



**Chromatographic analysis of VOC patterns in exhaled breath  
from smokers and nonsmokers**

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Keywords:	SPME-GC/MS, breath analysis, exhaled VOCs, smoking biomarkers

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3 **Chromatographic analysis of VOC patterns**  
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5 **in exhaled breath from smokers and nonsmokers**  
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31 **Keywords:** breath analysis, exhaled VOCs, smoking biomarkers, SPME-GC/MS  
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40 **ABSTRACT**  
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42 Cigarette smoking harms nearly every organ of the body and causes many diseases. The analysis of  
43 exhaled breath for Volatile Organic Compounds (VOCs) can provide fundamental information on  
44 active smoking and insight into the health damage that smoke is creating. Various exhaled (VOCs)  
45 have been reported as typical of smoking habit and recent tobacco consumption, but to date, no  
46 eligible biomarkers have been identified. Aiming to identify such potential biomarkers, in this pilot  
47 study we analysed the chemical patterns of exhaled breath from 26 volunteers divided in  
48 nonsmokers and smokers sampled at different periods of withdrawal from smoking. SPME-GC/MS  
49 method were applied.  
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3 Many breath VOCs were identified and quantified in very low concentrations, but only a few  
4 (toluene, pyridine, pyrrole, benzene, 2-butanone, 2-pentanone, 1-methyldecylamine) were found to  
5 be statistically significant variables. Probit prediction model based on statistical relevant VOCs-  
6 patterns (instead of individual VOCs) showed that the assessment of smoking status is heavily time  
7 dependent; it's possible recognise with high specificity and sensitivity smokers after a short-term  
8 exposure to tobacco (i.e. after 1 hour of smoking abstinence), whereas smokers after a long-term  
9 exposure to tobacco (i.e. after a night out of smoking) are more like nonsmokers.  
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## 20 **1. Introduction**

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22 Smoking is the inhalation of tobacco smoke containing over 4000 chemical compounds  
23 including toxic and carcinogenic constituents (Rodgman et al. 2013; Hecht, 2011). The harmful  
24 effects of tobacco smoking both for active and passive smokers are well known; active smoking is  
25 the main risk factor and independent predictor of short survival in lung cancer, and involuntary  
26 exposure to environmental tobacco smoke, i.e. passive smoking, has been extensively investigated  
27 with respect to its potential health effects (U.S. Dept. of Health and Human Services, 2014; Hakim  
28 et al., 2012; Hecht, 2011). Many more Volatile Organic Compounds (VOCs) can be found in the  
29 exhaled breath of smoking and passive smoking persons than in breath samples from nonsmokers.  
30 The origin of these VOCs is mainly exogenous due to the inhalation of tobacco smoke, but they are  
31 also produced in the body as result of a metabolic defence response of the body to the irritant effects  
32 of smoking and of the oxidative stress processes induced by cigarette constituents (endogenous  
33 origin) (Rodgman et al., 2013; Talhout et al., 2011; Piadé et al., 2013). In particular, cigarette  
34 smoking influences the levels of nitric oxide (NO) and carbon monoxide (CO) in breath, during  
35 smoking; NO concentration decreases and CO increases, whereas after just 1 hour quit smoking, the  
36 NO and CO levels will usually return to pre-exposure values (Nadif et al. 2010; Kendrick 2013).  
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38 Also many other exhaled VOCs, as alcohols, aldehydes, ketons, sulphur compounds, hydrocarbons  
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3 and nicotine-derived products, may be present in higher concentrations in the exhaled breath of  
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5 smokers as compared to nonsmokers (Phillips, 2004).  
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8 The effects of smoking are cumulative over time and quietly a health status threshold can be  
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10 overcome. The mechanisms of disease induction by tobacco smoke have well studied (U.S. Dept. of  
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12 Health and Human Services, 2014). Tobacco smoke produces fine particulate matter that deposit on  
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14 alveoli. Since nicotine creates addiction, it leads to prolonged exposure to tobacco smoke. In  
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16 addition, smokers may increase the depth of inhalation and hold the smoke in their lungs longer to  
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18 increase nicotine uptake. When smokers inhale smoke, each cigarette puff delivers a mixture of  
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20 carcinogens and toxicants. These exogenous inhaled VOCs pass the alveolar epithelial barrier and  
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22 enter the bloodstream by reaching cells/tissues of any organ in the body; endogenous VOCs  
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24 produced by cellular metabolism return to the alveolar volumes via the blood vessels and are  
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26 exhaled (Hakim, 2012).  
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31 As result of all these biochemical pathways, VOCs in alveolar air need time to decay to  
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33 background or smoking pre-exposure levels (ranging from a few minutes to hours), so that the VOC  
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35 pattern in exhaled breath of smokers may result altered compared to nonsmokers. A too short time  
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37 elapsed from the last smoked cigarette hinders the recover to normal condition. The inhaled VOCs  
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39 are retained in the alveolar volumes and trigger the above mentioned cumulative effects increasing  
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41 the risk of a lot of smoking-related diseases (cancer, inflammatory processes, pulmonary  
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43 emphysema, etc.). The health risk of tobacco smoking increases as the daily cigarette consumption  
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45 increases (U.S. Dept. of Health and Human Services, 2014; Alonso, 2010).  
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49 In the last years, the analysis of exhaled VOCs is playing an increasingly prominent role in  
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51 medical field, particularly in disease diagnostic. The complex pattern of hundreds of VOCs in the  
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53 exhaled breath is a “snapshot” of the various biochemical pathways that the volatile compounds  
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55 follow inside the body; it’s frequently referred as “breathprint” since it’s indicative of the health of  
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57 a subject. The great expectation is that breath analysis may become a powerful non-invasive tool in  
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3 clinical practise for disease diagnostics and metabolic status monitoring (Pereira et al., 2014;  
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5 Lourenço et al. 2014).

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8 Different analytical platforms can be used for analyse the breath VOC pattern. Spectrometric  
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10 technologies (GC-MS as gold standard, SIFT-MS, DMS, PTR-MS/PTR-ToFMS, FAIMS) are  
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12 sophisticated instrumental tools offering a lot of advantages in terms of selectivity and sensitivity,  
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14 but they are inexpensive, time consuming and cannot be easily miniaturized (Mathew et al. 2015;  
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16 Rattray et al. 2014, Garcia et al. 2014). Instead e-noses are portable analytical platforms, that  
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18 provides a potential, relatively cheap and easy devices for breath analysis (Garcia et al. 2014; Bikov  
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20 et al. 2015).

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24 Most of the publications on breath analysis deals with the detection of endogenous volatile  
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26 biomarkers that could be linked to specific diseases as consequence of the arising of relevant  
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28 pathogenic processes. On the contrary, a smaller number of works considers breath analysis as  
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30 challenging method to detect inhalational exposure to toxic or noxious vapours (Jarěno-Esteban et  
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32 al. 2013; Cheng et al. 2009; Witt et al. 2011; Capone et al. 2011). It's interesting to study how  
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34 smoking influences the exhaled VOC pattern in a healthy population and testing whether breath  
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36 analysis can be used to distinguish between smokers and nonsmokers.

