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Abstract: Leaves of three different sweet basil (*Ocimum basilicum* L.) cultivars (*Italico a foglia larga*, *Cammeo*, and *Italiano classico*) packed in polyethylene bags were stored at chilling (4 °C) or non-chilling temperature (12 °C) for 9 days. During storage, visual quality, physiological (respiration rate, ethylene production, ammonium content) and biochemical (antioxidant activity, total polyphenols and polyphenol profile) parameters were measured. Detached leaves stored at chilling temperature showed visual symptoms related to chilling injury, while ethylene production and ammonium content resulted associated to cultivar sensible to low temperature. Storage at 4 °C caused a depletion in polyphenols content and antioxidant capability, which was preserved at 12 °C; while stressful storage conditions did not enhance the phenolic metabolism. However, leaves stored at 12 °C showed a good amount in metabolites until the end of the storage, suggesting the possibility to extend the storability after the expiration date in order to recover bioactive compounds



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Dear Editor,

On behalf of my co-authors and myself, I am pleased to submit the manuscript entitled "Changes in visual quality, physiological and biochemical parameters assessed during the postharvest storage at chilling or non-chilling temperatures of three sweet basil (*Ocimum basilicum* L.) cultivars" to Food Chemistry for consideration for publication.

The objective of this study was to study changes in visual quality, physiological (respiration rate, ethylene production, ammonium content) and biochemical (antioxidant activity, total polyphenols and polyphenol profile) parameters taking place during postharvest storage of three basil cultivars at chilling (4 °C) or non-chilling temperatures (12 °C). To the best of our knowledge, this is the first report regarding the effect of storage at chilling stress temperature on changes in polyphenol profile of sweet basil leaves.

We confirm that neither the manuscript nor any part of it has been published or is under consideration for publication elsewhere (abstract excluded). Any reference to or use of previously published material protected by copyright is explicitly acknowledged in the manuscript.

All authors have given a substantial contribution to conception and study design, data acquisition analysis and interpretation, handled drafting of article and revised it critically for intellectual content and participated to final approval of the final version to be published.

In my opinion, the result of the present study could be of interest for Food Chemistry's readers and I hope that the paper might be acceptable for publication.

I thank you in advance for your attention to our manuscript.

Looking forward to your reply.

Yours Sincerely,
Rosaria Cozzolino, PhD

Highlights

- Darkened spots appeared on chilling injured basil leaves
- Respiratory metabolism was not promoted by chilling stress temperature
- Storage at 12°C preserved the polyphenols profile
- Ethylene biosynthesis might be an indicator of tissue sensitivity to chilling temperature
- Depletion in phenolic compounds was measured at chilling temperature

1 **Changes in visual quality, physiological and biochemical parameters assessed during the postharvest storage at**
2 **chilling or non-chilling temperatures of three sweet basil (*Ocimum basilicum* L.) cultivars**

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31 **Abstract**

32 Leaves of three different sweet basil (*Ocimum basilicum* L.) cultivars (*Italico a foglia larga*, *Cammeo*, and *Italiano*
33 *classico*) packed in macro-perforated polyethylene bags were stored at chilling (4 °C) or non-chilling temperature (12
34 °C) for 9 days. During storage, visual quality, physiological (respiration rate, ethylene production, ammonium content)
35 and biochemical (antioxidant activity, total polyphenols and polyphenol profile) parameters were measured. Detached
36 leaves stored at chilling temperature showed visual symptoms related to chilling injury, while ethylene production and
37 ammonium content resulted associated to cultivar sensibility to damage at low temperature. Storage at 4 °C caused a
38 depletion in polyphenols content and antioxidant capability, which was preserved at 12 °C. Regarding the polyphenols
39 profile, stressful storage conditions did not enhance the phenolic metabolism. However, leaves stored at 12 °C showed a
40 good amount in metabolites also at the end of the storage, suggesting the possibility to extend the storability after the
41 expiration date, for a possible recovery of bioactive compounds.

42

43 **Keywords:** *Ocimum basilicum* L.; chilling injury; visual quality; polyphenols; ultra performance liquid chromatography

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47 **1. Introduction**

48 Sweet basil (*Ocimum basilicum* L.) is a perennial crop that is widely diffused in Asia, Africa, South America, and in the
49 Mediterranean region. Sweet basil represents a rich source of phytochemicals and, similar to other culinary herbs, it is
50 extensively used for its organoleptic properties. In medicine, the importance of sweet basil is related to its antioxidant,
51 antimicrobial, and antiviral properties (Chiang, Ng, Cheng, Chiang & Lin, 2005). Indeed, the consumption of basil leaves
52 markedly increase glutathione S-transferase activity that partly controls chemical carcinogens in the stomach, liver, and
53 oesophagus (Aruna & Sivaramakrishnan, 1990). Sweet basil contains acidic phenolic compounds, such as cinnamic,
54 caffeic, sinapic, caftaric, rosmarinic and ferulic acid, and different flavonoids, such as apigenin and catechin (Baritoux,
55 Amiot, & Nicolas, 1991). These compounds act as strong antioxidants, free radical scavengers, and metal chelators (Cook
56 & Samman, 1996). Moreover, chicoric acid, present in different parts of basil (Lee & Scagel, 2009), is thought to
57 behaviour as an antioxidant, anti-inflammatory, antiviral, and immune-stimulator (Tsai, Chiou, Chan, Sung & Lin, 2012).
58 Fresh sweet basil, as well as fresh spices and culinary herbs, are considered perishable commodities with a very short
59 shelf life (Loughrin & Kasperbauer, 2001). In sweet basil leaves, it is well documented that storage at low temperature
60 (below 12 °C) causes chilling injury, characterized by brown discoloration of the middle areas of the leaf, stem browning
61 and collapse, wilting of the leaves and loss of glossy appearance and characteristic aroma (Cozzolino et al., 2016). In
62 addition, chilling stress temperature might cause changes in the polyphenolic compounds and antioxidant activity of
63 stored leaves. Many papers showed that chilling stress at a critical low temperature enhances phenolic metabolism, which
64 varies among commodities (Lattanzio, 2003). However, based on the authors' knowledge, this is the first report regarding
65 the effect of storage at chilling temperature on changes in polyphenol profile of sweet basil leaves. Starting from these
66 findings, this paper aims to study changes in visual quality, physiological (respiration rate, ethylene production,
67 ammonium content) and biochemical (antioxidant activity, total polyphenols and polyphenol profile) parameters taking
68 place during postharvest storage of three basil cultivars at chilling (4 °C) or non-chilling temperatures (12 °C).

69

70 **2. Materials and Methods**

71 *2.1. Chemicals and reagents*

72 Caffeic, ferulic, p-coumaric, gallic, caftaric, rosmarinic, and chlorogenic acids, epicatechin, rutin, quercetin, 2,2-
73 diphenyl-1-picrylhydrazyl (DPPH), butyl-parahydroxybenzoate, high pressure liquid chromatography (HPLC)-grade
74 methanol, sulphuric, metaphosphoric, acetic and formic acids, acetonitrile, ethanol and acetone were purchased from
75 Sigma-Aldrich (Milano, Italy). Apigenin, luteolin and hyperoside were purchased from Extrasynthese (Genay, France).

