.; Baeßler, B.; Petersen, S.E.; Lekadir, K. Image-Based Cardiac Diagnosis With Machine Learning: A Review. *Front. Cardiovasc. Med.* **2020**, *7*, 1. [[CrossRef](#)]
36. Danieli, M.G.; Tonacci, A.; Paladini, A.; Longhi, E.; Moroncini, G.; Allegra, A.; Sansone, F.; Gangemi, S. A machine learning analysis to predict the response to intravenous and subcutaneous immunoglobulin in inflammatory myopathies. A proposal for a future multi-omics approach in autoimmune diseases. *Autoimmun. Rev.* **2022**, *21*, 103105. [[CrossRef](#)]
37. Allegra, A.; Tonacci, A.; Sciacotta, R.; Genovese, S.; Musolino, C.; Pioggia, G.; Gangemi, S. Machine Learning and Deep Learning Applications in Multiple Myeloma Diagnosis, Prognosis, and Treatment Selection. *Cancers* **2022**, *14*, 606. [[CrossRef](#)]
38. Kumar, S.; Dispenzieri, A.; Katzmann, J.A.; Larson, D.R.; Colby, C.L.; Lacy, M.Q.; Hayman, S.R.; Buadi, F.K.; Leung, N.; Zeldenrust, S.R.; et al. Serum immunoglobulin free light-chain measurement in primary amyloidosis: Prognostic value and correlations with clinical features. *Blood* **2010**, *116*, 5126–5129. [[CrossRef](#)]
39. Kim, H.-S.; Kim, H.S.; Shin, K.-S.; Song, W.; Kim, H.J.; Kim, H.S.; Park, M.J. Clinical comparisons of two free light chain assays to immunofixation electrophoresis for detecting monoclonal gammopathy. *Biomed. Res. Int.* **2014**, *2014*, 647238. [[CrossRef](#)]
40. Rubinstein, S.M.; Stockerl-Goldstein, K. How to screen for monoclonal gammopathy in patients with a suspected amyloidosis. *J. Am. Coll. Cardiol. CardioOnc.* **2021**, *3*, 590–593. [[CrossRef](#)]
41. Sanchorawala, V. Light-chain (AL) amyloidosis: Diagnosis and treatment. *Clin. J. Am. Soc. Nephrol.* **2006**, *1*, 1331–1341. [[CrossRef](#)]
42. Palladini, G.; Russo, P.; Bosoni, T.; Verga, L.; Sarais, G.; Lavatelli, F.; Nuvolone, M.; Obici, L.; Casarini, S.; Donadei, S.; et al. Identification of amyloidogenic light chains requires the combination of serum-free light chain assay with immunofixation of serum and urine. *Clin. Chem.* **2009**, *55*, 499–504. [[CrossRef](#)]
43. Wisniewski, B.; Wechalekar, A. Confirming the diagnosis of amyloidosis. *Acta Haematol.* **2020**, *143*, 312–321. [[CrossRef](#)] [[PubMed](#)]
44. Gertz, M.A. Immunoglobulin light chain amyloidosis: 2016 update on diagnosis, prognosis, and treatment. *Am. J. Hematol.* **2016**, *91*, 947–956. [[CrossRef](#)] [[PubMed](#)]
45. Lane, T.; Fontana, M.; Martinez-Naharro, A.; Quarta, C.C.; Whelan, C.J.; Petrie, A.; Rowczenio, D.M.; Gilbertson, J.A.; Hutt, D.F.; Rezk, T.; et al. Natural History, Quality of Life, and Outcome in Cardiac Transthyretin Amyloidosis. *Circulation* **2019**, *140*, 16–26. [[CrossRef](#)] [[PubMed](#)]
46. Ladefoged, B.; Dybro, A.; Povlsen, J.A.; Vase, H.; Clemmensen, T.S.; Poulsen, S.H. Diagnostic delay in wild type transthyretin cardiac amyloidosis—A clinical challenge. *Int. J. Cardiol.* **2020**, *304*, 138–143. [[CrossRef](#)]
47. Adams, D.; Gonzalez-Duarte, A.; O’Riordan, W.D.; Yang, C.C.; Ueda, M.; Kristen, A.V.; Tournev, I.; Schmidt, H.H.; Coelho, T.; Berk, J.L.; et al. Patisiran, an RNAi Therapeutic, for Hereditary Transthyretin Amyloidosis. *N. Engl. J. Med.* **2018**, *379*, 11–21. [[CrossRef](#)]
48. Bloom, M.W.; Gorevic, P.D. Cardiac Amyloidosis. *Ann. Intern. Med.* **2023**. [[CrossRef](#)]

49. Ruberg, F.L.; Grogan, M.; Hanna, M.; Kelly, J.W.; Maurer, M.S. Transthyretin Amyloid Cardiomyopathy: JACC State-of-the-Art Review. *J. Am. Coll. Cardiol.* **2019**, *73*, 2872. [[CrossRef](#)]
50. Canepa, M.; Fumagalli, C.; Tini, G.; Vincent-Tompkins, J.; Day, S.M.; Ashley, E.A.; Mazzarotto, F.; Ware, J.S.; Michels, M.; Jacoby, D.; et al. Temporal Trend of Age at Diagnosis in Hypertrophic Cardiomyopathy: An Analysis of the International Sarcomeric Human Cardiomyopathy Registry. *Circ. Heart Fail.* **2020**, *13*, e007230. [[CrossRef](#)]
51. Canepa, M.; Vianello, P.F.; Porcari, A.; Merlo, M.; Scarpa, M. Cardiac amyloidosis: A changing epidemiology with open challenges. *Vessel Plus* **2022**, *6*, 30. [[CrossRef](#)]
52. Gillmore, J.D.; Maurer, M.S.; Falk, R.H.; Merlini, G.; Damy, T.; Dispenzieri, A.; Wechalekar, A.D.; Berk, J.L.; Quarta, C.C.; Grogan, M.; et al. Nonbiopsy diagnosis of cardiac transthyretin amyloidosis. *Circulation* **2016**, *133*, 2404–2412. [[CrossRef](#)]
53. Fontana, M.; Chung, R.; Hawkins, P.N.; Moon, J.C. Cardiovascular magnetic resonance for amyloidosis. *Heart Fail. Rev.* **2015**, *20*, 133–144. [[CrossRef](#)]
54. Pandey, T.; Jambhekar, K.; Shaikh, R.; Lensing, S.; Viswamitra, S. Utility of the inversion scout sequence (TI scout) in diagnosing myocardial amyloid infiltration. *Int. J. Cardiovasc. Imaging* **2013**, *29*, 103–112. [[CrossRef](#)]
55. Aquaro, G.D.; Pugliese, N.R.; Perfetto, F.; Cappelli, F.; Barison, A.; Masci, P.G.; Passino, C.; Emdin, M. Myocardial signal intensity decay after gadolinium injection: A fast and effective method for the diagnosis of cardiac amyloidosis. *Int. J. Cardiovasc. Imaging* **2014**, *30*, 1105–1115. [[CrossRef](#)]