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40 In this work, we used breath analysis for the assessment of active smoking habit and recent use of  
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42 tobacco. From a clinical point of view, prediction of smoking habit and smoking abstinence is  
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44 indispensable because some patients do not admit to being regular smokers and claim to smoke a  
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46 smaller number of cigarettes than they actually smoked. Different exhaled VOCs have been  
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48 proposed as biomarkers of smoking status (carbon monoxide, 2,5-dimethylfuran, benzene, toluene,  
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50 xylene, 1,3-butadiene, acetonitrile) (Kendrick et al., 2013; Sandberg et al., 2011; Al-Sheyab et al.,  
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52 2015; Alonso et al., 2010; Buszewski et al., 2009; Gordon et al., 2002; Lirk et al., 2004; Crespo et  
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54 al., 2012; Jarěno-Esteban et al., 2013), but the correlation of a single compound to smoking habit is  
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56 complex and dependent on many other factors (e.g. other exogenous sources different from  
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3 cigarettes, comorbidity, dependence of compound concentration from time since smoking, large  
4 inter- and intra- individuals variability). In general, identifying which exhaled VOCs are biomarkers  
5 of a specific pathology is more complex than formerly believed, because the relation between a  
6 biomarker and a specific disease is multi-fold. Indeed, an exhaled VOC can be biomarker of several  
7 diseases, and one particular disease can be characterized by different VOC biomarkers. A specific  
8 VOC pattern, instead of individual VOCs, is probably the biomarker that most realistically  
9 represents a specific morbidity (Pereira et al.,2014; Lourenço et al., 2014).  
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19 In particular, in relation with possible smoking biomarkers, exhaled carbon monoxide is a  
20 simple marker that can be measured by commercial end-user devices for the assessment of smoking  
21 cessation, but the correlation the correlation between decreasing CO levels in exhaled breath and  
22 reducing tobacco consumption is not so good. What limits the usefulness of CO as a biomarker of  
23 smoking habit is that it can also come from sources different from smoking. In addition, the actual  
24 level of CO (8 ppm or less) indicating that a patient continues to smoke is controversial. To date, no  
25 reliable cut point has been established to allow differentiation between smokers and nonsmokers  
26 (Kendrick et al., 2013; Sandberg et al., 2011; Al-Sheyab et al., 2015).  
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37 According to what reported in literature, toluene and xylenes may be already present in indoor air,  
38 and they have been found in the breath of light smokers and nonsmokers; they show a heavy time-  
39 dependence after smoking and a large variability from subject to subject, so that they could be  
40 smoking markers only for heavy smokers and for recent use of tobacco (Alonso et al., 2010).  
41 Benzene, which is carcinogenic, is more significant since it's frequently identified in higher  
42 concentrations in the breath samples of smokers compared to non-smokers. For this reason, it has  
43 been proposed as potential marker of smoking, but it also may come from other sources of air  
44 pollution (traffic, combustion process). Moreover, during smoking benzene rapidly increases in  
45 exhaled breath of smokers and after smoking it declines to values similar found in nonsmokers  
46 (within about 1 hour); it may be used only for detect recent use of tobacco (Buszewski et al., 2009;  
47 Alonso, 2010).  
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3 Cigarette smoke causes also an exposure to 1,3-butadiene, that is in B2 group like a probable human  
4 carcinogen with a unit risk factor 30 times higher than benzene. 1,3-butadiene shows short  
5 residence times (less than 30 min after smoking) in the body, so that also this compound cannot be  
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10 useful as effective marker for smoking habit (Hecht, 2011; Gordon et al., 2002).

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12 Increased levels of acetonitrile, a constituent of cigarette smoke, were found in the breath of  
13 smokers, but the studies that validate its use as marker of active smoking behaviour are still few  
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16 (Phillips, 2004; Buszewski et al., 2009; Lirk et al., 2004; Crespo et al., 2012).

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18 Other authors studied the presence of aldehydes and carboxylic acids in the exhaled breath of  
19 healthy population (nonsmokers, ex-smokers and smokers): differently to previously mentioned  
20 VOCs, the metabolic origin of these compounds are known to derive from oxidative stress  
21 processes of cells. They found that nonanal in exhaled breath is associated to the status of being a  
22 smoker or ex-smoker, independently of the age, gender and amount of tobacco smoked (Jarěno-  
23 Esteban et al., 2013).

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25 This work was intended to contribute to the discussion: 1) which is the medical significance of  
26 VOCs in exhaled air?, 2) what are the differences between the exhaled breath of smokers and  
27 nonsmokers?, 3) which VOCs can be smoking biomarkers?

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29 Here, solid-phase microextraction (SPME) and gas chromatography coupled with mass  
30 spectrometry (GC/MS) was used for the analysis of volatiles in breath. We identified and quantified  
31 the VOCs present in the alveolar breath samples of groups of nonsmokers and subgroups of  
32 smokers sampled at different periods of withdrawal from smoking (after 1 hour abstinence and 1  
33 night out of smoking), and we used the patterns of VOCs concentrations to perform statistical  
34 analysis (Mann-Whitney test, predictive Probit model) in order to asses smoking habit.

## 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 **2. Experimental**

### 56 57 58 59 60 **2.1 Subjects**

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3 Breath analyses were conducted in 26 healthy adult volunteers (10 nonsmokers and 16  
4 smokers) recruited within the university campus in Lecce, Italy. All smokers aren't heavy smokers  
5 (<10 cigarettes/day). The smokers individuals were asked to fill three bags in three different times:  
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7 a) in the morning, fast, 1 night out of smoking and before smoking the first cigarette of the day (1°  
8 bag) ("blank" smokers), b) after 1 hour abstinence of smoking the 1<sup>st</sup> cigarette of the day (2° bag),  
9  
10 c) after 1 hour of abstinence of smoking the n<sup>th</sup>- cigarette of the day (3° bag). The collected bags  
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12 from subgroups of smokers were labelled as  $F_{i\_j\text{bag}}$  ( $i=1,\dots,16, j=1,2,3$ ). Nonsmokers individuals  
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14 were asked to fill one bag in the morning, fast; the codes for nonsmokers bags were  $N_{Fi}$ ,  $i=1,\dots,10$ .  
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16 We collected total 58 bags from smokers and nonsmokers individuals.  
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23 The study was designed to assess whether the group of all smokers ( $F_{i\_j\text{bag}}$ ,  $i=1,\dots,16$ ,  
24  $j=1,2,3$ ), can be distinguished from the group of nonsmokers ( $N_{Fi}$ ,  $i=1,\dots,10$ ). The sub-group of  
25 "blank" smokers ( $F_{i\_1\text{bag}}$ ,  $i=1,\dots,16$ ) was also compared with the nonsmokers group ( $N_{Fi}$ ,  
26  $i=1,\dots,10$ ), in order to evaluate if the smoking habit in regular smokers can be detected in "blank"  
27 conditions, i.e. after the longest time of abstinence of smoking in a day, i.e. 1 night out of smoking.  
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29 Finally, the sub-group of smokers after the 1<sup>st</sup> and the n<sup>th</sup>- smoked cigarette of the day ( $F_{i\_2\text{bag}}$  +  
30  $F_{i\_3\text{bag}}$ ,  $i=1,\dots,16$ , 1 h smoking abstinence) was compared with the subgroup of "blank" smokers (  
31  $F_{i\_1\text{bag}}$ ,  $i=1,\dots,16$ , 1 night out of smoking) in order to assess the recent use of tobacco for a regular  
32 smoker. A scheme of the study design and groups definition was shown in figure.  
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## 49 2.2 Breath sampling

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51 We sampled breath of the volunteers by a commercial breath sampling system (QUINTRON,  
52 USA). The system consists in a special 3-way valve and two bags, one (discard bag, 250 ml)  
53 discards the expired air contained in the first respiratory airways (dead-volume) and the other  
54 (collecting bag, 750 ml) collects the only alveolar air (end tidal). When the discard bag is full, a  
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3 membrane-valve opens automatically and the collecting bag fills with the alveolar air. A 1-way  
4 stopcock (into which a SPME septum was fitted) was inserted into the Luer port on the collecting  
5 bag. The volunteer was asked for doing a single deep exhalation through the mouthpiece of the  
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7 breath sampling system.  
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### 11 12 13 14 15 16 **2.3 Analytical procedure**

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18 The GC-MS analysis was performed on Agilent 5973 mass spectrometer (MSD) coupled with  
19 6890N series gas chromatograph (Agilent Technologies) with a split-splitless injector. The injector  
20 temperature was 250 °C. A DB-WAX capillary column (60 m, 0.25 mm I.D., 0.25 µm thickness)  
21 was used. The oven temperature program was as follows: initial 40 °C held for 5 min, ramped at 3  
22 °C/min to 140 °C held for 10 min, next ramped at 10 °C/min to 230 °C and held for 3 min. Electron  
23 impact ionization was applied at 70 eV. The MS analyses were carried out in full-scan mode with a  
24 scan range 30-500 amu at 3.2 scans/s. The identification of the volatile compounds was achieved by  
25 comparing mass spectra with those of the data system library (NIST 98, P > 80%). A manual SPME  
26 holder with a SPME fiber assembly Carboxen/Polydimethylsiloxane (CAR/PDMS) from Supelco  
27 (black fiber, Supelco, Bellefonte, USA) was used. Internal standard (I.S. bromobenzene, 23 ppbv)  
28 was added by a suitable multiple-steps dilution into the collecting bag after breath sample collection  
29 in order to apply semiquantitative analysis method to evaluate the concentrations of the identified  
30 volatile compounds. Since the breath exhaled at the body temperature is highly humid, it inevitably  
31 condenses on the colder inner walls of the collecting bag; it was necessary to slightly heat the bag in  
32 order to return the condensate breath to the vapour phase. This was realized by firmly placing the  
33 collecting bag onto a by a heating plate at 38°C, during the exposure of the SPME fiber for 30 min.  
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### 58 **2.4 Statistical analysis**