76

77 *2.2. Plant material and experimental set-up*

78 Plants of three sweet basil (*Ocimum basilicum* L.) cultivars (*Italico a foglia larga*, Enza Zaden srl, Tarquinia, Italy;
79 *Cammeo*, Sais Sementi, Cesena, Italy; *Italiano classico*, Sementi Fuscello, Andria, Italy) were cultivated in greenhouses
80 and purchased from a local farm (Ortoflora, Fasano, Italy). Three consecutive experiments, one for each basil cultivar,
81 were performed. In each experiment and for each cultivar, 120 marketable basil plants were delivered to the Postharvest
82 Laboratory of the ISPA-CNR. Basil leaves of each cultivar were detached from plants and pooled. Then, approximately
83 200 g of detached basil leaves were placed in each polypropylene tray and packed in a macro-perforated polyethylene
84 bag, in order to avoid a modification of the atmosphere inside packages and to limit dehydration. In each experiment,
85 basil leaves bags were stored at chilling (4 °C) or non-chilling temperature (12 °C) for 9 days. Twelve bags for each
86 cultivar and experiment were prepared: two replicates for two temperatures (4 or 12 °C) for three storage time (3, 6 and
87 9 days). All plastic materials were purchased from Carton Pack (Rutigliano, Italy). Basil leaves of each cultivar at time
88 0 and after 3, 6 and 9 days were analysed to evaluate the sensory visual quality, physiological (respiration rate, ethylene
89 production and ammonium content), and biochemical parameters (antioxidant activity, total polyphenols and
90 polyphenol profile).

91

92 *2.3. Sensory visual quality, respiration rate, ethylene production and ammonium content*

93 A group of five trained researchers used a colour photographic scale associated with a brief description as a reference to
94 subjectively assess the sensory visual quality (VQ). Five quality levels, according to the following scale, were used: 5 =
95 excellent, fresh appearance; 4 = good; 3 = fair (limit of marketability); 2 = poor (just below the limit of marketability);
96 and 1 = very bad, inedible. A visual quality of 3 was considered the minimum threshold of acceptance for sale or
97 consumption, and values below 3 indicated a waste product (Cozzolino et al., 2016).

98 The respiration rate was measured using a closed system measured the respiration rate, as reported by Kader (2002).
99 Approximately 100 g of leaves for each replicate was placed into 6 L sealed plastic jars (one jar x replicate), where CO₂
100 was allowed to accumulate up to 0.1%. The time taken to reach this threshold was measured by monitoring the CO₂
101 concentration at regular time intervals. For CO₂ analysis, 1 mL of gas sample was taken from the head space of the
102 plastic jars through a rubber septum and injected into a gas chromatograph (P200 Micro GC, Agilent, Santa Clara, CA,
103 USA) equipped with a thermal conductivity detector. Carbon dioxide was analysed with a retention time of 16 s and a
104 total run time of 120 s on a 10-m porous polymer (PPU) column at a constant temperature of 70 °C. The respiration rate

105 of the leaves of each basil cultivar was measured on detached leaves at 0 day (after the exposure for 12 h at 4 and 12
106 °C) and after 3, 6 and 9 days of cold storage at 4 and 12 °C and results were expressed as mL CO₂/kg/h.
107 Ethylene production (μL C₂H₄/kg/h) was measured using a closed system (Kader, 1992). Fresh produce (about 50 g for
108 each replicate), was placed into 4.20 L sealed plastic-jars, where ethylene was allowed to accumulate until 0.1 ppm
109 (standard concentration). The time taken to reach this threshold was measured by monitoring the CO₂ concentration at
110 regular time intervals. Then, the gas sample was taken from the headspace through a rubber septum and measured using
111 ethylene analyser (Easy-1 Absoger, Les Barthes, France). The ethylene concentration was then referred to the sample
112 weight, the headspace volume in the jars, and the elapsed time. Ethylene from the leaves of each basil cultivar was
113 measured at 0 day (after the exposure for 12 h at 4 and 12 °C) and after 3, 6 and 9 days of cold storage at 4 and 12 °C.
114 For ammonium (NH₄⁺) analysis, five grams of chopped basil leaves was homogenised (Ultra-Turrax T-25, IKA,
115 Staufen, Germany) for 2 min in 20 mL of deionised water. The mixture was centrifuged for 5 min at 6,440×g, and 0.5
116 mL of the extract was mixed with 5 mL of nitroprusside reagent (phenol and hypochlorite in alkali reaction mixture)
117 and heated at 37 °C for 20 min. The colour development was determined using a spectrophotometer (UV-1800,
118 Shimadzu, Kyoto, Japan) reading the absorbance at 635 nm. The concentration of NH₄⁺ was expressed as μmoles
119 NH₄⁺/g, using ammonium sulphate as standard (0-10 μg/mL, R² = 0.99).

120

121 *2.4. Extraction of the polyphenol portion*

122 In each experiment, basil leaves were triturated in a mixture of methanol-acetic acid-water (8: 1: 1) in a ratio 1/5 (w/v)
123 and maintained overnight in the dark at 4 °C. The supernatants were recovered by centrifugation (11,600 x g, 15 min;
124 Beckman Italia, Cassina de' Pecchi, Milano, Italy) and stored at -26°C until time of analysis.

125

126 *2.4.1. Free radical scavenging activity*

127 The free radical scavenging activity was determined using the stable radical DPPH assay (Brand-Williams, Cuvelier, &
128 Berset, 1995). The analysis was performed in microplates by adding 7.5 μL of an extract to 303 μL of a methanol-
129 DPPH solution (153 mM). Next, the absorbance at λ= 517 nm was measured (Cary 50 MPR, Varian, Palo Alto, CA,
130 USA). The absorbance of DPPH in the absence of antioxidants (control sample) was used to determine the baseline
131 value. The EC₅₀ value defined the sample concentration (in mg) necessary to inhibit the DPPH radical activity by 50%
132 during a 60 min incubation.

133 *2.4.2. Total polyphenols content*

134 The total phenolic content was determined using Folin–Ciocalteu reagent (Singleton & Rossi, 1965). The absorbance at
135 $\lambda = 760$ nm was determined at room temperature using a Cary 73 UV/Vis spectrophotometer (Varian, Palo Alto, CA,
136 USA). Quantification was based on a standard curve generated using gallic acid at concentrations ranging from 0.5 mM
137 to 5 mM ($y = 0.5488x + 0.0148$, $R^2 = 0.9919$). The results were expressed as mg of gallic acid equivalent (GAE)/g of the
138 fresh product \pm standard deviation (SD).