56. Fontana, M.; Pica, S.; Reant, P.; Abdel-Gadir, A.; Treibel, T.A.; Banyersad, S.M.; Maestrini, V.; Barcella, W.; Rosmini, S.; Bulluck, H.; et al. Prognostic Value of Late Gadolinium Enhancement Cardiovascular Magnetic Resonance in Cardiac Amyloidosis. *Circulation* **2015**, *132*, 1570–1579. [[CrossRef](#)]
57. Karamitsos, T.D.; Piechnik, S.K.; Banyersad, S.M.; Fontana, M.; Ntusi, N.B.; Ferreira, V.M.; Whelan, C.J.; Myerson, S.G.; Robson, M.D.; Hawkins, P.N.; et al. Noncontrast T1 mapping for the diagnosis of cardiac amyloidosis. *JACC Cardiovasc. Imaging* **2013**, *6*, 488–497. [[CrossRef](#)]
58. Barison, A.; Aquaro, G.D.; Pugliese, N.R.; Cappelli, F.; Chiappino, S.; Vergaro, G.; Mirizzi, G.; Todiere, G.; Passino, C.; Masci, P.G.; et al. Measurement of myocardial amyloid deposition in systemic amyloidosis: Insights from cardiovascular magnetic resonance imaging. *J. Intern. Med.* **2015**, *277*, 605–614. [[CrossRef](#)]
59. Rahman, J.E.; Helou, E.F.; Gelzer-Bell, R.; Thompson, R.E.; Kuo, C.; Rodriguez, E.R.; Hare, J.M.; Baughman, K.L.; Kasper, E.K. Noninvasive diagnosis of biopsy-proven cardiac amyloidosis. *J. Am. Coll. Cardiol.* **2004**, *43*, 410–415. [[CrossRef](#)]
60. Cyrille, N.B.; Goldsmith, J.; Alvarez, J.; Maurer, M.S. Prevalence and prognostic significance of low QRS voltage among the three main types of cardiac amyloidosis. *Am. J. Cardiol.* **2014**, *114*, 1089–1093. [[CrossRef](#)]
61. Rapezzi, C.; Merlini, G.; Quarta, C.C.; Riva, L.; Longhi, S.; Leone, O.; Salvi, F.; Ciliberti, P.; Pastorelli, F.; Biagini, E.; et al. Systemic cardiac amyloidoses: Disease profiles and clinical courses of the 3 main types. *Circulation* **2009**, *120*, 1203–1212. [[CrossRef](#)]
62. Ramanathan, C.; Ghanem, R.; Jia, P.; Ryu, K.; Rudy, Y. Noninvasive electrocardiographic imaging for cardiac electrophysiology and arrhythmia. *Nat. Med.* **2004**, *10*, 422–428. [[CrossRef](#)]
63. Murtagh, B.; Hammill, S.C.; Gertz, M.A.; Kyle, R.A.; Tajik, A.J.; Grogan, M. Electrocardiographic findings in primary systemic amyloidosis and biopsy-proven cardiac involvement. *Am. J. Cardiol.* **2005**, *95*, 535–537. [[CrossRef](#)]
64. Merlo, M.; Porcari, A.; Pagura, L.; Cameli, M.; Vergaro, G.; Musumeci, B.; Biagini, E.; Canepa, M.; Crotti, L.; Imazio, M.; et al. A national survey on prevalence of possible echocardiographic red flags of amyloid cardiomyopathy in consecutive patients undergoing routine echocardiography: Study design and patients characterization—the first insight from the AC-TIVE Study. *Eur. J. Prev. Cardio.* **2021**, *29*, e173–e177. [[CrossRef](#)] [[PubMed](#)]
65. Boldrini, M.; Cappelli, F.; Chacko, L.; Restrepo-Cordoba, M.A.; Lopez-Sainz, A.; Giannoni, A.; Aimo, A.; Baggiano, A.; Martinez-Naharro, A.; Whelan, C.; et al. Multiparametric Echocardiography Scores for the Diagnosis of Cardiac Amyloidosis. *JACC Cardiovasc. Imaging.* **2020**, *13*, 909–920. [[CrossRef](#)] [[PubMed](#)]
66. Wu, Z.W.; Zheng, J.L.; Kaung, L.; Yan, H. Machine learning algorithms to automate differentiating cardiac amyloidosis from hypertrophic cardiomyopathy. *Int. J. Cardiovasc. Imaging* **2022**, *39*, 339–348. [[CrossRef](#)] [[PubMed](#)]
67. Goto, S.; Solanki, D.; John, J.E.; Yagi, R.; Homilies, M.; Ichihara, G.; Katsumata, Y.; Gagging, H.K.; Itabashi, Y.; Macrae, C.A.; et al. Multinational Federated Learning Approach to Train ECG and Echocardiogram Models for Hypertrophic Cardiomyopathy Detection. *Circulation* **2022**, *146*, 755–769. [[CrossRef](#)] [[PubMed](#)]
68. Duffy, G.; Cheng, P.P.; Yuan, N.; He, B.; Kwan, A.C.; Shun-Shin, M.J.; Alexander, K.M.; Ebinger, J.; Lungren, M.P.; Rader, F.; et al. High-Throughput Precision Phenotyping of Left Ventricular Hypertrophy With Cardiovascular Deep Learning. *JAMA Cardiol.* **2022**, *7*, 386–395. [[CrossRef](#)] [[PubMed](#)]
69. Sammani, A.; Jansen, M.; de Vries, N.M.; de Jonge, N.; Baas, A.F.; Te Riele, A.S.J.M.; Asselbergs, F.W.; Oerlemans, M.I.F.J. Automatic Identification of Patients With Unexplained Left Ventricular Hypertrophy in Electronic Health Record Data to Improve Targeted Treatment and Family Screening. *Front. Cardiovasc. Med.* **2022**, *9*, 768847. [[CrossRef](#)] [[PubMed](#)]
70. Davies, D.R.; Redfield, M.M.; Scott, C.G.; Minamisawa, M.; Grogan, M.; Dispenzieri, A.; Chareonthaitawee, P.; Shah, A.M.; Shah, S.J.; Wehbe, R.M.; et al. A Simple Score to Identify Increased Risk of Transthyretin Amyloid Cardiomyopathy in Heart Failure With Preserved Ejection Fraction. *JAMA Cardiol.* **2022**, *7*, 1036–1044. [[CrossRef](#)]
71. Vassilios, V.S.; Cameron, D.; Prasad, S.K.; Gatehouse, P.D. Magnetic resonance imaging: Physics basics for the cardiologist. *JRSM Cardiovasc. Dis.* **2018**, *7*, 2048004018772237.