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3 Statistical analysis was performed using STATISTICA software. For descriptive statistics, two-  
4 sided testing and median with interquartile (IQR) range were used. Non parametric Mann-Whitney  
5 test was preferred in alternative to *t*-test for independent samples. By Mann-Whitney test, statistical  
6 significance of differences between the two compared groups (Figure 1) was assessed in terms of  
7 most significant variables (VOCs), and a  $p < 0.05$  was considered significant. Further a Probit model  
8 was used as non-linear regression model for binary responses; the method is more suitable to breath  
9 metabolomics data that typically show nonlinear patterns. This model describes the relationship  
10 between one or more continuous independent variable(s) to dichotomous (binary) variables, here  
11 corresponding to breath VOCs and binary categorical variables (i.e. the two compared groups)  
12 respectively. In particular, we retained all and only the statistical relevant variables highlighted by  
13 Mann-Whitney test, and we evaluated how this VOC pattern can predict a correct class  
14 membership.  
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### 33 3. Results and discussion

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36 Eighty-three different compounds were identified in the collected breath samples. Their  
37 concentration was calculated by normalizing peaks areas to internal standard area (I.S.) and  
38 referring to its concentration (Table 1). They belong to different classes of compounds: alcohols,  
39 aldehydes, ketons and hydrocarbons. The breath patterns of the group of the nonsmokers and the  
40 subgroup of “blank” smokers sampled in the morning, fast, 1 night out of smoking and before  
41 smoking the 1<sup>st</sup> cigarette of the day, were found less rich in VOCs than breath pattern of the  
42 subgroup of smokers after 1 h from smoking the 1<sup>st</sup> and the n<sup>th</sup> cigarette of the day. At a first  
43 qualitative analysis of data distribution in the sample population, we observed an increased  
44 occurrence of heterocyclic aromatic organic compound and alkanes hydrocarbons in the group of  
45 smokers. Given that the main objective of this study was to evaluate which VOCs can allow a real  
46 determination of smoking habit and/or smoking event, we performed Mann-Whitney test on data set  
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3 by groups. By applying Mann-Whitney test the most informative features regarding the statistical  
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5 differences between the two groups under analysis were filtered.  
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8 First, we consider the whole group of smokers regardless of whether they had smoked recently  
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10 or not (*analysis 1*). In *analysis 1*, the complete group of the smokers sampled three times in a day  
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12  $Fi\_jbag$  ( $i=1,\dots,16$ ;  $j=1$  in the morning, fast, 1 night out of smoking and before smoking the 1<sup>st</sup>  
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14 cigarette of the day;  $j=2$  smokers after 1 h from the 1<sup>st</sup> smoked cigarette of the day;  $j=3$  smokers  
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16 after 1 h from the  $n^{\text{th}}$  smoked cigarette of the day) was compared to the group of nonsmokers  $NFi$   
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18 ( $i=1,\dots,10$ ). By Mann-Whitney test, the VOCs found to be variables with statistical relevance  
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20 ( $p<0.05$ ) in analysis 1 were: a) toluene, b) pyridine, c) pyrrole. Descriptive statistic results (median,  
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22 25<sup>th</sup>-75<sup>th</sup> percentiles, minimum/maximum) for these compounds were respectively shown in Figure  
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24 2a, 2b, 2c for nonsmokers and smokers in order to easily compare the differences between the two  
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26 groups. A trend in higher occurrence and higher concentrations in the complete group of smokers  
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28 were registered for these VOCs in our sample population. The presence of toluene, pyridine and  
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30 pyrrole in human breath was confirmed by literature (Alonso et al., 2010; Schmidt et al., 2015;  
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32 Pennazza et al., 2010; Ji et al., 2002; Mansoor et al., 2014; Kapishon et al., 2013; Bazemore et al.,  
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34 2006; Marco et al., 2015; Filipiak et al., 2014; Hanai et al. 2012; Talty et al., 2013). Moreover, in  
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36 next analysis (*analysis 2*), the sub-group of the “blank” smokers  $Fi\_1bag$  (sampled in the morning,  
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38 fast, 1 night out of smoking and before smoking the 1st cigarette of the day) was compared with the  
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40 group of nonsmokers  $NFi$ ; by this comparison the Mann-Whitney test didn't show any VOCs really  
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42 statistically significant. We can mention compound nonane, 2,3-dimethyl ( $p=0.093$ ), an alkane  
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44 found more in smokers than in nonsmokers (Figure 2d).  
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51 Toluene is an aromatic hydrocarbon widely used as an industrial feedstock and as a solvent, and it's  
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53 one of the many compounds found in cigarettes (Hecht, 2011; Rodgman et al., 2013; Talhout et al.,  
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55 2011). It's thus commonly identified in breath samples independently of the source (Schmidt et al.,  
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57 2015; Pennazza et al., 2010). Toluene is classified as an intoxicative inhalants, but although less  
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3 dangerous than benzene, under the Guidelines for Carcinogen Risk Assessments (US. EPA, 2005),  
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5 the EPA considers that there is an inadequate information to assess the carcinogenic potential of  
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7 toluene. We identified toluene in greater amount in the smokers breath (Figure 2a).  
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11 Pyridine is a basic heterocyclic organic compound used as a precursor to agrochemicals and  
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13 pharmaceuticals; it's also an important solvent and reagent as well as a cigarette constituent (Hecht,  
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15 2011; Rodgman et al., 2013). It's known it causes damage to cells (Ji et al., 2002; Mansoor et al.,  
16  
17 2014). Pyridine was also proposed as a VOC marker for exposure to tobacco smoke (Kapishon et  
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19 al., 2013); other work identified pyridines and pyrazines as compounds responsible for the typical  
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21 malodour of smokers breath, being likely generated during cigar pyrolysis by cleavage of nicotine  
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23 or by Maillard reaction (Bazemore et al., 2006).  
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27 Pyrrole is another heterocyclic aromatic organic compound typically listed as a chemical constituent  
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29 in cigarettes and cigarette smoke (Hecht, 2011; Rodgman et al., 2013; Talhout et al., 2011; Piadé et  
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31 al., 2013; Marco et al., 2015). It has been detected both in the exhaled breath of lung cancer patients  
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33 and in the headspace of cancer cells *in vitro* (Filipiak et al., 2014; Hanai et al., 2012). Abnormal  
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35 concentrations of pyrrole are also related to a less known *Pyroluria*, or *pyrrole disorder*, a  
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37 metabolic condition with many physical and psychological symptoms, suffered by 10% of the  
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39 population (Talty et al., 2013), it's caused by stress that depletes the body of certain vitamins  
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41 (mainly vitamin B6) and minerals (mainly magnesium and zinc) before they are able to be  
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43 absorbed.  
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47 By considering the VOC pattern with statistical relevance ( $p < 0.05$ ) for analysis 1 (toluene,  
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49 pyridine, pyrrole), we performed a Probit Regression by comparing the nonsmokers with the  
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51 complete group of smokers. The predictive results with a good significance level  $p$  ( $p = 0.0043$ ) are  
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53 shown in Table 2. The classification test has an high sensitivity (100%) but a low specificity (20%)  
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55 due to the high number of false positives. The sensitivity measures the percentage of positives that  
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57 are correctly identified as such (i.e. here the smokers who are correctly identified as such), whereas  
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3 the specificity measures the percentage of negatives nonsmokers that are correctly identified as such  
4 (i.e. here the nonsmokers who are correctly identified as such). The test is even null specificity if we  
5 consider the “blank” smokers compared to the nonsmokers, whereas the sensitivity is optimum  
6 (100%) (analysis 2, table 3). In both data analysis, the high number of false positives may be  
7 influenced by the small sample size of this pilot study, and of course a more extended experiment  
8 on larger population could better show the small relations between nonsmokers and “blank”  
9 smokers. However, we can argue that in our sample population, all the smokers were light smokers;  
10 the breath patterns of nonsmokers and subset group of “blank” smokers, are really quite similar,  
11 hence the classification test, that forces them to belong to two different classes in analysis 1, gives  
12 unavoidably a misclassification for nonsmokers. Prediction model based on statistical relevant  
13 VOCs-patterns showed that assessment of smoking status is heavily time dependent. Since the level  
14 of exhaled VOCs decays with time since smoking, the discrimination between nonsmokers and  
15 smokers depend on the time lag between the smoking event and the breath sample collection. A  
16 breath sampling after the longest period since smoking for a regular smoker in a day (i.e. as after 1  
17 night out of smoking) ensures the recovery to pre-exposure or “non-smoker” condition; the “blank”  
18 smokers of our sample population are thus realistically more like non-smokers. Our highly sensitive  
19 test can thus be deemed effective only at ruling out a smoking habit when negative.  
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41 Later, we focused on how breath pattern is altered by cigarette smoking, aiming in finding  
42 VOCs markers of smoking event. At this aim we analysed data distribution within the group of  
43 smokers, by comparing the sub-group of the smokers after 1 h from the 1st and the nth- smoked  
44 cigarette of the day ( $Fi_{2bag} + Fi_{3bag}$ ) with the sub-group of “blank” smokers i.e. sampled in the  
45 morning, fast, 1 night out of smoking and before smoking the 1st cigarette of the day ( $Fi_{1bag}$ )  
46 (*analysis 3*). First, Mann-Whitney test filtered the most informative features regarding the statistical  
47 differences between the two groups of smokers after 1 h from last smoked cigarette and “blank”  
48 smokers. More statistical relevant variables ( $p < 0.05$ ) compared to *analyses 1* and *2* were found: a)  
49 toluene, b) pyridine, c) pyrrole, d) benzene, e) 2-butanone, f) 2-pentanone, g) 1-methyldecylamine.  
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3 Descriptive statistic results (median, 25<sup>th</sup>-75<sup>th</sup> percentiles, minimum/maximum) for these  
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5 compounds were respectively shown in Figure 4a,b,c,d,e,f,g. It can be easily observed that the  
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7 smoking event altered the breath pattern by increasing the levels of the identified VOCs with  
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9 statistical relevance. In addition to toluene, pyridine and pyrrole, another aromatic hydrocarbon, i.e.  
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11 benzene known to be a cigarette smoking product and to have carcinogenic effects on health, was  
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13 found in higher concentrations in the exhaled breath of smokers after smoking. Benzene is generally  
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15 considered a marker of lung cancer (Alonso et al., 2010; Buszewski et al., 2009; Gordon et al.,  
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17 2002; Filipiak et al., 2014). More interesting is the role of 2-butanone and 2-pentanone, that are two  
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19 important ketones whose origin could be both exogenous and endogenous. Ketone bodies are  
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21 physiologically produced when the body uses fat instead of glucose for energy (Buszewski et al.,  
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23 2007). It was observed variation of ketones levels as a function of tumor growth in *in-vitro* cancer  
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25 cells studies suggesting that ketogenetic pathways may be involved in lung cancer (Hakim et al.,  
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27 2012, Kischkel et al., 2010). Several recent studies reported their increased characteristic presence  
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29 in the breath of Río et al., 2015; Fu et al., 2014).  
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35 As in the previous analysis, we didn't use the 7 identified statistical relevant VOCs as  
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37 individual breath markers, whereas we used the 7-VOCs chemical patterns to assess if the recent  
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39 tobacco consumption (1 h abstinence from smoking). At this aim Probit Regression was applied to  
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41 VOCs data pattern and a prediction of class membership to the groups of "blank" smokers and  
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43 smokers after smoking was get. In this case a good prediction ( $p < 0.01$ ) was obtained with both high  
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45 sensitivity (84%) and specificity (100%) (Table 4). The high sensitivity means that it's possible to  
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47 correctly detect smokers, who effectively smoked 1 h before sampling, as smokers after a smoking  
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49 event. The full specificity means that it's possible to correctly identify smokers, who effectively  
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51 abstained from smoking before testing. Such information are fundamental for physician to assess  
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53 the degree of compliance with a smoking cessation programme.  
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#### 4. Conclusions