139 *2.4.3. Polyphenols profile*

140 An ACQUITY Ultra Performance LCTM system (Waters, Milford, MA, USA) linked to a PDA 2996 photodiode array
141 detector (Waters) was used for ultra-high-performance liquid chromatography (UPLC) analyses. The Empower
142 software controlled the instruments and acquired and processed the data. The extracts and standards (previously
143 dissolved in methanol) were filtered (0.45- μ m; Waters) before analysis. The analyses were performed at 30 °C using a
144 reversed phase column (BEH C₁₈, 1.7 μ m, 2.1 x 100 mm; Waters) following the method of Fratianni et al. (2013). The
145 mobile phase consisted of solvent A (7.5 mM acetic acid) and solvent B (acetonitrile) at a flow rate of 250 μ L/min.
146 Gradient elution was employed, starting with 5% B for 0.8 min, followed by 5-20% B over 5.2 min; isocratic 20% B for
147 0.5 min; 20-30% B for 1 min; isocratic 30% B for 0.2 min; 30-50% B over 2.3 min; 50-100% B over 1 min; isocratic
148 100% B for 1 min; and 100-5% B over 0.5 min. At the end of this process, the system equilibrated the column under the
149 initial conditions for 2.5 min. The pressure ranged from 6000 to 8000 psi during the chromatographic run. The effluent
150 was introduced into an LC detector (scanning range: 210-400 nm, resolution: 1.2 nm). The injection volume was 5 μ L.
151 Each sample was tested in triplicate, and the results were expressed as the mean values \pm standard deviations.

152

153 *2.5. Statistical analysis*

154 For each experiment, a multifactor ANOVA for $P \leq 0.05$ was performed with the aim of evaluating the effect of
155 temperature (at chilling, 4 °C, or non-chilling, 12 °C), storage (3, 6 and 9 days) and their interaction on sensory visual
156 quality, physiological (respiration rate, ethylene production, ammonium content) and biochemical parameters (total
157 polyphenols content and antioxidant activity).

158 In addition, for each experiment, individual phenolic compounds measured by UPLC were processed performing a one-
159 way ANOVA for $P \leq 0.05$, for fresh and stored samples at chilling or non-chilling temperatures at different storage time,
160 with data means arranged in a completely randomized design. The mean values ($n=2$) for individual phenols
161 (determined by UPLC) were separated using the Student–Newman–Keuls (SNK) test ($P \leq 0.05$). Statistical analysis was
162 performed using the Statistica 6.0 software.

163

164 **3. Results and discussion**

165 *3.1 Effect of storage at chilling or non-chilling temperatures (4 or 12 °C) on basil leaves sensory visual quality and*
166 *physiological parameters*

167 Visual quality was significantly affected by temperature and storage in all cultivars (*Italico a foglia larga*, *Cammeo* and
168 *Italiano classico*), as reported in Table 1. During storage, all samples (4 °C and 12 °C) exhibited significant reductions
169 in VQ scores (Table 2); in addition, leaves stored at 12 °C exhibited VQ mean values 20%, 30% and 35% higher than
170 samples stored at 4 °C for cv *Italico a foglia larga*, *Italiano classico* and *Cammeo*, respectively (Table 2). The low VQ
171 scores assigned to basil leaves stored at 4 °C during the entire conservation was mainly due to the loss of freshness and
172 to the appearance of slightly darkened spots, which increased on the entire leaf surface at the end of the storage. These
173 symptoms, which were mainly observed in the *Italiano classico* basil leaves, were previously reported on other basil
174 cultivars and represent a clear manifestation of chilling injury damage at stress temperature (Wongsheree, Ketsa & van
175 Doorn, 2009).

176 Regarding respiration rate, all cultivars exhibited similar initial values of approximately 33.0 mL CO₂/kg/h (Figure 1 A-
177 D-G), according to the data reported by Bekhradi et al. (2015) for *Dolly*, a Genovese basil cultivar. The respiration rate
178 was exclusively affected by storage time in *Italico a foglia larga* (Table 1), showing slight changes during the storage at
179 both temperatures (Figure 1 A). Moreover, the respiration rate was affected by temperature, storage and interaction in
180 *Cammeo* and only by temperature in *Italiano Classico* (Table 1). In the last two cultivars, increased respiration rates
181 were detected in leaves stored at 12 °C compared with 4 °C. This effect was showed only at the end of storage in
182 *Cammeo* (Figure 1 D), whereas it was measured throughout the entire storage in *Italiano Classico* (Figure 1 G).
183 Cantwell & Reid (1993) reported similar results for basil leaves stored for 3 days at 0 °C or 10 °C. Thus, it seemed that
184 the storage of sweet basil leaves at chilling stress temperature (4 °C) did not cause an increase in respiratory
185 metabolism, which is consistent with previous reports on chilling-sensitive vegetables (Saltveit & Morris, 2000).

186 As for ethylene production, the three cultivars of basil exhibited a different pattern (Figure 1 B-E-H). In particular,
187 ethylene production was affected only by storage in *Italico a foglia larga*, which showed low values with slight changes
188 during storage at both temperatures (Figure 1 B), according to the respiration rate results. Whereas, ethylene was
189 affected by temperature, storage and their interaction in the other two cultivars studied (Table 1). In detail, in *Cammeo*
190 very low ethylene values were measured until 6 days of storage (both at 4 °C and 12 °C). Then, at 9th day of storage, an
191 increase in ethylene production of approximately 2-fold was detected in leaves stored at 12 °C respect to the ones stored
192 at 4 °C (Figure 1 E). In *Italiano classico*, a severe increase in ethylene content in leaves stored at 4 °C was measured
193 starting from the 3th day in storage, whereas no ethylene accumulation was noted in leaves stored at 12 °C (Figure 1 H).

194 This last result is consistent with data reported by Aharoni et al. (2010) on sweet basil leaves stored at 6 or 12 °C for 1
195 week. The increase in ethylene biosynthesis during storage has been suggested as an indicator of tissue sensitivity to
196 chilling temperatures in different vegetables (Lafuente, Zacarias, Martínez-Téllez, Sanchez-Ballesta & Dupille, 2001;
197 Megías et al., 2016). The induction of ethylene production is always associated with an accumulation of transcripts for
198 the ethylene biosynthesis enzymes (Lado, Rodrigo, Cronje & Zacarías, 2015). The different patterns in ethylene
199 production revealed by the different cultivars might lead to consider *Italico a foglia larga* as a genotype with low
200 sensitivity to chilling temperature of 4 °C, whereas *Italiano classico* could be considered a cultivar that has a high
201 sensitivity to injury at low temperature. Thus, the sensibility to chilling injury exhibited by different basil cultivars
202 could be considered as a genotypic trait, as previously highlighted by Cantwell & Reid (1993).