72. Antonopoulos, A.S.; Almogheer, B.; Azzu, A.; Alati, E.; Papagkikas, P.; Cheong, J.; Clague, J.; Wechalekar, K.; Baksi, J.; Alpendurada, F. Typical and atypical imaging features of cardiac amyloidosis. *Hell. J. Cardiol.* **2021**, *62*, 312–314. [[CrossRef](#)]
73. Kolossváry, M.; Karády, J.; Szilveszter, B.; Kitslaar, P.; Hoffmann, U.; Merkely, B.; Maurovich-Horvat, P. Radiomic Features Are Superior to Conventional Quantitative Computed Tomographic Metrics to Identify Coronary Plaques With Napkin-Ring Sign. *Circ. Cardiovasc. Imaging* **2017**, *10*, e006843. [[CrossRef](#)] [[PubMed](#)]
74. Kolossváry, M.; Kellermayer, M.; Merkely, B.; Maurovich-Horvat, P. Cardiac computed tomography radiomics: A comprehensive review on radiomic techniques. *J. Thorac. Imaging* **2018**, *33*, 26–34. [[CrossRef](#)] [[PubMed](#)]
75. Ioannou, A.; Patel, R.K.; Razvi, Y.; Porcari, A.; Knight, D.; Martinez-Naharro, A.; Kotecha, T.; Venneri, L.; Chacko, L.; Brown, J.; et al. Multi-Imaging Characterization of Cardiac Phenotype in Different Types of Amyloidosis. *JACC Cardiovasc. Imaging*, 2022; *Epub ahead of print*. [[CrossRef](#)] [[PubMed](#)]
76. Li, X.; Li, J.; Lin, L.; Shen, K.; Tian, Z.; Sun, J.; Zhang, C.; An, J.; Jin, Z.; Vliegenthart, R.; et al. Left and right ventricular myocardial deformation and late gadolinium enhancement: Incremental prognostic value in amyloid light-chain amyloidosis. *Cardiovasc. Diagn. Ther.* **2020**, *10*, 470–480. [[CrossRef](#)]
77. Giusca, S.; Steen, H.; Montenbruck, M.; Patel, A.R.; Pieske, B.; Erley, J.; Kelle, S.; Korosoglou, G. Multi-parametric assessment of left ventricular hypertrophy using late gadolinium enhancement, T1 mapping and strain-encoded cardiovascular magnetic resonance. *J. Cardiovasc. Magn. Reson.* **2021**, *23*, 92. [[CrossRef](#)]
78. Mohty, D.; Pradel, S.; Magne, J.; Fadel, B.; Boulogne, C.; Petitalot, V.; Raboukhi, S.; Darodes, N.; Damy, T.; Aboyans, V.; et al. Prevalence and prognostic impact of left-sided valve thickening in systemic light-chain amyloidosis. *Clin. Res. Cardiol.* **2016**, *106*, 331–340. [[CrossRef](#)]
79. Steen, H.; Giusca, S.; Montenbruck, M.; Patel, A.R.; Pieske, B.; Florian, A.; Erley, J.; Kelle, S.; Korosoglou, G. Left and right ventricular strain using fast strain-encoded cardiovascular magnetic resonance for the diagnostic classification of patients with chronic non-ischemic heart failure due to dilated, hypertrophic cardiomyopathy or cardiac amyloidosis. *J. Cardiovasc. Magn. Reson.* **2021**, *23*, 45. [[CrossRef](#)]
80. Sciacca, V.; Eckstein, J.; Körperich, H.; Fink, T.; Bergau, L.; El Hamriti, M.; Imnadze, G.; Guckel, D.; Fox, H.; Gerçek, M.; et al. Magnetic-Resonance-Imaging-Based Left Atrial Strain and Left Atrial Strain Rate as Diagnostic Parameters in Cardiac Amyloidosis. *J. Clin. Med.* **2022**, *11*, 3150. [[CrossRef](#)]
81. Nemes, A.; Földeák, D.; Domsik, P.; Kalapos, A.; Kormányos, Á.; Borbényi, Z.; Forster, T. Right Atrial Deformation Analysis in Cardiac Amyloidosis—Results from the Three-Dimensional Speckle-Tracking Echocardiographic MAGYAR-Path Study. *Arq. Bras. Cardiol.* **2018**, *111*, 384–391. [[CrossRef](#)]
82. Higashi, H.; Inoue, K.; Inaba, S.; Nakao, Y.; Kinoshita, M.; Miyazaki, S.; Miyoshi, T.; Akazawa, Y.; Kawakami, H.; Uetani, T.; et al. Restricted left atrial dilatation can visually differentiate cardiac amyloidosis from hypertrophic cardiomyopathy. *ESC Heart Fail.* **2021**, *8*, 3198–3205. [[CrossRef](#)]
83. Liu, H.; Bai, P.; Xu, H.-Y.; Li, Z.-L.; Xia, C.-C.; Zhou, X.-Y.; Gong, L.-G.; Guo, Y.-K. Distinguishing Cardiac Amyloidosis and Hypertrophic Cardiomyopathy by Thickness and Myocardial Deformation of the Right Ventricle. *Cardiol. Res. Pract.* **2022**, *2022*, 4364279. [[CrossRef](#)]
84. Nochioka, K.; Quarta, C.C.; Claggett, B.; Roca, G.Q.; Rapezzi, C.; Falk, R.H.; Solomon, S.D. Left atrial structure and function in cardiac amyloidosis. *Eur. Heart J. Cardiovasc. Imaging* **2017**, *18*, 1128–1137. [[CrossRef](#)] [[PubMed](#)]
85. Eckstein, J.; Moghadasi, N.; Körperich, H.; Weise Valdés, E.; Sciacca, V.; Paluszkiwicz, L.; Burchert, W.; Piran, M. A Machine Learning Challenge: Detection of Cardiac Amyloidosis Based on Bi-Atrial and Right Ventricular Strain and Cardiac Function. *Diagnostics* **2022**, *12*, 2693. [[CrossRef](#)] [[PubMed](#)]
86. Kellman, P.; Hansen, M.S. T1-mapping in the heart: Accuracy and precision. *J. Cardiovasc. Magn. Reson.* **2014**, *16*, 2. [[CrossRef](#)] [[PubMed](#)]
87. Giri, S.; Chung, Y.C.; Merchant, A.; Mihai, G.; Rajagopalan, S.; Raman, S.V.; Simonetti, O.P. T2 quantification for improved detection of myocardial edema. *J. Cardiovasc. Magn. Reson.* **2009**, *11*, 56. [[CrossRef](#)] [[PubMed](#)]
88. Kellman, P.; Hernando, D.; Arai, A.E. Myocardial fat imaging. *Curr. Cardiovasc. Imaging Rep.* **2010**, *3*, 83–91. [[CrossRef](#)] [[PubMed](#)]