In conclusion, chromatographic measurements showed that tobacco smoking undoubtedly causes a voluntary inhalation of hundreds of harmful substances into lungs as evidenced by the alteration of breath VOCs profile. Indeed, cigarette smoking directly affects the level of several VOCs in human breath, some of which are toxic and carcinogenic, increasing the risk of lung cancer and other tumour diseases. Smokers showed higher concentrations of toluene, pyridine and pyrrole, as compared to nonsmokers. As consequence of a smoking event also the level of other VOCs, as benzene, 2-butanone, 2-pentanone, 1-methyldecylamine, increases as compared to the reference breath profile of a “blank” smoker who really observed a longer time of abstinence from smoking (i.e. 1 night out of smoking). Our statistical approach to data analysis was based on filtering the statistical relevant variables (VOCs), that highlights the differences between two classes of samples, by non parametric Mann-Whitney test, and on using the breath patterns of statistically relevant VOCs to assess the membership to the two classes under analysis by non-linear Probit regression model.

The prediction results based on our dataset showed that the assessment of smoking status is heavily time dependent. Time is required for breath VOC to reach background or pre-exposure levels ranging from a few minutes to hours. A too short time elapsed from the last smoked cigarette leads to a perturbation of the alveolar air composition; the inhaled VOCs are retained in the alveolar volumes and trigger the smoking cumulative effects increasing the risk for smoking-related diseases. Probit prediction model based on statistical relevant VOCs-patterns showed that it's possible recognise with high specificity and sensitivity smokers after a short-term exposure to tobacco (i.e. after 1 hour of smoking abstinence), whereas smokers after a long-term exposure to tobacco (i.e. after a night out of smoking) are more like non-smokers.

However, the present authors are aware of the many pitfalls that a statistical analysis can lead (confounders, false positive or “voodoo” correlations, overfitting); they admit that, up to now, the

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3 small sample size used in this pilot study doesn't allow to establish statistically significant  
4 correlations for biomedical considerations. Larger data set are needed in order to draw the  
5 conclusion that a given VOCs pattern is a biomarker for a smoking status and/or smoking event  
6 based on a reliable pattern recognition methodology. An extended study on a larger sample  
7 population of both light and heavy smokers will be considered as well as a more detailed study of  
8 the effect of time from smoking at different sampling times from smoking.  
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### 18 **Declaration of Interests**

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20 The other authors declare that they have no conflicts of interest.  
21  
22

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30  
31  
32

### 33 **References**

34  
35  
36  
37 Alonso, M., Castellanos, M., Sanchez, J.M. (2010) Evaluation of potential breath biomarkers for  
38 active smoking: assessment of smoking habits, *Anal Bioanal Chem* 396, 2987-2995.  
39

40  
41 Al-Sheyab, N., Kheirallah, K. A., Thomson Mangnall, L. J., Gallagher, R. (2015). Agreement  
42 Between Exhaled Breath Carbon Monoxide Threshold Levels and Self-Reported Cigarette Smoking  
43 in a Sample of Male Adolescents in Jordan, *Int. J. Environ. Res. Public Health* 12, 841-854.  
44  
45

46  
47 Bazemore, R., Harrison, C., Greeberg, M. (2006). Identification of Components Responsible for  
48 the Odor of Cigar Smoker's Breath, *J. Agric. Food Chem.* 54, 497-501.  
49  
50

51  
52 Bikov, A.; Lazar, Z.; Horvath, I. (2015). Established methodological issues in electronic nose  
53 research: how far are we from using these instruments in clinical settings of breath analysis?, *J.*  
54 *Breath Res* 9, 034001  
55  
56  
57  
58  
59  
60