203 Ammonium content was affected only by interaction in *Italico a foglia larga* and by temperature, storage duration and
204 interaction in *Cammeo* and *Italiano Classico* (Table 1). As reported in Figure 1 C-F-I, during storage, starting from an
205 initial value of approximately 0.5 µg NH₄⁺/g, an increase in ammonium was measured in basil leaves of cv *Italico a*
206 *foglia larga*, *Cammeo* and *Italiano classico* stored at 4 °C. Similar NH₄⁺ accumulation was observed in vegetables that
207 were stored in stressful conditions (Cefola, Amodio, Rinaldi, Vanadia, & Colelli, 2010; Cefola, Pace, Colelli, &
208 Cantwell, 2015). On the contrary, during storage at 12 °C, the ammonium content exhibited a slight reduction in *Italico*
209 *a foglia larga* and *Cammeo* (Figure 1 C-F) or remained almost constant in *Italiano Classico* (Figure 1 D). Generally,
210 during storage low ammonium accumulation is associated with a good preservation of overall quality for leafy
211 vegetable (Tudela, Marín, Garrido, Cantwell, Medina-Martínez & Gil, 2013; Cefola & Pace 2015).

212

213 3.2 Effect of storage at chilling or non-chilling temperatures (4 or 12 °C) on basil leaves antioxidant activity and 214 total polyphenols

215 Antioxidant activity and total phenols content were affected by temperature, storage duration and their interaction in
216 *Cammeo* and *Italiano Classico* cultivars (Table 1).

217 Fresh leaves of all three cultivars exhibited an excellent antioxidant activity, with EC₅₀ values ranging between 0.78 mg
218 (*Italiano classico*) and 1.05 mg (*Italico a foglia larga*) (Figure 2 A-B-C). Such values are of remarkable interest, mainly
219 compared with previous works that reported EC₅₀ values between 1 and 4 mg (extraction in ethanol and water,
220 respectively) (Sulaiman, Sajak, Ooi, Supriatno & Seow 2011). Our results were more similar to those of Hakkim, Gowri
221 Shankar & Girija (2007), which, however, worked using previously dried leaves. Storage at chilling or non-chilling
222 temperatures (4 or 12 °C) affected the antioxidant activity in different modes (Figure 2 A-B-C). At the end of storage at
223 4 °C, cv *Italico a foglia larga* and *Italiano classico* exhibited a nearly 100% loss of antioxidant capability respect to

224 fresh samples (2.015 mg *versus* 1.05 mg and 1.34 mg *versus* 0.78 mg, respectively, Figure 2 A and 2 C); the loss was
225 even more evident in the case of cv *Cammeo*; in fact, about 7 times more of extract (5.99 mg of extract instead of the
226 initial 0.836 mg, Figure 2 B) was necessary to inhibit the activity of the stable radical DPPH (Figure 2 B). Storage at
227 12°C effectively preserved the antioxidant activity of the three cultivars for 9 days, and the loss of antioxidant power
228 was minimal. Both considering the antioxidant activity observed at the 9th day of storage and the trend exhibited
229 throughout storage, cv *Italiano classico* could be considered to exhibit superior quality compared with the other two
230 varieties and probably related to its different chilling sensibility (Wongsheree, Ketsa & van Doorn, 2009).
231 The content of total polyphenols in the fresh detached leaves was approximately 2.5 mg GAE /g in all basil cultivars
232 studied (Figure 2 D-E-F). Such data are consistent with Dogan, Turan, Dogan, Arslan, & Alkan (2005), and Shiga,
233 Shoji, Shimada, Hashida, Goto & Yoshihara (2009). At the 9th day of storage at 4°C, cv *Italico a foglia larga* and
234 *Italiano classico* exhibited respectively losses of 46% and 20% respect to fresh leaves (Figure 2 D and 2 F); at the end
235 of the storage, the loss of polyphenols was much more marked in cultivar *Cammeo* basil leaves (78%) stored at 4°C. On
236 the other hand, storage at 12°C seemed strongly limit this event in all cultivars (Figure 2 D-E-F). Anyway, data of
237 antioxidant activity and polyphenols content resulted highly associated; a reduction in total phenol content generally
238 corresponded to a loss in antioxidant capability, mainly highlighted at chilling temperature.

239

240 3.3 Effect of storage at chilling or non-chilling temperatures (4 or 12 °C) on polyphenol profile

241 The polyphenol profile obtained by UPLC analysis is reported in Table 3, which shows the results of the three one-way
242 ANOVA carried out on the three experiments (*Italico a foglia larga*, *Cammeo* and *Italiano classico*) for fresh samples (0
243 day) and basil leaves stored at chilling (4 °C) or no chilling temperature (12 °C) at different storage time (3, 6 and 9
244 days). Rosmarinic, caffeic, gallic acid and apigenin were found in all the three cultivars starting from time zero. However,
245 some differences characterising the cultivars, as well as the time and temperature of storage, complemented the scenario,
246 confirming the results obtained by Jayasinghe, Gotoh, Aoki, & Wada (2003). Rosmarinic acid was the most abundant
247 polyphenol (1.5-1.4 mg/gr of fresh product) detected in all the three cultivars. At the end of storage, chilling (4 °C)
248 caused different loss of rosmarinic acid respect to fresh samples (analysed at 0 day) depending on cultivar, ranging from
249 30% (*Italiano classico*) to 85% (*Cammeo*), with an intermediate behaviour in *Italico a foglia larga* (52%). This reduction
250 is probably due to the antioxidant role of rosmarinic acid (Jayasinghe et al., 2003) in chilling stressed basil leaves. On the
251 other hand, the non-chilling temperature allowed to preserve this important molecule, reporting a decrease of the amount
252 of rosmarinic acid oscillating from 6% (*Cammeo*) to 19% (*Italiano Classico*) until 47% (*Italico a foglia larga*). As regard
253 caffeic acid, present just after detaching from plants, in the highest quantities in *Cammeo* (508.7 ±33.0 µg/g fw) and

254 *Italiano Classico* ($547.0 \pm 36 \mu\text{g/g fw}$), the trend was different, depending on the storage temperature used. At 12°C an
255 oscillating trend was measured in all cultivars; moreover at the end of the storage *Italiano classico* and *Italico a foglia*
256 *larga* showed an amount in caffeic acid similar to fresh leaves, whereas a slight reduction was observed in *Cammeo*
257 leaves (Table 3). Despite, storage at 4°C led to a linear decrease of the amount of caffeic acid, and this was particularly
258 evident in the *Cammeo* cultivar (Table 3).

259 Just after detachment, gallic acid was measured in the highest amount in the *Italico a foglia larga* ($117 \pm 16 \mu\text{g/g fw}$),
260 whereas a content 4-fold lower were detected in the other two cultivars analysed (Table 3). In all cultivars, storage at 12°C
261 preserved the initial content, although with some fluctuations; a similar behaviour was also showed in basil leaves of
262 cv *Cammeo* and *Italiano Classico* stored at 4°C . Differently, in *Italico a foglia larga* storage at 4°C caused a severe
263 reduction of about 4-fold respect to initial value at the end of storage (Table 3).