89. Xue, H.; Brown, L.A.E.; Nielles-Vallespin, S.; Plein, S.; Kellman, P. Automatic in-line quantitative myocardial perfusion mapping: Processing algorithm and implementation. *Magn. Reson. Med.* **2020**, *83*, 712–730. [[CrossRef](#)]
90. Kellman, P.; Hansen, M.S.; Nielles-Vallespin, S.; Nickander, J.; Themudo, R.; Ugander, M.; Xue, H. Myocardial perfusion cardiovascular magnetic resonance: Optimized dual sequence and reconstruction for quantification. *J. Cardiovasc. Magn. Reson.* **2017**, *19*, 43. [[CrossRef](#)]
91. Nickander, J.; Themudo, R.; Sigfridsson, A.; Xue, H.; Kellman, P.; Ugander, M. Females have higher myocardial perfusion, blood volume and extracellular volume compared to males: An adenosine stress cardiovascular magnetic resonance study. *Sci. Rep.* **2020**, *10*, 10380. [[CrossRef](#)]
92. Schulz-Menger, J.; Bluemke, D.A.; Bremerich, J.; Flamm, S.D.; Fogel, M.A.; Friedrich, M.G.; Kim, R.J.; von Knobelsdorff-Brenkenhoff, F.; Kramer, C.M.; Pennell, D.J.; et al. Standardized image interpretation and post-processing in cardiovascular magnetic resonance: 2020 update—Society for Cardiovascular Magnetic Resonance (SCMR): Board of Trustees Task Force on Standardized Post-Processing. *J. Cardiovasc. Magn. Reson.* **2020**, *22*, 19. [[CrossRef](#)]

93. Bai, W.; Sinclair, M.; Tarroni, G.; Oktay, O.; Rajchl, M.; Vaillant, G.; Lee, A.M.; Aung, N.; Lukaschuk, E.; Sanghvi, M.M.; et al. Automated cardiovascular magnetic resonance image analysis with fully convolutional networks. *J. Cardiovasc. Magn. Reson.* **2018**, *20*, 65. [[CrossRef](#)] [[PubMed](#)]
94. Bernard, O.; Lalande, A.; Zotti, C.; Cervenansky, F.; Yang, X.; Heng, P.A.; Cetin, I.; Lekadir, K.; Camara, O.; Gonzalez Ballester, M.A.; et al. Deep learning techniques for automatic MRI cardiac multi-structures segmentation and diagnosis: Is the problem solved? *IEEE Trans. Med. Imaging* **2018**, *37*, 2514–2525. [[CrossRef](#)] [[PubMed](#)]
95. Xue, H.; Davies, R.H.; Brown, L.A.E.; Knott, K.D.; Kotecha, T.; Fontana, M.; Plein, S.; Moon, J.C.; Kellman, P. Automated inline analysis of myocardial perfusion MRI with deep learning. *Radiol. Artif. Intell.* **2020**, *2*, e200009. [[CrossRef](#)]
96. Tao, Q.; Yan, W.; Wang, Y.; Paiman, E.H.M.; Shamonin, D.P.; Garg, P.; Plein, S.; Huang, L.; Xia, L.; Sramko, M.; et al. Deep learning-based method for fully automatic quantification of left ventricle function from cine MR images: A multivendor, multicenter study. *Radiology* **2019**, *290*, 81–88. [[CrossRef](#)] [[PubMed](#)]
97. Davies, R.H.; Augusto, J.B.; Bhuva, A.; Xue, H.; Treibel, T.A.; Ye, Y.; Hughes, R.K.; Bai, W.; Lau, C.; Shiwani, H.; et al. Precision measurement of cardiac structure and function in cardiovascular magnetic resonance using machine learning. *J. Cardiovasc. Magn. Reson.* **2022**, *24*, 16. [[CrossRef](#)]
98. Xue, H.; Artico, J.; Fontana, M.; Moon, J.C.; Davies, R.H.; Kellman, P. Landmark Detection in Cardiac MRI by Using a Convolutional Neural Network. *Radiology: Artif. Intell.* **2021**, *3*, e200197. [[CrossRef](#)]
99. Jiang, S.; Zhang, L.; Wang, J.; Li, X.; Hu, S.; Fu, Y.; Wang, X.; Hao, S.; Hu, C. Differentiating between cardiac amyloidosis and hypertrophic cardiomyopathy on non-contrast cine-magnetic resonance images using machine learning-based radiomics. *Front. Cardiovasc. Med.* **2022**, *9*, 1001269. [[CrossRef](#)]
100. Antonopoulos, A.S.; Boutsikou, M.; Simantiris, S.; Angelopoulos, A.; Lazaros, G.; Panagiotopoulos, I.; Oikonomou, E.; Kanoupaki, M.; Tousoulis, D.; Mohiaddin, R.H.; et al. Machine learning of native T1 mapping radiomics for classification of hypertrophic cardiomyopathy phenotypes. *Sci. Rep.* **2021**, *11*, 23596. [[CrossRef](#)]
101. Martini, N.; Aimo, A.; Barison, A.; Della Latta, D.; Vergaro, G.; Aquaro, G.D.; Ripoli, A.; Emdin, M.; Chiappino, D. Deep learning to diagnose cardiac amyloidosis from cardiovascular magnetic resonance. *J. Cardiovasc. Magn. Reson.* **2020**, *22*, 84. [[CrossRef](#)]
102. Satriano, A.; Afzal, Y.; Sarim Afzal, M.; Fatehi Hassanabad, A.; Wu, C.; Dykstra, S.; Flewitt, J.; Feuchter, P.; Sandomato, R.; Heydari, B.; et al. Neural-Network-Based Diagnosis Using 3-Dimensional Myocardial Architecture and Deformation: Demonstration for the Differentiation of Hypertrophic Cardiomyopathy. *Front. Cardiovasc. Med.* **2020**, *7*, 584727. [[CrossRef](#)]
103. Moharram, M.A.; Lamberts, R.R.; Whalley, G.; Williams, M.J.A.; Coffey, S. Myocardial tissue characterisation using echocardiographic deformation imaging. *Cardiovasc. Ultrasound* **2019**, *17*, 27. [[CrossRef](#)] [[PubMed](#)]
104. Haland, T.F.; Almaas, V.M.; Hasselberg, N.E.; Saberniak, J.; Leren, I.S.; Hopp, E.; Edvardsen, T.; Haugaa, K.H. Strain echocardiography is related to fibrosis and ventricular arrhythmias in hypertrophic cardiomyopathy. *Eur. Heart J. Cardiovasc. Imaging* **2016**, *17*, 613–621. [[CrossRef](#)] [[PubMed](#)]