1  
2  
3 Buszewski, B., Keszy, M., Ligor, T., Amann, A. (2007). Human exhaled air analytics:  
4 biomarkers of diseases, *Biomed. Chromatogr.* 21, 553–566.  
5  
6

7 Buszewski, B., Ulanowska, A., Ligor, T., Denderz, N., Amann, A. (2009). Analysis of exhaled  
8 breath smokers, passive smokers and non-smokers by solid-phase microextraction gas  
9 chromatography/mass spectrometry, *Biomed. Chromatogr.* 23, 551-556.  
10  
11

12  
13 Capone, S.; Mazzotta, L.; Francioso, L.; Epifani, M.; Siciliano P. (2011). Gas microsensor array  
14 for breath analysis: An explorative study of smoking status risk, *Proc. of 4th IEEE International*  
15 *Workshop on Advances in Sensors and Interfaces (IWASI)*, 121-124.  
16  
17

18  
19 Cheng, Z. J.; Warwick, G.; Yates, D. H.; Thomas, P. S. (2009). An electronic nose in the  
20 discrimination of breath from smokers and non-smokers: a model for toxin exposure, *J. Breath Res.*  
21 3, 036003 (5pp).  
22  
23

24  
25 Crespo, E., Devasena, S., Sikkens, C., Centeno, R., Cristescu, S. M., Harren Frans, J. M. (2012).  
26 Proton-transfer reaction mass spectrometry (PTRMS) in combination with thermal desorption (TD)  
27 for sensitive off-line analysis of volatiles, *Rapid Commun. Mass Spectrom.* 26, 990–996.  
28  
29

30  
31 Fernández del Río, R., O'Hara, M.E., Holt, A., Pemberton, P., Shah, T., Whitehouse, T.,  
32 Mayhew, C.A. (2015). Volatile Biomarkers in Breath Associated With Liver Cirrhosis —  
33 Comparisons of Pre- and Post-liver Transplant Breath Samples, *EBioMedicine* 2, 1243–1250.  
34  
35

36  
37 Filipiak, W., Filipiak, A., Sponring, A., Schmid, T., Zelger, B., Ager, C., Klodzinska, E., Denz,  
38 H., Pizzini, A., Lucciarini, P. (2014). Comparative analyses of volatile organic compounds (VOCs)  
39 from patients, tumors and transformed cell lines for the validation of lung cancer-derived breath  
40 markers, *Journal of Breath Research* 8, 027111.  
41  
42

43  
44 Fu, X.-A., Li, M., Knipp, R. J, Nantz, M. H., Bousamra, M. (2014). Noninvasive detection of  
45 lung cancer using exhaled breath. *Cancer Medicine* 3, 174–181.  
46  
47

48  
49 García, R. A.; Morales, V.; Martín, S.; Vilches, E.; Toledano, A. (2014). Volatile Organic  
50 Compounds Analysis in Breath Air in Healthy Volunteers and Patients Suffering Epidermoid  
51 Laryngeal Carcinomas, *Chromatographia* 77, 501-509.  
52  
53

54  
55 García, R. A.; Morales, V.; Toledano, A. (2014). Cancer diagnosis by breath analysis - what is  
56 the future? *Bioanalysis* 5, 2331-2333.  
57  
58  
59  
60

1  
2  
3 Gordon, S. M., Wallace, L. A., Brinkman, M. C., Callahan, P. J., Kenny D. V. (2002). Volatile  
4 Organic Compounds as Breath Biomarkers for Active and Passive Smoking, *Environ Health*  
5 *Perspect* 110, 689–698.  
6  
7

8  
9 Hakim, M., Broza, Y. Y., Barash, O., Peled, N., Phillips, M., Amann, A., Haick, H. (2012).  
10 Volatile Organic Compounds of Lung Cancer and Possible Biochemical Pathways, *Chem.* 112,  
11 5949–5966.  
12  
13

14  
15 Hanai, Y., Shimono, K., Oka, H., Baba, Y., Yamazaki, K., Beauchamp, G. K. (2012). Analysis  
16 of volatile organic compounds released from human lung cancer cells and from the urine tumor-  
17 bearing mice, *Cancer Cell International* 12, 7.  
18  
19

20  
21 Hecht, S. S. (2011). Tobacco smoke carcinogens and lung cancer, in: *Chemical Carcinogenesis*,  
22 ed. Penning T M, © Springer 53-71.  
23  
24

25  
26 Jarëno-Esteban, J. J., Munoz-Lucas, M. Ángeles, Carrillo-Aranda, B., Maldonado-Sanz, J. Á.,  
27 de Granda-Orive, I., Aguilar-Ros, A., Civera-Tejuca, C., Gutiérrez-Ortega, C., Callol-Sánchez, L.  
28 M. (2013). Volatile Organic Compounds in Exhaled Breath in a Healthy Population: Effect of  
29 Tobacco Smoking, *Arch Bronconeumol.* 49, 457–461.  
30  
31

32  
33 Ji, L., Melkonian, G., Riveles, K., Talbot, P. (2002). Identification of pyridine compounds in  
34 cigarette smoke solution that inhibit growth of the chick chorioallantoic membrane, *Toxicol Sci.*  
35 *Sep*, 69, 217-25.  
36  
37

38  
39 Kapishon, V., Koyanagi, G. K., Blagojevic, V., Bohme, D. K. (2013). Atmospheric pressure  
40 chemical ionization mass spectrometry of pyridine and isoprene: potential breath exposure and  
41 disease biomarkers, *J. Breath Res.* 7, 026005.  
42  
43

44  
45 Kendrick, A. H. (2013) Exhaled carbon monoxide devices in smoking cessation: physiology,  
46 controversies and equipment, *The buyers' guide to respiratory care products*, 13, 180-189, © GASP  
47 0117 955 0101  
48  
49

50  
51 Kischkel, S.; Wolfram, M.; Sawacki, A., Straker, E. M., Trefz, P.; Amann, A.; Schubert, J. K.  
52 (2010). Breath biomarkers for lung cancer detection and assessment of smoking related effects —  
53 confounding variables, influence of normalization and statistical algorithms, *Clinica Chimica Acta*  
54 411, 1637–1644.  
55  
56  
57  
58  
59  
60

1  
2  
3 Lirk, P., Bodrogi, F., Deibl, M., Kähler, Ch., Colvin, J., Moser, B., Pinggera, G., Raifer, H.,  
4 Rieder, J., Schobersberger, W. (2004). Quantification of recent smoking behaviour using proton  
5 transfer reaction-mass spectrometry (PTR-MS), *Wien Klin Wochenschr* 116/1–2, 21–25,  
6 ©Springer-Verlag 2004.  
7  
8

9  
10 Lourenço, C.; Turner, C. (2014). Breath Analysis in Disease Diagnosis: Methodological  
11 Considerations and Applications, *Metabolites* 4, (2), 465–498.  
12  
13

14 Mansoor, S., Gupta, N., Falatoonzadeh, P., Kuppermann, B.D., Kenney M.C. (2014). 2-  
15 ethylpyridine, a cigarette smoke component, causes mitochondrial damage in human retinal  
16 pigment epithelial cells in vitro, *Indian J Ophthalmol.* 62, 16-22.  
17  
18

19  
20 Marco, E., Grimalt, J. O. (2015). A rapid method for the chromatographic analysis of volatile  
21 organic compounds in exhaled breath of tobacco cigarette and electronic cigarette smokers, *Journal*  
22 *of Chromatography A.* 1410, 51–59.  
23  
24

25  
26 Mathew, T. L.; Pownraj, P.; Abdua, S.; Pullithadathil, B.; Technologies for Clinical Diagnosis  
27 Using Expired Human Breath Analysis (2015), *Diagnostics*, 5, 27-60.  
28  
29

30  
31 Nadif, R.; Matran, R.; Maccario, J.; Bechet, M.; Le Moual, N.; Scheinmann, P.; Bousquet, J.;  
32 Kauffmann, F.; Pin, I. (2010). Passive and active smoking and exhaled nitric oxide levels according  
33 to asthma and atopy in adults, *Ann Allergy Asthma Immunol.* 104, 5, 385-93.  
34  
35