264 Apigenin was the only flavonoid initially present in all the three cultivars; in particular it was detected in *Italiano classico*
265 in an amount of $42.0 (\pm 2.8) \mu\text{g/g fw}$, 7-fold-higher than the content measured in the other two cultivars (Table 3).
266 Moreover, at the end of storage, in *Italico a foglia larga* an increase in the content of apigenin of 28% (12°C) and 41%
267 (4°C), respect to the fresh product, was measured. The increase of apigenin turned more marked in *Cammeo* (5.4 and 4.1
268 times more after storage of 9 days at 12° and at 4°C , respectively). At the end of the storage, it is to emphasize the strong
269 increase of apigenin found in the *Italiano Classico* stored at 12°C compared to the storage at chilling temperature, which
270 maintained a level of apigenin similar to that observed in the fresh product (Table 3).

271 The three cultivars exhibited, although in a specific way, other secondary metabolites, not always present in the fresh
272 product, the synthesis of which was dependent on the temperature and the time of storage. Among the phenolic acids, at
273 time zero we found chlorogenic acid in cultivars *Italico a foglia larga* and *Italiano classico*, but not in *Cammeo*. After 9
274 days of storage in *Italico a foglia larga* chlorogenic acid increased of about 3 and 4-fold in samples stored at 4 and 12°C ,
275 respectively. Therefore, in the *Italiano classico* a similar increase was measured only at 12°C , whereas no significant
276 changes were detected in leaves stored at 4°C respect to fresh samples (Table 3). Chicoric acid was found in the cultivar
277 *Italico a foglia larga* starting from the 6th day and the 9th day of storage at chilling and no-chilling temperatures,
278 respectively. Cultivar *Cammeo* resulted to contain chicoric acid from the 3rd day of storage. However, while the *Italico a*
279 *foglia larga* was probably able to synthesize chicoric acid during storage, in cultivar *Cammeo* the metabolite disappeared
280 at the 9th day of storage at 4°C , but continued to be present in the samples stored at 12°C (Table 3). Caftaric acid, was
281 initially detected only in *Cammeo* ($16.0 \pm 1.5 \mu\text{g/g fw}$) and *Italiano Classico* ($24.5 \pm 0.8 \mu\text{g/g fw}$). During storage at 4°C ,
282 it was detected only at 3rd day in *Italiano Classico*, while it was never detected in *Cammeo*. Differently during storage at
283 12°C the amount in caftaric acid doubled in both cultivars respect to fresh samples. As regard cv *Italico a foglia larga*,

284 caftaric acid was shown only at the end of the storage at 12 °C (Table 3). Among flavonoids, quercetin was detected in
285 cultivar *Cammeo* only after 9 days at 12 °C, while in *Italico a foglia larga* starting from the 3rd day of storage at both
286 temperatures (Table 3). The loss of basil leaves membrane structure, under chilling stress temperature, involves an
287 overproduction of radical oxygen specie (ROS) including $\cdot\text{O}_2^-$, HOOH and hydroxyl radical and singlet oxygen, leading to
288 an oxidative stress. There is evidence to confirm that phenolic compounds can detoxify ROS by donating a hydrogen and
289 decomposing in peroxides. Thus, the reduction in phenolic compounds, such as rosmarinic, gallic, chlorogenic and caffeic
290 acids, in basil leaves stored at chilling stress temperature, might be considered the result of the scavenging of ROS (Dua,
291 Agrawal, Singh & Mahajan, 2016). On the other hand, at 12 °C, the polyphenols content was preserved and in many
292 cases enhanced as consequence of the biochemical mechanisms of biosynthesis, which are promoted after the detachment
293 of basil leaves from the plants as well as in wounded vegetable (Saltveit, 2000). Thus, the presence of rosmarinic, caffeic
294 and caftaric acids in important amount, even after 9 days, in basil leaves stored at 12 °C, could give an important
295 "functional" meaning, suggesting the possibility of a recovery of bioactive compounds at the end of the storage.
296 Rosmarinic acid might be metabolised by the microbiome to produce caffeic acid and derivatives, including caftaric acid
297 (Bel-Rhliid, Crespy, Pagé-Zoerkler, Nagy, Raab & Hansen, 2009). Caffeic and caftaric acid can be subsequently
298 metabolised by the microbiome, producing simpler molecules with a positive impact on both the microbiome and
299 organism (Gonthier et al., 2006).

300

301 **4. Conclusion**

302 Postharvest storage of detached basil leaves at chilling temperature (4 °C) caused the manifestation of dark spots on leaf
303 surface. Moreover, chilling temperature did not enhanced the respiratory metabolism, while it caused a significant
304 increase in ethylene production and ammonium content. Thus, these two last parameters could be indicator of basil
305 cultivar chilling sensibility. In addition, storage at 4 °C caused a depletion in antioxidant activity and polyphenols
306 content. The significant reduction in the main phenolic compounds (rosmarinic, gallic, chlorogenic and caffeic acids)
307 measured in chilling injured basil leaves might be associated to the no-enzymatic antioxidant action performed by these
308 metabolites. On the other hand, the good preservation or the enhancement in phenolic compounds measured even after 9
309 days of storage at 12 °C, led to consider basil leaves as a good source of bioactive molecules also after the expiration
310 date.

311

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314

315 **References**

- 316 Aharoni, N., Kenigsbuch, D., Chalupowicz, D., Faura-Mlinski, M., Aharon, Z., Maurer, D., Ovadia, A., & Lers, A.
317 (2010). Reducing chilling injury and decay in stored sweet basil. *Israel Journal of Plant Sciences*, 58, 167-181.
318
- 319 Aruna, K., & Sivaramakrishnan, V.M. (1990). Plant products as protective agents against cancer. *Indian Journal of*
320 *Experimental Biology*, 28, 1008-1011.
321
- 322 Baritoux, O., Amiot, M.J., Nicolas, J. (1991). Enzymatic browning of basil (*Ocimum basilicum* L.) studies on
323 phenolic compounds and polyphenol oxidase. *Sciences des aliments*, 11, 49-62.
324
- 325 Bekhradi, F., Luna, M. C., Delshad, M., Jordan, M. J., Sotomayor, J. A., Martínez-Conesa, C., & Gil, M. I. (2015).
326 Effect of deficit irrigation on the postharvest quality of different genotypes of basil including purple and green
327 Iranian cultivars and a Genovese variety. *Postharvest Biology and Technology*, 100, 127-135.
328
- 329 Bel-Rhlid, R., Crespy, V., Pagé-Zoerkler, N., Nagy, K., Raab, T., & Hansen, C. H. (2009). Hydrolysis of
330 rosmarinic acid from rosemary extract with esterases and *Lactobacillus johnsonii* *in vitro* and in a gastrointestinal
331 model. *J. Agric. Food Chem.*, 57, 7700-7705.
332
- 333 Brand-Williams, W., Cuvelier, M.E. & Berset, C. (1995). Use of a free radical method to evaluate antioxidant
334 activity. *Food Sci. Technol-Leb.* 28, 25-30.
335
- 336 Cantwell, M.I. & Reid, M.S. (1993) Postharvest Physiology and Handling of Fresh Culinary Herbs. *Journal of*
337 *Herbs Spices & Medicinal Plants* 1, 93-125
338
- 339 Cefola M., Amodio M.L., Rinaldi R., Vanadia S. & Colelli, G. (2010). Exposure to 1-methylcyclopropene (1-MCP)
340 delays effects of ethylene on fresh-cut broccoli raab (*Brassica rapa* L.). *Postharvest Biology and Technology* 58,
341 29-35.
342
- 343 Cefola, M., Pace, B., Colelli, G. & Cantwell, M. (2015). Compositional and marketable quality of fresh-cut florets
344 of four specialty brassicas in relation to controlled atmosphere storage. *Acta Horticulturae (ISHS)* 1071, 455-462.