105. Popovic, Z.B.; Kwon, D.H.; Mishra, M.; Buakhamsri, A.; Greenberg, N.L.; Thamilarasan, M.; Flamm, S.D.; Thomas, J.D.; Lever, H.M.; Desai, M.Y. Association between regional ventricular function and myocardial fibrosis in hypertrophic cardiomyopathy assessed by speckle tracking echocardiography and delayed hyperenhancement magnetic resonance imaging. *J. Am. Soc. Echocardiogr.* **2008**, *21*, 1299–1305. [[CrossRef](#)]
106. Delgado, V.; Marsan, N.A. Global and regional longitudinal strain assessment in hypertrophic cardiomyopathy. *Circ. Cardiovasc. Imaging* **2019**, *12*, e009586. [[CrossRef](#)] [[PubMed](#)]
107. Pagourelas, E.D.; Mirea, O.; Duchenne, J.; Van Cleemput, J.; Delforge, M.; Bogaert, J.; Kuznetsova, T.; Voigt, J.U. Echo parameters for differential diagnosis in cardiac amyloidosis: A head-to-head comparison of deformation and nondeformation parameters. *Circ. Cardiovasc. Imaging* **2017**, *10*, e005588. [[CrossRef](#)]
108. Gotschy, A.; Von Deuster, C.; Van Gorkum, R.J.H.; Gastl, M.; Vintschger, E.; Schwotzer, R.; Flammer, A.J.; Manka, R.; Stoeck, C.T.; Kozerke, S. Characterizing cardiac involvement in amyloidosis using cardiovascular magnetic resonance diffusion tensor imaging. *J. Cardiovasc. Magn. Reson.* **2019**, *21*, 56. [[CrossRef](#)]
109. Satoskar, A.A.; Efebera, Y.; Hasan, A.; Brodsky, S.; Nadasdy, G.; Dogan, A.; Nadasdy, T. Strong transthyretin immunostaining: Potential pitfall in cardiac amyloid typing. *Am. J. Surg. Pathol.* **2011**, *35*, 1685–1690. [[CrossRef](#)]
110. Solomon, A.; Murphy, C.L.; Westermarck, P. Unreliability of immunohistochemistry for typing amyloid deposits. *Arch. Pathol. Lab. Med.* **2008**, *132*, 14. [[CrossRef](#)]
111. Gilbertson, J.A.; Theis, J.D.; Vrana, J.A.; Lachmann, H.; Wechalekar, A.; Whelan, C.; Hawkins, P.N.; Dogan, A.; Gillmore, J.D. A comparison of immunohistochemistry and mass spectrometry for determining the amyloid fibril protein from formalin-fixed biopsy tissue. *J. Clin. Pathol.* **2015**, *68*, 314–317. [[CrossRef](#)]
112. Lachmann, H.; Booth, D.R.; Booth, S.E.; Bybee, A.; Gilbertson, J.A.; Gillmore, J.D.; Pepys, M.B.; Hawkins, P.N. Misdiagnosis of Hereditary Amyloidosis as AL (Primary) Amyloidosis. *N. Engl. J. Med.* **2002**, *346*, 1786–1791. [[CrossRef](#)]
113. Satoskar, A.A.; Burdge, K.; Cowden, D.J.; Nadasdy, G.M.; Hebert, L.A.; Nadasdy, T. Typing of amyloidosis in renal biopsies: Diagnostic pitfalls. *Arch. Pathol. Lab. Med.* **2007**, *131*, 917–922. [[CrossRef](#)] [[PubMed](#)]
114. Schönland, S.O.; Hegenbart, U.; Bochtler, T.; Mangatter, A.; Hansberg, M.; Ho, A.D.; Lohse, P.; Röcken, C. Immunohistochemistry in the classification of systemic forms of amyloidosis: A systematic investigation of 117 patients. *Blood* **2012**, *119*, 488–493. [[CrossRef](#)] [[PubMed](#)]

115. Abildgaard, N.; Rojek, A.M.; Møller, H.E.; Palstrøm, N.B.; Nyvold, C.G.; Rasmussen, L.M.; Hansen, C.T.; Beck, H.C.; Marcussen, N. Immunoelectron microscopy and mass spectrometry for classification of amyloid deposits. *Amyloid* **2020**, *27*, 59–66. [CrossRef] [PubMed]
116. Vrana, J.A.; Gamez, J.D.; Madden, B.J.; Theis, J.D.; Bergen, H.R., 3rd; Dogan, A. Classification of amyloidosis by laser microdissection and mass spectrometry-based proteomic analysis in clinical biopsy specimens. *Blood* **2009**, *114*, 4957–4959. [CrossRef]
117. Brambilla, F.; Lavatelli, F.; Di Silvestre, D.; Valentini, V.; Rossi, R.; Palladini, G.; Obici, L.; Verga, L.; Mauri, P.; Merlini, G. Reliable typing of systemic amyloidoses through proteomic analysis of subcutaneous adipose tissue. *Blood* **2012**, *119*, 1844–1847. [CrossRef]
118. Palstrøm, N.B.; Rojek, A.M.; Møller, H.E.H.; Hansen, C.T.; Matthiesen, R.; Rasmussen, L.M.; Abildgaard, N.; Beck, H.C. Classification of Amyloidosis by Model-Assisted Mass Spectrometry-Based Proteomics. *Int. J. Mol. Sci.* **2021**, *23*, 319. [CrossRef]
119. Leguit, R.J.; Vink, A.; de Jonge, N.; Minnema, M.C.; Oerlemans, M.I.F. Endomyocardial biopsy with co-localization of a lymphoplasmacytic lymphoma and AL amyloidosis. *Cardiovasc. Pathol.* **2021**, *53*, 107348. [CrossRef]
120. Winburn, I.; Ishii, T.; Sumikawa, T.; Togo, K.; Yasunaga, H. Estimating the Prevalence of Transthyretin Amyloid Cardiomyopathy in a Large In-Hospital Database in Japan. *Cardiol. Ther.* **2019**, *8*, 297–316. [CrossRef]
121. Isogai, T.; Yasunaga, H.; Matsui, H.; Ueda, T.; Tanaka, H.; Horiguchi, H.; Fushimi, K. Hospital volume and cardiac complications of endomyocardial biopsy: A retrospective cohort study of 9508 adult patients using a nationwide inpatient database in Japan. *Clin. Cardiol.* **2015**, *38*, 164–170. [CrossRef]
122. Statistics Bureau, Ministry of Internal Affairs and Communications Japan. Monthly Report, 2018. Available online: <http://www.stat.go.jp/english/data/jinsui/tsuki/index.html> (accessed on 13 March 2019).