36  
37 Pennazza, G., Santonico, M., Martinelli, E., D'Amico, A., Di Natale, C. (2010). Interpretation  
38 of exhaled volatile organic compounds, chapt.8, in: *Exhaled Breath - European Respiratory*  
39 *Monograph*, 49, 115-119.  
40  
41

42  
43 Pereira, J., Porto-Figueira, P., Cavaco, C., Taunk, K., Rapole, S., Dhakne, R., Nagarajaram, H.,  
44 Camara, J. S. (2014). Breath Analysis as a Potential and Non-Invasive Frontier in Disease  
45 Diagnosis: An Overview. *Metabolites*, 5, 3-55.  
46  
47

48  
49 Phillips, M. (2004). Editorial: Smoke gets in your eyes... and in your breath, *Wien Klin*  
50 *Wochenschr* 116/1-2, ©Springer-Verlag  
51  
52

53 Piadé, J.-J., Wajrock, S., Jaccard, G., Janeke, G. (2013). Formation of mainstream cigarette  
54 smoke constituents prioritized by the World Health Organization – Yield patterns observed in  
55 market surveys, clustering and inverse correlations, *Food and Chemical Toxicology* 55, 329–347.  
56  
57  
58  
59  
60

1  
2  
3 Rattray, N. J. W.; Hamrang, Z.; Trivedi, D. K.; Goodacre, R.; Fowler, S. J. , Taking your breath  
4 away: metabolomics breathes life in to personalized medicine (2014), Trends in Biotechnology, 32,  
5 10, 538-548.  
6  
7

8  
9 Rodgman, A., Perfetti, T. A. (2013). The Chemical components of tobacco and tobacco smoke,  
10 2nd edition, CRC Press © by Taylor & Francis Group, International Standard Book Number-  
11 13:978-1-4665-1552-9.  
12  
13

14 Sandberg, A., Sköld, C. M., Grunewald, J., Eklund, A., Wheelock, Å. M. (2011). Assessing  
15 Recent Smoking Status by Measuring Exhaled Carbon Monoxide Levels, PLoS ONE 6, e28864.  
16  
17

18 Schmidt, K., Podmore, I. (2015). Current Challenges in Volatile Organic Compounds Analysis  
19 as Potential Biomarkers of Cancer, Journal of Biomarkers. 981458, 16 .  
20  
21

22 Talhout, R., Schulz, T., Florek, E., van Benthem, J., Wester, P., Opperhuizen, A. (2011).  
23 Hazardous Compounds in Tobacco Smoke, Int. J. Environ. Res. Public Health 8, 613-628.  
24  
25

26 Talty, C., Dahlitz, M. (2013). Pyrrole Disorder for Therapists, The Neuropsychotherapist 3, 58-  
27 66.  
28  
29

30  
31 U.S. Department of Health and Human Services. The Health Consequences of Smoking—50  
32 Years of Progress: A Report of the Surgeon General. Atlanta: U.S. Department of Health and  
33 Human Services, Centers for Disease Control and Prevention, National Center for Chronic Disease  
34 Prevention and Health Promotion, Office on Smoking and Health, 2014.  
35 <https://www.surgeongeneral.gov/library/reports/50-years-of-progress/full-report.pdf>  
36  
37  
38  
39

40  
41 U.S. Environmental Protection Agency. Integrated Risk Information System (IRIS) on Toluene.  
42 (2005). National Center for Environmental Assessment, Office of Research and Development,  
43 Washington, DC.  
44  
45

46  
47 Witt, K.; Reulecke, S.; Voss, A. (2011). Discrimination and characterization of breath from  
48 smokers and non-smokers via electronic nose and GC/MS analysis, Conf Proc IEEE Eng Med Biol  
49 Soc. 2011, 3664-3667.  
50  
51  
52  
53  
54  
55  
56  
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58  
59  
60

**Table 1** Compounds identified in the exhaled breath of the sample population. Concentration ranges (min-max in ppbv) were reported for the groups of nonsmoker (NF), “blank” smokers (Fi\_1bag) and smokers after 1 h from smoking last cigarette (Fi\_2bag + Fi\_3bag).

RT (min)	Compound	Min-Max (ppbv)			RT (min)	Compound list	Min-Max (ppbv)		
		Non Smokers	“Blank” Smokers (1 night out of smoking)	Smokers (after 1h from smoking last cigarette)			Non Smokers	“Blank” Smokers (1 night out of smoking)	Smokers (after 1h from smoking last cigarette)
7,038	2-Methyl-1,3-Butadiene (isoprene)	0 ÷ 99,9	0 ÷ 403	0 ÷ 810	31,617	Pyridine, 3-methyl-	/	/	0 ÷ 5,9
7,052	1-Methyldecylamine	/	/	0 ÷ 591	33,471	Tetradecane, 5-methyl-	/	0 ÷ 2,5	/
8,005	1,3-Pentadiene	/	/	0 ÷ 359	34,640	2-Cyclopenten-1-one	/	/	0 ÷ 3,8
9,369	Acetone	0 ÷ 33,4	0 ÷ 69	0 ÷ 96	35,295	2-Ciclopenten-2-Methyl-1-One	/	/	0 ÷ 5,3
10,570	2-Methyl Furan	/	/	0 ÷ 9,8	35,449	Acetic acid 2-Ethylexyester	/	/	0 ÷ 5,1
11,654	2-Ethyl Butanale	/	/	0 ÷ 6,4	35,975	10-Methylnonadecane	/	0 ÷ 2,6	0 ÷ 5,9
11,720	2-Butanone	/	/	0 ÷ 51,4	35,981	Heptadecane, 8-methyl-	/	/	0 ÷ 4,5
11,780	Butanale	/	/	0 ÷ 21,9	36,090	Nonanal	/	0 ÷ 1,6	/
13,062	Ethanol	/	/	0 ÷ 74,5	36,800	Cyclohexane, (1,2-dimethylbutyl)-	/	0 ÷ 6,6	/
13,611	Benzene	/	/	0 ÷ 34,7	36,812	Cyclohexane, 1,2-diethyl-, cis-	0 ÷ 2,6	0 ÷ 3,5	0 ÷ 1,8
14,656	2-Pentanone	/	/	0 ÷ 30,9	36,820	Cyclohexane, 1-ethyl-1, 3-dimethyl-, trans-	0 ÷ 4,1	0 ÷ 2,0	0 ÷ 3,2
15,045	Pentanale	/	/	0 ÷ 66,6	37,288	2-Heptene, 4-methyl-, (E)-	0 ÷ 0,9	0 ÷ 2,8	0 ÷ 0,8
15,522	Cyclopentanol	/	/	0 ÷ 8,9	38,327	Benzene 1,2,3triethyl-5methyl	0 ÷ 5,1	0 ÷ 10,8	0 ÷ 1,1
15,629	Decane	/	0 ÷ 10,8	0 ÷ 4,0	38,583	Hydrazine, 1,2-dimethyl-	/	/	0 ÷ 9,5
17,687	Toluene	0 ÷ 7,5	0 ÷ 12,9	0 ÷ 22,8	39,313	Benzenamine, 4-(trifluotomethyl)-	/	0 ÷ 1,5	/
19,632	Hexanal	/	/	0 ÷ 24,6	39,545	Nonane, 3-methyl-	/	0 ÷ 2,9	/
20,169	1,3-butanediamine	/	0 ÷ 1,5	/	39,553	Furfural	/	/	0 ÷ 2,4
22,169	1,2-Dimethyl-Benzene	/	/	0 ÷ 4,3	40,006	Decane, 5,6-dipropyl-	0 ÷ 1,5	0 ÷ 5,9	0 ÷ 7,7
22,205	Ethanone, 1-cyclopropyl-	/	/	0 ÷ 10,6	40,013	1,3-cyclopentamedione, 2-methyl-	0 ÷ 5,9	/	0 ÷ 9,1
23,031	1-Butanol	/	0 ÷ 18,3	0 ÷ 37,1	40,016	Cyclohexanone,5methyl-2-(1methylethyl)	/	0 ÷ 12,6	/
23,043	1H-Pyrrole, 1-methyl-	/	/	0 ÷ 7,4	40,618	1-hexanol	/	0 ÷ 8,6	/
23,710	3-Heptanone	/	/	0 ÷ 8,5	40,626	1-Hexanol, 2-ethyl-	0 ÷ 12,6	0 ÷ 26,6	0 ÷ 17,9
25,563	Pyridine	/	/	0 ÷ 19,9	40,846	Decane5,6dipropyl	/	0 ÷ 8,7	/
25,771	Dodecane	/	0 ÷ 1,9	/	40,853	Nonane, 2,3-dimethyl	/	0 ÷ 6,1	0 ÷ 7,0
26,046	Cyclooctanone	/	/	0 ÷ 4,8	40,857	dl-2-Ethylhexyl chloroformate	/	/	0 ÷ 1,6
26,312	Heptadecane	/	0 ÷ 16,4	/	42,233	Pyrrrole	/	/	0 ÷ 17,5
26,963	Benzene, chloro-	0 ÷ 4,1	/	0 ÷ 3,8	48,942	Pyranoic acid	/	/	0 ÷ 9,0
27,474	Pyridine, 2-methyl-	/	/	0 ÷ 2,7	43,186	Propanoic acid	/	/	0 ÷ 3,4
28,150	1-Tridecene	/	0 ÷ 4,9	/	48,972	Butanoic acid	/	/	0 ÷ 12,9
28,664	1-Pentanol	/	/	0 ÷ 17,4	49,986	Butyrolactone	/	/	0 ÷ 10,5
28,723	Methyl-Ethyl-Benzene	/	0 ÷ 4,3	0 ÷ 4,5	50,052	Cyclohexane, 1-methyl-4-(1-methylethenyl)-cis-	/	0 ÷ 9,7	/
29,120	1,3,5,7-Cyclooctatetraene	/	0 ÷ 13,5	/	52,675	Benzene, 1,1'-(1-methyl-2-butynylidene)bis-	/	/	0 ÷ 12,6
29,154	Styrene	0 ÷ 8,9	0 ÷ 18,8	0 ÷ 12,7	52,707	Hydroxy Toluene Butilated	0 ÷ 102	0 ÷ 41,6	0 ÷ 25,5
29,924	Benzene, 1-methyl-2-(1-methylethyl)-	0 ÷ 14,2	0 ÷ 3,4	0 ÷ 14,5	55,945	Phenol, 2, 4-bis(1,1-dimethylethyl)	/	0 ÷ 7,6	0 ÷ 3,2
29,925	1-Methyl-4-(1-Methylethyl)	0 ÷ 21,9	0 ÷ 19,5	0 ÷ 32,2	56,049	Diethyl Phthalate	/	0 ÷ 8,6	/
29,954	Methyl-Benzene	/	0 ÷ 33,6	/	57,446	3-Hydroxy-4-methoxybenzoic acid	/	/	0 ÷ 17,9
30,237	Nonane,2,2,4,4,6,8,8-heptamethyl-	/	0 ÷ 7,6	0 ÷ 6,7	58,490	Propanoic acid, 2-methyl-, 3-hydroxy-2,4, 4-trimethylpentyl ester	/	0 ÷ 2,5	0 ÷ 7,9
30,256	Nonane	/	0 ÷ 7,2	0 ÷ 2,1	58,845	Benzoic acid, 4-hydroxy-3,5-dimethoxy-	0 ÷ 31,8	0 ÷ 11,8	0 ÷ 4,4
30,726	2-Octonal	/	/	0 ÷ 9,1	59,699	Butylated hydroxytoluene	0 ÷ 104	0 ÷ 176	0 ÷ 125