345

346 Cefola, M. & Pace, B. (2015). Application of oxalic acid to preserve the overall quality of rocket and baby spinach
347 leaves during storage. *Journal of Food Processing and Preservation*, 39, 2523-2532.

348

349 Chiang, L.C., Ng, L.T., Cheng, P.W., Chiang, W., & Lin, C.C. (2005). Antiviral activities of extracts and selected
350 pure constituents of *Ocimum basilicum*. *Clinical and Experimental Pharmacology and Physiology*, 32, 811-816.

351

352 Cook, N.C., Samman S. (1996). Flavonoids-Chemistry, metabolism, cardioprotective effects, and dietary sources.
353 *The Journal of Nutritional Biochemistry*, 7, 66-76.

354

355 Cozzolino, R., Pace, B., Cefola, M., Martignetti, A., Stocchero, M., Fratianni, F., Nazzaro, F., & De Giulio, B.
356 (2016). Assessment of volatile profile as potential marker of chilling injury of basil leaves during postharvest
357 storage. *Food Chemistry*, 213, 361-368.

358

359 Dogan, S., Turan, P., Dogan, M., Arslan, O., & Alkan, M. (2005). Purification and characterization of *Ocimum*
360 *basilicum* L. polyphenol oxidase *Journal of Agricultural and Food Chemistry*, 53, 10224-10230.

361

362 Dua, A., Agrawal, S., Singh, A., Mahajan, R. (2016). Antioxidant and Antimicrobial Potential of Polyphenols from
363 Foods. In: (N.Garg, S.M. Abdel-Aziz, A. Aeron eds) *Microbes in Food and Health*, Springer, pp 43-63.

364

365 Fratianni, F., Nazzaro, F., Marandino, A., Fusco, M.d.R., Coppola, R., De Feo, V., De Martino, L. (2013).
366 Biochemical composition, antimicrobial activities, and anti-quorum-sensing activities of ethanol and ethyl acetate
367 extracts from *Hypericum connatum* Lam. (*Guttiferae*) *Journal of Medicinal Food*, 16, 454-459.

368

369 Gonthier, M.P., Remesy, C., Scalbert, A., Cheynier, V., Souquet, J.M., Poutanen, K. & Aura, A.M. (2006).
370 Microbial metabolism of caffeic acid and its esters chlorogenic and caftaric acids by human faecal microbiota *in*
371 *vitro*. *Biomedicine and Pharmacotherapy*, 60, 536-540.

372

373 Hakkim, F.L., Gowri Shankar, C. & Girija S. (2007). Chemical composition and antioxidant property of holy basil
374 (*Ocimum sanctum* L.) leaves, stems, and inflorescence and their *in vitro* callus cultures *Journal of Agricultural and*
375 *Food Chemistry*, 55, 9109-9117.

376

377 Jayasinghe, C., Gotoh, N., Aoki, T., & Wada, S. (2003). Phenolics composition and antioxidant activity of sweet
378 basil (*Ocimum basilicum* L.). *Journal of Agricultural and Food Chemistry* 51, 4442-4449.

379

380 Kader, A.A. (1992). Postharvest biology and technology: An overview. In: A.A. Kader. Postharvest Technology of
381 Horticultural Crops. (2nd Ed.). University of California: ANR Publication, Oakland. CA.

382

383 Kader, A.A. (2002). Postharvest Technology of Horticultural Crops. (3rd Ed.). University of California: ANR
384 Publication, Oakland, CA.

385

386 Lado, J., Rodrigo, M.J., Cronje, P., & Zacarías, L. (2015). Involvement of lycopene in the induction of tolerance to
387 chilling injury in grapefruit. *Postharvest Biology and Technology*, 100, 176-186.

388

389 Lafuente, M. T., Zacarias, L., Martínez-Téllez, M. A., Sanchez-Ballesta, M. T. & Dupille, E. (2001). Phenylalanine
390 ammonia-lyase as related to ethylene in the development of chilling symptoms during cold storage of citrus fruits.
391 *Journal of Agricultural and Food Chemistry*, 49, 6020-6025.

392

393 Lattanzio, V. (2003). Bioactive polyphenols: their role in quality and storability of fruit and vegetables. *Journal of*
394 *Applied Botany*, 77(5/6), 128-146.

395

396 Lee, J. & Scagel C.F. (2009). Chicoric acid found in basil (*Ocimum basilicum* L.) leaves. *Food Chemistry* 115,
397 650-656.

398

399 Loughrin, J.H., & Kasperbauer, M.J. (2001). Light reflected from coloured mulches affects aroma and phenol
400 content of sweet basil (*Ocimum basilicum* L.) leaves. *Journal of Agricultural and Food Chemistry* 49, 1331-1335.

401

402 Megías, Z., Martínez, C., Manzano, S., García, A., del Mar Reboloso-Fuentes, M., Valenzuela, J. L. & Jamilena,
403 M. (2016). Ethylene biosynthesis and signaling elements involved in chilling injury and other postharvest quality
404 traits in the non-climacteric fruit of zucchini (*Cucurbita pepo*). *Postharvest Biology and Technology*, 113, 48-57.
405

406 Saltveit, M.E. & Morris, L.L. (2000) Wound induced changes in phenolic metabolism and tissue browning are
407 altered by heat shock. *Postharvest Biology and Technology* 21, 61-69.
408

409 Shiga, T., Shoji, K., Shimada, H., Hashida, S., Goto, F. & Yoshihara, T. (2009). Effect of light quality on
410 rosmarinic acid content and antioxidant activity of sweet basil, *Ocimum basilicum* L. *Plant Biotechnology* 26, 255-
411 259.
412

413 Singleton, V.L. & Rossi, J.A. (1965). Colorimetry of total phenolics with a phosphomolybdic-phosphotungstic acid
414 reagents. *American Journal of Enology and Viticulture*, 16, 144-158.
415

416 Sulaiman, S.F., Sajak, A.A.B., Ooi, K.L., Supriatno, E. & Seow M. (2011). Effect of solvents in extracting
417 polyphenols and antioxidants of selected raw vegetables. *Journal of Food Composition and Analysis* 24, 506-515.
418