123. Tsuchihashi-Makaya, M.; Hamaguchi, S.; Kinugawa, S.; Yokota, T.; Goto, D.; Yokoshiki, H.; Kato, N.; Takeshita, A.; Tsutsui, H.; JCARE-CARD Investigators. Characteristics and outcomes of hospitalized patients with heart failure and reduced vs preserved ejection fraction. Report from the Japanese Cardiac Registry of Heart Failure in Cardiology (JCARE-CARD). *Circ. J.* **2009**, *73*, 1893–1900. [CrossRef]
124. Huda, A.; Castaño, A.; Niyogi, A.; Schumacher, J.; Stewart, M.; Bruno, M.; Hu, M.; Ahmad, F.S.; Deo, R.C.; Shah, S.J. A machine learning model for identifying patients at risk for wild-type transthyretin amyloid cardiomyopathy. *Nat Commun.* **2021**, *12*, 2725. [CrossRef] [PubMed]
125. Agibetov, A.; Seirer, B.; Dachs, T.M.; Koschutnik, M.; Dalos, D.; Rettl, R.; Duca, F.; Schrutka, L.; Agis, H.; Kain, R.; et al. Machine Learning Enables Prediction of Cardiac Amyloidosis by Routine Laboratory Parameters: A Proof-of-Concept Study. *J. Clin. Med.* **2020**, *9*, 1334. [CrossRef] [PubMed]
126. Makin, O.S.; Serpell, L.C. Structures for amyloid fibrils. *FEBS J.* **2005**, *272*, 5950–5961. [CrossRef]
127. Cao, Y.; Tang, X.; Yuan, M.; Han, W. Computational studies of protein aggregation mediated by amyloid: Fibril elongation and secondary nucleation. *Prog. Mol. Biol. Transl. Sci.* **2020**, *170*, 461–504. [CrossRef] [PubMed]
128. Tian, J.; Wu, N.; Guo, J.; Fan, Y. Prediction of amyloid fibril-forming segments based on a support vector machine. *BMC Bioinform.* **2009**, *10* (Suppl. S1), S45. [CrossRef]
129. Kawashima, S.; Kanehisa, M. Aaindex: Amino acid index database. *Nucleic Acids Res.* **2000**, *28*, 374. [CrossRef]
130. Liu, B.; Liu, F.; Wang, X.; Chen, J.; Fang, L.; Chou, K.-C. Pse-in-one: A web server for generating various modes of pseudo components of DNA, RNA, and protein sequences. *Nucleic Acids Res.* **2015**, *43*, 65–71. [CrossRef] [PubMed]
131. Li, Y.; Zhang, Z.; Teng, Z.; Liu, X. Predamyl-mlp: Prediction of amyloid proteins using multilayer perceptron. *Comput. Math. Methods Med.* **2020**. [CrossRef]
132. Teng, Z.; Zhang, Z.; Tian, Z.; Li, Y.; Wang, G. ReRF-Pred: Predicting amyloidogenic regions of proteins based on their pseudo amino acid composition and tripeptide composition. *BMC Bioinform.* **2021**, *22*, 545. [CrossRef]
133. Chou, K.-C. Pseudo amino acid composition and its applications in bioinformatics, proteomics and system biology. *Curr. Proteom.* **2009**, *6*, 262–274. [CrossRef]
134. Liao, B.; Jiang, J.-B.; Zeng, Q.-G.; Zhu, W. Predicting apoptosis protein subcellular location with pseaac by incorporating tripeptide composition. *Protein Pept. Lett.* **2011**, *18*, 1086–1092. [CrossRef] [PubMed]
135. Burdukiewicz, M.; Sobczyk, P.; Rödiger, S.; Duda-Madej, A.; Mackiewicz, P.; Kotulska, M. Amyloidogenic motifs revealed by n-gram analysis. *Sci. Rep.* **2017**, *7*, 12961. [CrossRef] [PubMed]
136. Keresztes, L.; Szögi, E.; Varga, B.; Farkas, V.; Perczel, A.; Grolmusz, V. The budapest amyloid predictor and its applications. *Biomolecules* **2021**, *11*, 500. [CrossRef] [PubMed]
137. Beerten, J.; Van Durme, J.; Gallardo, R.; Capriotti, E.; Serpell, L.; Rousseau, F.; Schymkowitz, J. WALTZ-DB: A benchmark database of amyloidogenic hexapeptides. *Bioinformatics* **2015**, *31*, 1698–1700. [CrossRef]
138. Auriemma Citarella, A.; Di Biasi, L.; Risi, M.; Tortora, G. Snarer: New molecular descriptors for snare proteins classification. *BMC Bioinform.* **2022**, *23*, 148. [CrossRef]
139. Auriemma Citarella, A.; Di Biasi, L.; De Marco, F.; Tortora, G. ENTAIL: YET aNoTher amyloid fibrils cLassifier. *BMC Bioinform.* **2022**, *23*, 517. [CrossRef]
140. Dimopoulos, M.A.; Alexanian, R. Waldenstrom's macroglobulinemia. *Blood* **1994**, *83*, 1452–1459. [CrossRef]
141. Lee, C.C.; Perchiacca, J.M.; Tessier, P.M. Toward aggregation-resistant antibodies by design. *Trends Biotechnol.* **2013**, *31*, 612–620. [CrossRef]
142. Tangakani, A.M.; Kumar, S.; Nagarajan, R.; Velmurugan, D.; Gromiha, M.M. GAP: Towards almost 100 percent prediction for β -strand-mediated aggregating peptides with distinct morphologies. *Bioinformatics* **2014**, *30*, 1983–1990. [CrossRef]

143. Fernandez-Escamilla, A.M.; Rousseau, F.; Schymkowitz, J.; Serrano, L. Prediction of sequence-dependent and mutational effects on the aggregation of peptides and proteins. *Nat. Biotechnol.* **2004**, *22*, 1302–1306. [[CrossRef](#)]