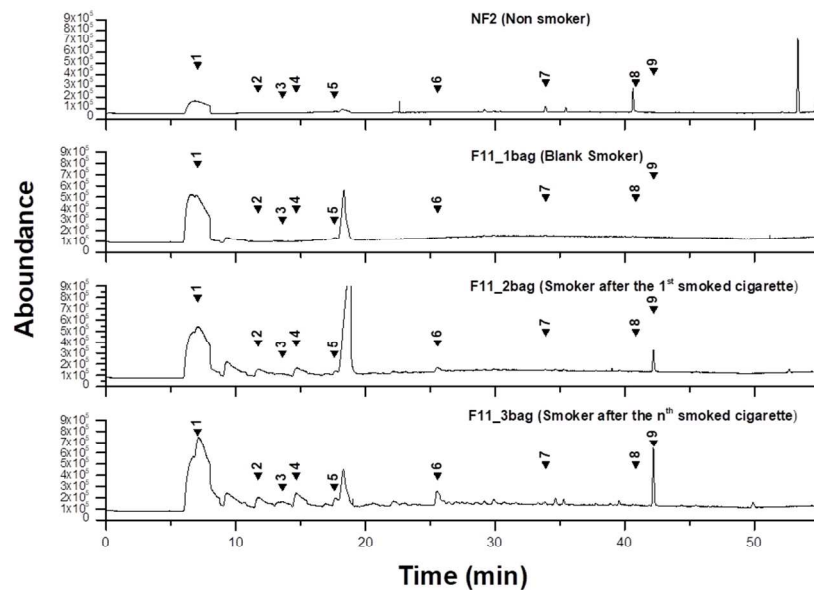
**Table 2** Confusion matrix of the predictive Probit model (analysis 1,  $p=0.0043$ ; analysis 2,  $p=0.0374$ ; analysis 3,  $p=0.0000016$ )

	<b>Predicted condition</b>	<b>True condition</b>	
<b>Analysis 1</b> (smoker vs nonsmoker) $p=0.0043$	Predicted condition positive (smoker) Predicted condition negative (nonsmoker) Accuracy (ACC)= $(TP+TN)/\text{Total population}=86\%$	<b>True positive</b> (TP)=48 <b>False negative</b> (FN)=0 <b>Sensitivity</b> or True Positive Rate $TPR=TP/(TP+FN)=100\%$ <b>False negative rate</b> (FNR), Miss rate= $FN/(FN+TP)=0\%$	<b>False positive</b> (FP)=8 <b>True negative</b> (TN)=2 <b>False positive rate</b> (FPR) or Fall-out = $1-SPC=80\%$ <b>Specificity (SPC)</b> or True Negative Rate (TNR) $TNR=TN/(TN+FP)=20\%$
<b>Analysis 2</b> ("blank" smoker vs nonsmoker) $p=0.0374$	Predicted condition positive ("blank" smoker) Predicted condition negative (nonsmoker) Accuracy (ACC)= $(TP+TN)/\text{Total population}=61\%$	<b>True positive</b> (TP)=16 <b>False negative</b> (FN)=0 <b>Sensitivity</b> or True Positive Rate $TPR=TP/(TP+FN)=100\%$ <b>False negative rate</b> (FNR), Miss rate= $FN/(FN+TP)=0\%$	<b>False positive</b> (FP)=10 <b>True negative</b> (TN)=0 <b>False positive rate</b> (FPR) or Fall-out = $1-SPC=100\%$ <b>Specificity</b> (SPC) or True Negative Rate (TNR) $TNR=TN/(TN+FP)=0\%$
<b>Analysis 3</b> (smoker after 1 h from smoking last cigarette vs "blank" smoker) $p=0.0000016$	Predicted condition positive (smoker after 1 h from smoking last cigarette) Predicted condition negative ("blank" smoker) Accuracy (ACC)= $(TP+TN)/\text{Total population}=89\%$	<b>True positive</b> (TP)=27 <b>False negative</b> (FN)=5 <b>Sensitivity</b> or True Positive Rate $TPR=TP/(TP+FN)=84\%$ <b>False negative rate (FNR),</b> Miss rate= $FN/(FN+TP)=16\%$	<b>False positive</b> (FP)=0 <b>True negative</b> (TN)=16 <b>False positive rate (FPR) or Fall-out=</b> $1-SPC=0\%$ <b>Specificity (SPC)</b> or True Negative Rate (TNR) $TNR=TN/(TN+FP)=100\%$



Scheme of the study design and definition of the groups. Analysis 1: the complete group of the smokers sampled three times in a day  $F_{i\_j\text{bag}}$  ( $i=1, \dots, 16$ ;  $j=1$  in the morning, fast, 1 night out of smoking and before smoking the 1st cigarette of the day;  $j=2$  smokers after 1 h from the 1st smoked cigarette of the day;  $j=3$  smokers after 1 h from the  $n$ th smoked cigarette of the day) compared to the group of nonsmokers  $N_{Fi}$  ( $i=1, \dots, 10$ ). Analysis 2: sub-group of the "blank" smokers  $F_{i\_1\text{bag}}$  compared with the group of nonsmokers  $N_{Fi}$ . Analysis 3: sub-group of the smokers after 1 h from the 1st and the  $n$ th- smoked cigarette of the day ( $F_{i\_2\text{bag}} + F_{i\_3\text{bag}}$ ) compared with the sub-group of "blank" smokers i.e. sampled in the morning, fast, 1 night out of smoking and before smoking the 1st cigarette of the day ( $F_{i\_1\text{bag}}$ ).