419 Tsai, Y.L., Chiou, S.Y., Chi Chan, K.C., Sung, J.M. & Lin, S.D. (2012). Caffeic acid derivatives, total phenols,
420 antioxidant and antimutagenic activities of *Echinacea purpurea* flower extracts. *LWT - Food Science and*
421 *Technology* 46 169-176.
422

423 Tudela, J.A., Marín, A., Garrido, Y., Cantwell, M., Medina-Martínez, M.S. & Gil, M.I. (2013). Off-odour
424 development in modified atmosphere packaged baby spinach is an unresolved problem. *Postharvest Biology and*
425 *Technology*, 75, 75-85.
426

427 Wongsheree, T., Ketsa, S., & van Doorn, W.G. (2009). The relationship between chilling injury and membrane
428 damage in lemon basil (*Ocimum × citriodourum*) leaves. *Postharvest Biology and Technology*, 51, 91-96.
429

430 **Figure Captions**

431 Figure 1. Effect of storage at chilling (4 °C) or non-chilling temperatures (12 °C) on respiration rate, ethylene
432 production and ammonium content of basil leaves belonging to cultivar *Italico a foglia larga* (A, B, C), *Cammeo* (D, E,
433 F) and *Italiano classico* (G, H, I).

434

435 Figure 2. Effect of storage at chilling (4 °C) or non-chilling temperatures (12 °C) on antioxidant activity and total
436 phenol content of basil leaves belonging to cultivar *Italico a foglia larga* (A, D), *Cammeo* (B, E) and *Italiano classico*
437 (C, F).

438

439

440

Table 1

Table 1. Effects of temperature (at 4 or 12 °C), storage (3, 6 and 9 days) and their interaction on sensory visual quality, physiological and biochemical parameters of the leaves of *Italico a foglia larga*, *Cammeo* and *Italiano Classico* basil cultivars.

Parameters	Experiment 1: <i>Italico a foglia larga</i>			Experiment 2: <i>Cammeo</i>			Experiment 3: <i>Italiano Classico</i>		
	Temperature (A)	Storage (B)	A x B	Temperature (A)	Storage (B)	A x B	Temperature (A)	Storage (B)	A x B
	(4 or 12 °C)	(3, 6 and 9 days)		(4 or 12 °C)	(3, 6 and 9 days)		(4 or 12 °C)	(3, 6 and 9 days)	
Visual Quality score (5-1)	**	***	ns	***	***	ns	**	***	ns
Respiration rate (mL CO ₂ kg ⁻¹ h ⁻¹)	ns	*	ns	*	**	**	***	ns	ns
Ethylene production (μL Kg ⁻¹ h ⁻¹)	ns	**	ns	***	***	**	***	***	***
Ammonium content (μmole NH ₄ ⁺ g ⁻¹ fw)	ns	ns	*	***	***	***	***	***	**
Antioxidant activity (EC ₅₀ , mg)	ns	ns	ns	***	***	***	***	***	***
Total Phenols (mg of gallic acid equivalent GAE g ⁻¹)	ns	ns	ns	***	*	***	*	***	***

ns: not significant; *for $P \leq 0.05$; ** ≤ 0.01 ; *** ≤ 0.001 . For storage temperature dataset of 6 samples (2 replicates x 3 storage durations), for storage duration dataset of 4 samples (2 replicates x 2 storage temperatures) were used. Fw: fresh weight.

Table 2

Table 2. Effect of temperature (4 or 12 °C) and storage (3, 6 and 9 days) on visual quality score (5 = excellent, fresh appearance, 4 = good, 3 = fair, limit of marketability, 2 = poor, just below the limit of marketability, 1 = very bad, inedible) of basil leaves from *Italico a foglia larga*, *Cammeo* and *Italiano Classico* cultivars

Cultivar name	Temperature (°C)		Significance	Storage (days)			Significance
	4	12		3	6	9	
<i>Italico a foglia larga</i>	2.8 b	3.6 a	**	4.1 a	3.2 b	2.2 c	***
<i>Cammeo</i>	2.5 b	3.9 a	***	4.4 a	3.5 a	1.8 b	***
<i>Italiano Classico</i>	2.4 b	3.4 a	**	4.2 a	2.6 b	1.8 b	***

For each cultivar and for each factor (temperature or storage time) different letters indicates mean significantly different for the following levels: **($P \leq 0.01$), *** ($P \leq 0.001$). For temperature mean values of 6 data (2 replicates x 3 storage times, after 3, 6, 9 days); for each storage time mean values of 4 data (2 replicates x 2 temperature at 4 and 12 °C). Method: 95.0 percent Student–Newman–Keuls (SNK).

Table 3

Table 3. Phenolic compounds of basil leaves from *Italico a foglia larga* (Experiment 1), *Cammeo* (Experiment 2) and *Italiano Classico* (Experiment 3) cultivars identified by UPLC in the fresh samples (just after detachment, at 0 days, d), and in leaves stored (for 3, 6 and 9 d) at chilling (4 °C) or no chilling temperature (12 °C)

Phenolic compounds (µg/g)	STORAGE CONDITION							P
	0d	3d-12°C	3d-4 °C	6d-12°C	6d-4°C	9d-12°C	9d-4°C	
Experiment 1: <i>Italico a foglia larga</i>								
gallic acid	117.27 ^a	25.26 ^b	114.57 ^a	23.42 ^b	49.23 ^b	117.16 ^a	35.55 ^b	***
caftaric acid	0.00 ^b	0.00 ^b	0.00 ^b	0.00 ^b	0.00 ^b	7.95 ^a	0.00 ^b	**
chicoric acid	0.00 ^d	0.00 ^d	0.00 ^d	0.00 ^d	16.03 ^c	147.18 ^a	99.10 ^b	****
cholorogenic acid	52.93 ^d	167.89 ^b	177.87 ^b	106.30 ^c	107.56 ^c	229.45 ^a	154.48 ^b	***
catechin	19.55 ^a	0.93 ^{bc}	1.28 ^{bc}	1.04 ^{bc}	2.07 ^b	0.00 ^c	0.00 ^c	****
caffeic acid	285.04 ^a	129.66 ^d	168.03 ^{bc}	87.16 ^{de}	119.38 ^{de}	200.82 ^b	136.81 ^{cd}	****
rosmarinic acid	1508 ^a	895 ^b	856 ^{bc}	483 ^d	717 ^c	812 ^b	732 ^{bc}	****
apigenin	6.66 ^{ab}	7.07 ^{ab}	7.44 ^{ab}	6.47 ^{ab}	4.07 ^b	8.15 ^a	9.64 ^a	*
quercetin	0.00 ^c	8.31 ^b	12.41 ^b	7.31 ^b	22.22 ^a	7.12 ^b	0.00 ^c	***
Experiment 2: <i>Cammeo</i>								
gallic acid	37.17 ^{bc}	57.11 ^b	91.50 ^a	58.91 ^b	32.50 ^c	59.00 ^b	41.64 ^{bc}	**
caftaric acid	24.46 ^b	0.00 ^d	0.00 ^d	9.29 ^c	0.00 ^d	52.84 ^a	0.00 ^d	****
chicoric acid	0.00 ^e	27.13 ^d	40.66 ^c	108.77 ^o	0.00 ^e	83.39 ^b	0.00 ^e	****
cholorogenic acid	0.00 ^b	10.51 ^b	0.00 ^b	4.62 ^b	4.06 ^b	36.35 ^a	0.00 ^b	**
caffeic acid	508.7 ^a	49.8 ^d	98.9 ^d	259.5 ^c	84.9 ^d	388.4 ^b	91.3 ^d	****
rosmarinic acid	1405 ^a	1094 ^b	511 ^c	1451 ^a	484 ^{cd}	1321 ^{ab}	206 ^d	****
apigenin	4.98 ^{cd}	8.56 ^{cd}	0.00 ^d	15.92 ^{abc}	11.21 ^{bc}	27.01 ^a	20.58 ^{ab}	**
quercetin	0.00 ^b	0.00 ^b	0.00 ^b	0.00 ^b	0.00 ^b	2.65 ^a	0.00 ^b	*
Experiment 3: <i>Italiano Classico</i>								
gallic acid	46.16 ^c	86.82 ^a	41.78 ^c	29.17 ^d	63.41 ^b	48.65 ^c	51.35 ^c	***
caftaric acid	16.08 ^c	7.99 ^d	36.15 ^a	27.71 ^b	0.00 ^e	29.77 ^b	0.00 ^e	****