144. Maurer-Stroh, S.; Debulpaep, M.; Kuemmerer, N.; Lopez de la Paz, M.; Martins, I.C.; Reumers, J.; Morris, K.L.; Copland, A.; Serpell, L.; Serrano, L.; et al. Exploring the sequence determinants of amyloid structure using position-specific scoring matrices. *Nat. Methods* **2010**, *7*, 237–242. [[CrossRef](#)] [[PubMed](#)]
145. Trovato, A.; Seno, F.; Tosatto, S.C. Te PASTA server for protein aggregation prediction. *Protein Eng. Des. Sel.* **2007**, *20*, 521–523. [[CrossRef](#)] [[PubMed](#)]
146. Conchillo-Solé, O.; de Groot, N.S.; Avilés, F.X.; Vendrell, J.; Daura, X.; Ventura, S. AGGRESCAN: A server for the prediction and evaluation of “hot spots” of aggregation in polypeptides. *BMC Bioinform.* **2007**, *8*, 65. [[CrossRef](#)] [[PubMed](#)]
147. Garbuzynskiy, S.O.; Lobanov, M.Y.; Galzitskaya, O.V. FoldAmyloid: A method of prediction of amyloidogenic regions from protein sequence. *Bioinformatics* **2010**, *26*, 326–332. [[CrossRef](#)] [[PubMed](#)]
148. Prabakaran, R.; Rawat, P.; Kumar, S.; Gromiha, M.M. ANuPP: A versatile tool to predict aggregation nucleating regions in peptides and proteins. *J. Mol. Biol.* **2020**, *1*, 166707. [[CrossRef](#)]
149. Prabakaran, R.; Rawat, P.; Tangakani, A.M.; Kumar, S.; Gromiha, M.M. Protein aggregation: In silico algorithms and applications. *Biophys. Rev.* **2021**, *13*, 71–89. [[CrossRef](#)]
150. Buck, P.M.; Kumar, S.; Wang, X.; Agrawal, N.J.; Trout, B.L.; Singh, S.K. Computational methods to predict therapeutic protein aggregation. *Methods Mol. Biol.* **2012**, *899*, 425–451.
151. Buck, P.M.; Kumar, S.; Singh, S.K. On the role of aggregation prone regions in protein evolution, stability, and enzymatic catalysis: Insights from diverse analyses. *PLoS Comput. Biol.* **2013**, *9*, e1003291. [[CrossRef](#)]
152. Prabakaran, R.; Goel, D.; Kumar, S.; Gromiha, M.M. Aggregation prone regions in human proteome: Insights from large-scale data analyses. *Proteins* **2017**, *85*, 1099–1118. [[CrossRef](#)]
153. Rawat, P.; Kumar, S.; Gromiha, M.M. An in-silico method for identifying aggregation rate enhancer and mitigator mutations in proteins. *Int. J. Biol. Macromol.* **2018**, *118*, 1157–1167. [[CrossRef](#)]
154. Rawat, P.; Prabakaran, R.; Kumar, S.; Gromiha, M.M. AggreRATE-Pred: A mathematical model for the prediction of change in aggregation rate upon point mutation. *Bioinformatics* **2020**, *36*, 1439–1444. [[CrossRef](#)]
155. Rawat, P.; Prabakaran, R.; Sakthivel, R.; Mary Thangakani, A.; Kumar, S.; Gromiha, M.M. CPAD 20: A repository of curated experimental data on aggregating proteins and peptides. *Amyloid* **2020**, *27*, 128–133. [[CrossRef](#)] [[PubMed](#)]
156. Rawat, P.; Prabakaran, R.; Kumar, S.; Gromiha, M.M. AbsoluRATE: An in-silico method to predict the aggregation kinetics of native proteins. *BBA-Proteins Proteom.* **2021**, *1*, 140682. [[CrossRef](#)] [[PubMed](#)]
157. Briney, B.; Inderbitzin, A.; Joyce, C.; Burton, D.R. Commonality despite exceptional diversity in the baseline human antibody repertoire. *Nature* **2019**, *566*, 393–397. [[CrossRef](#)] [[PubMed](#)]
158. David, M.P.C.; Concepcion, G.P.; Padlan, E.A. Using simple artificial intelligence methods for predicting amyloidogenesis in antibodies. *BMC Bioinform.* **2010**, *11*, 79. [[CrossRef](#)]
159. Rawat, P.; Prabakaran, R.; Kumar, S.; Gromiha, M.M. Exploring the sequence features determining amyloidosis in human antibody light chains. *Sci Rep.* **2021**, *11*, 13785. [[CrossRef](#)]
160. Bonnefous, L.; Kharoubi, M.; Bézard, M.; Oghina, S.; Le Bras, F.; Pouillot, E.; Molinier-Frenkel, V.; Fanen, P.; Deux, J.F.; Audard, V.; et al. Assessing Cardiac Amyloidosis Subtypes by Unsupervised Phenotype Clustering Analysis. *J. Am. Coll. Cardiol.* **2021**, *78*, 2177–2192. [[CrossRef](#)]
161. Ruberg, F.L. Phenotype Mapping in Cardiac Amyloidosis. *J. Am. Coll. Cardiol.* **2021**, *78*, 2193–2195. [[CrossRef](#)]
162. Poterucha, T.J.; Elias, P.; Ruberg, F.L.; DeLuca, A.; Kinkhabwala, M.; Johnson, L.L.; Griffin, J.M.; Pandey, S.; Einstein, A.J.; Maurer, M.S. False Positive 99mTc-Pyrophosphate Scanning Leading to Inappropriate Tafamidis Prescriptions. *JACC Cardiovasc. Imaging* **2021**, *14*, 2042–2044. [[CrossRef](#)]