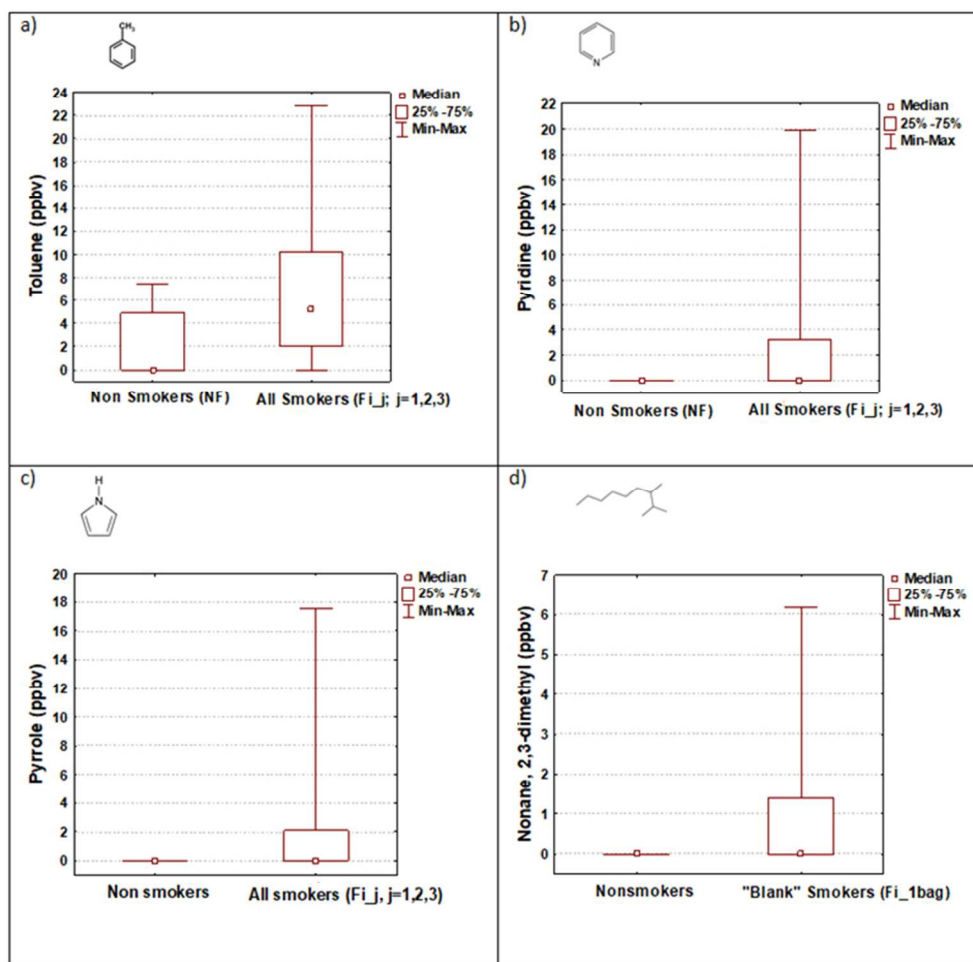
247x176mm (150 x 150 DPI)



GC chromatograms of the exhaled air samples from a nonsmoker (NF2) and a smoker subject (F11), the last sampled in three different times in a day (F11\_1bag: in the morning, fast, 1 night out of smoking and before smoking the 1st cigarette of the day; F11\_2bag: after 1 h from smoking the 1st cigarette of the day); F11\_3 bag: after 1 h from smoking the nth cigarette of the day. Significant VOCs: 1) 1-Methyldecylamine; 2) 2-Butanone; 3) Benzene; 4) 2-Pentanone; 5) Toluene; 6) Pyridine; 7) I.S.; 8) Nonane, 2,3-dimethyl; 9) Pyrrole.

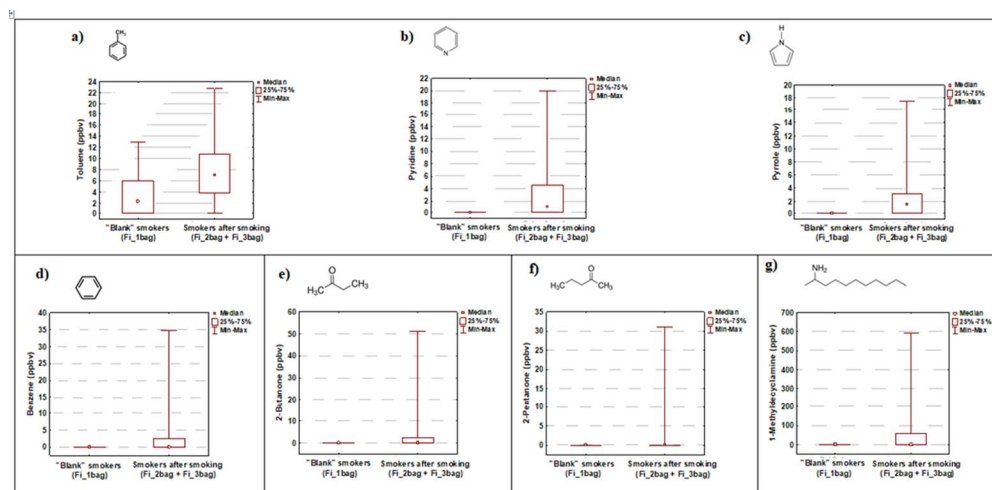
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Box plot of exhaled significant VOCs. For analysis 1: a) toluene, b) pyridine, c) pyrrole concentrations (in ppbv) (Analysis 1: the complete group of the smokers sampled three times in a day  $F_{i_j}$ bag ( $i=1, \dots, 16$ ;  $j=1$  in the morning, fast, 1 night out of smoking and before smoking the 1st cigarette of the day;  $j=2$  smokers after 1 h from the 1st smoked cigarette of the day;  $j=3$  smokers after 1 h from the  $n$ th smoked cigarette of the day) compared to the group of nonsmokers  $N_{Fi}$  ( $i=1, \dots, 10$ )). For analysis 2: d) nonane, 2,3-dimethyl- from nonsmokers and "blank" smokers (Analysis 2: sub-group of the "blank" smokers  $F_{i_1}$ bag compared with the group of nonsmokers  $N_{Fi}$ ). The bottom and top of the box are 25th and 75th percentile; the little square inside the box the median (50th percentile), and the whiskers indicate the lowest and the highest data.

138x135mm (150 x 150 DPI)



Box plot of exhaled significant VOCs. For analysis 3: a) toluene, b) pyridine, c) pyrrole, d) benzene, e) 2-butanone, f) 2-pentanone, g) 1-methyldecylamine concentrations (in ppbv) from blank smokers and smokers after cigarette smoking (Analysis 3: sub-group of the smokers after 1 h from the 1st and the nth-smoked cigarette of the day (Fi\_2bag + Fi\_3bag) compared with the sub-group of "blank" smokers i.e. sampled in the morning, fast, 1 night out of smoking and before smoking the 1st cigarette of the day (Fi\_1bag). The bottom and top of the box are 25th and 75th percentile, the little square inside the box the median (50th percentile), and the whiskers indicate the lowest and the highest data.

208x102mm (150 x 150 DPI)