163. Hanna, M.; Ruberg, F.L.; Maurer, M.S.; Dispenzieri, A.; Dorbala, S.; Falk, R.H.; Hoffman, J.; Jaber, W.; Soman, P.; Witteles, R.M.; et al. Cardiac Scintigraphy With Technetium-99m-Labeled Bone-Seeking Tracers for Suspected Amyloidosis: JACC Review Topic of the Week. *J. Am. Coll. Cardiol.* **2020**, *75*, 2851–2862. [[CrossRef](#)]
164. Hughes, S.E. The pathology of hypertrophic cardiomyopathy. *Histopathology* **2004**, *44*, 412–427. [[CrossRef](#)] [[PubMed](#)]
165. López-Cuenca, D.; Muñoz-Esparza, C.; Peñalver, M.N.; Alberola, A.G.; Blanes, J.R.G. Hypertrophic or hypertensive cardiomyopathy? *Int. J. Cardiol.* **2016**, *203*, 891–892. [[CrossRef](#)] [[PubMed](#)]
166. Maron, M.S.; Rowin, E.J.; Maron, B.J. How to image hypertrophic cardiomyopathy. *Circ. Cardiovasc. Imaging* **2017**, *10*, e005372. [[CrossRef](#)]
167. Lemery, R.; Kleinebenne, A.; Nihoyannopoulos, P.; Aber, V.; Alfonso, F.; McKenna, W.J. Q waves in hypertrophic cardiomyopathy in relation to the distribution and severity of right and left ventricular hypertrophy. *J. Am. Coll. Cardiol.* **1990**, *16*, 368–374. [[CrossRef](#)] [[PubMed](#)]
168. Maron, B.J.; Wolfson, J.K.; Ciró, E.; Spirito, P. Relation of electrocardiographic abnormalities and patterns of left ventricular hypertrophy identified by 2-dimensional echocardiography in patients with hypertrophic cardiomyopathy. *Am. J. Cardiol.* **1983**, *51*, 189–194. [[CrossRef](#)]
169. Usui, M.; Inoue, H.; Suzuki, J.; Watanabe, F.; Sugimoto, T.; Nishikawa, J. Relationship between distribution of hypertrophy and electrocardiographic changes in hypertrophic cardiomyopathy. *Am. Heart J.* **1993**, *126*, 177–183. [[CrossRef](#)]

170. Tison, G.H.; Zhang, J.; Delling, F.N.; Deo, R.C. Automated and interpretable patient ECG profiles for disease detection, tracking, and discovery. *Circ. Cardiovasc. Qual. Outcomes* **2019**, *12*, e005289. [[CrossRef](#)]
171. Morita, S.X.; Kusunose, K.; Haga, A.; Sata, M.; Hasegawa, K.; Raita, Y.; Reilly, M.P.; Fifer, M.A.; Maurer, M.S.; Shimada, Y.J. Deep learning analysis of echocardiographic images to predict positive genotype in patients with hypertrophic cardiomyopathy. *Front. Cardiovasc. Med.* **2021**, *8*, 669860. [[CrossRef](#)]
172. Ko, W.-Y.; Siontis, K.C.; Attia, Z.I.; Carter, R.E.; Kapa, S.; Ommen, S.R.; Demuth, S.J.; Ackerman, M.J.; Gersh, B.J.; Arruda-Olson, A.M.; et al. Detection of hypertrophic cardiomyopathy using a convolutional neural network-enabled electrocardiogram. *J. Am. Coll. Cardiol.* **2020**, *75*, 722–733. [[CrossRef](#)]
173. Zhang, J.; Gajjala, S.; Agrawal, P.; Tison, G.H.; Hallock, L.A.; Beussink-Nelson, L.; Lassen, M.H.; Fan, E.; Aras, M.A.; Jordan, C.; et al. Fully automated echocardiogram interpretation in clinical practice. *Circulation* **2018**, *138*, 1623–1635. [[CrossRef](#)]
174. Wasfy, J.H.; Singal, G.; O'Brien, C.; Blumenthal, D.M.; Kennedy, K.F.; Strom, J.B.; Spertus, J.A.; Mauri, L.; Normand, S.L.T.; Yeh, R.V. Enhancing the prediction of 30-day readmission after percutaneous coronary intervention using data extracted by querying of the electronic health record. *Circ. Cardiovasc. Qual. Outcomes* **2015**, *8*, 477–485. [[CrossRef](#)] [[PubMed](#)]
175. García-García, E.; González-Romero, G.M.; Martín-Pérez, E.M.; Zapata Cornejo, E.D.; Escobar-Aguilar, G.; Cárdenas Bonnet, M.F. Real-World Data and Machine Learning to Predict Cardiac Amyloidosis. *Int. J. Environ. Res. Public Health* **2021**, *18*, 908. [[CrossRef](#)] [[PubMed](#)]
176. Subbiah, V. The next generation of evidence-based medicine. *Nat. Med.* **2023**, *29*, 49–58. [[CrossRef](#)]
177. Maurer, M.S.; Schwartz, J.H.; Gundapaneni, B.; Elliott, P.M.; Merlini, G.; Waddington-Cruz, M.; Kristen, A.V.; Grogan, M.; Witteles, R.; Damy, T.; et al. Tafamidis Treatment for Patients with Transthyretin Amyloid Cardiomyopathy. *N. Engl. J. Med.* **2018**, *379*, 1007–1016. [[CrossRef](#)] [[PubMed](#)]
178. Falk, R.H.; Alexander, K.M.; Liao, R.; Dorbala, S. AL (Light-Chain) Cardiac Amyloidosis: A Review of Diagnosis and Therapy. *J. Am. Coll. Cardiol.* **2016**, *68*, 1323–1341. [[CrossRef](#)] [[PubMed](#)]
179. Comenzo, R.L. Out, Out—Making Amyloid's Candle Briefer. *N. Engl. J. Med.* **2015**, *373*, 1167–1169. [[CrossRef](#)] [[PubMed](#)]
180. Rossi, M.; Varrà, G.G.; Porcari, A.; Saro, R.; Pagura, L.; Lalario, A.; Dore, F.; Bussani, R.; Sinagra, G.; Merlo, M. Re-Definition of the Epidemiology of Cardiac Amyloidosis. *Biomedicines* **2022**, *10*, 1566. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.