chlorogenic acid	44.36 ^c	36.60 ^c	79.43 ^b	90.44 ^b	80.94 ^b	138.70 ^a	21.44 ^c	***
catechin	0.00 ^b	48.65 ^a	0.00 ^b	0.00 ^b	0.00 ^b	0.00 ^b	0.00 ^b	****
caffeic acid	547 ^a	241 ^e	455 ^{bc}	407 ^{cd}	313 ^{de}	541 ^b	318 ^{de}	**
rosmarinic acid	1672 ^{ab}	1692 ^a	1593 ^{ab}	811 ^d	485 ^e	1365 ^{bc}	1180 ^c	****
apigenin	42.00 ^b	42.76 ^b	54.88 ^b	37.12 ^b	46.74 ^b	230.50 ^a	45.46 ^b	****
rutin	0.00 ^d	81.14 ^b	0.00 ^d	42.03 ^c	96.85 ^a	0.00 ^d	0.00 ^d	****

For each experiment and for each compound identified, mean values (n=2) followed by different letters (a-e) are significantly different for $P \leq 0.05$.

*for $P \leq 0.05$; ** ≤ 0.01 ; *** ≤ 0.001 ; **** ≤ 0.0001 . Method: 95.0 percent Student–Newman–Keuls (SNK).

Figure 1
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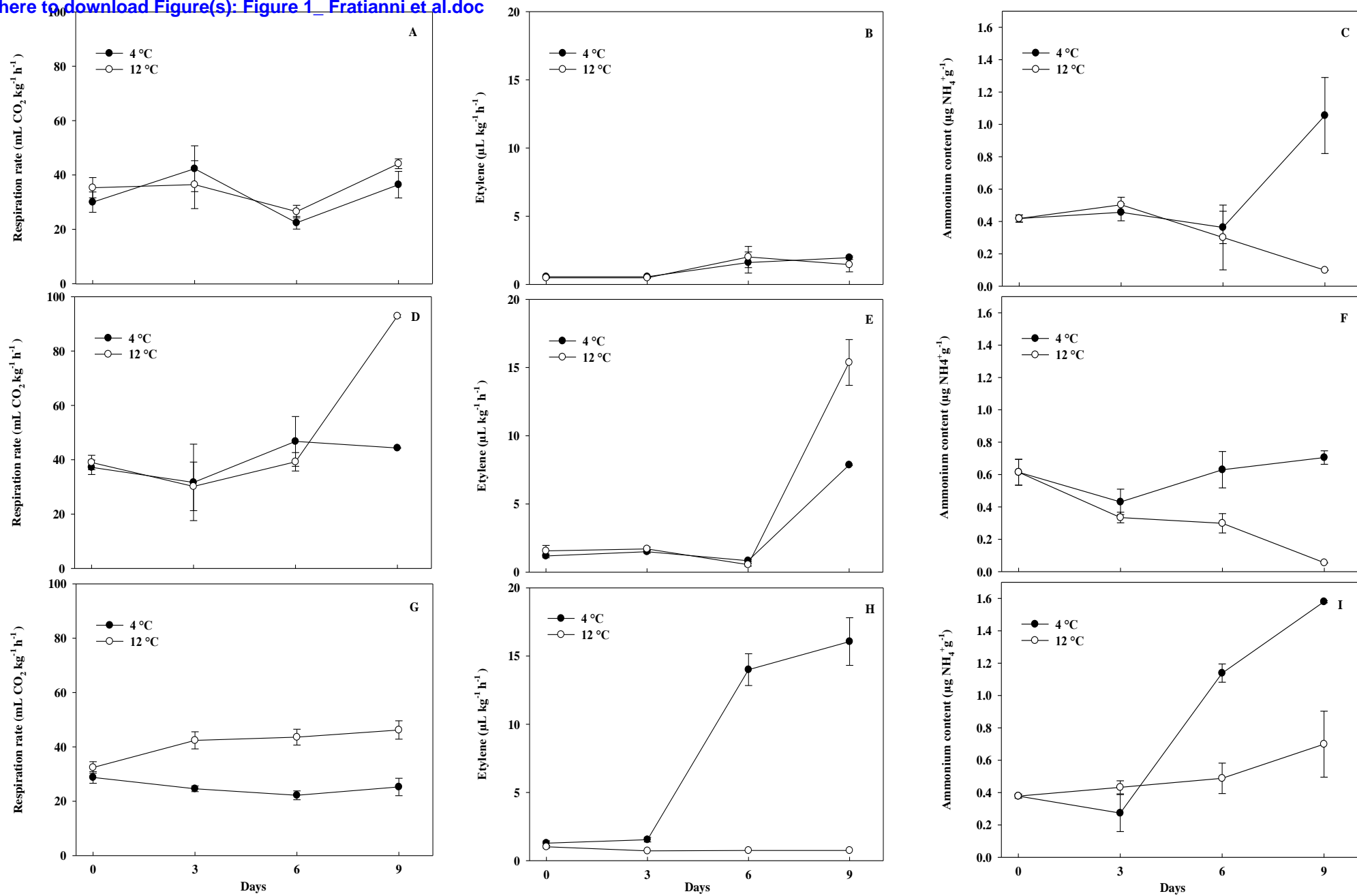


Figure 1

Figure 2

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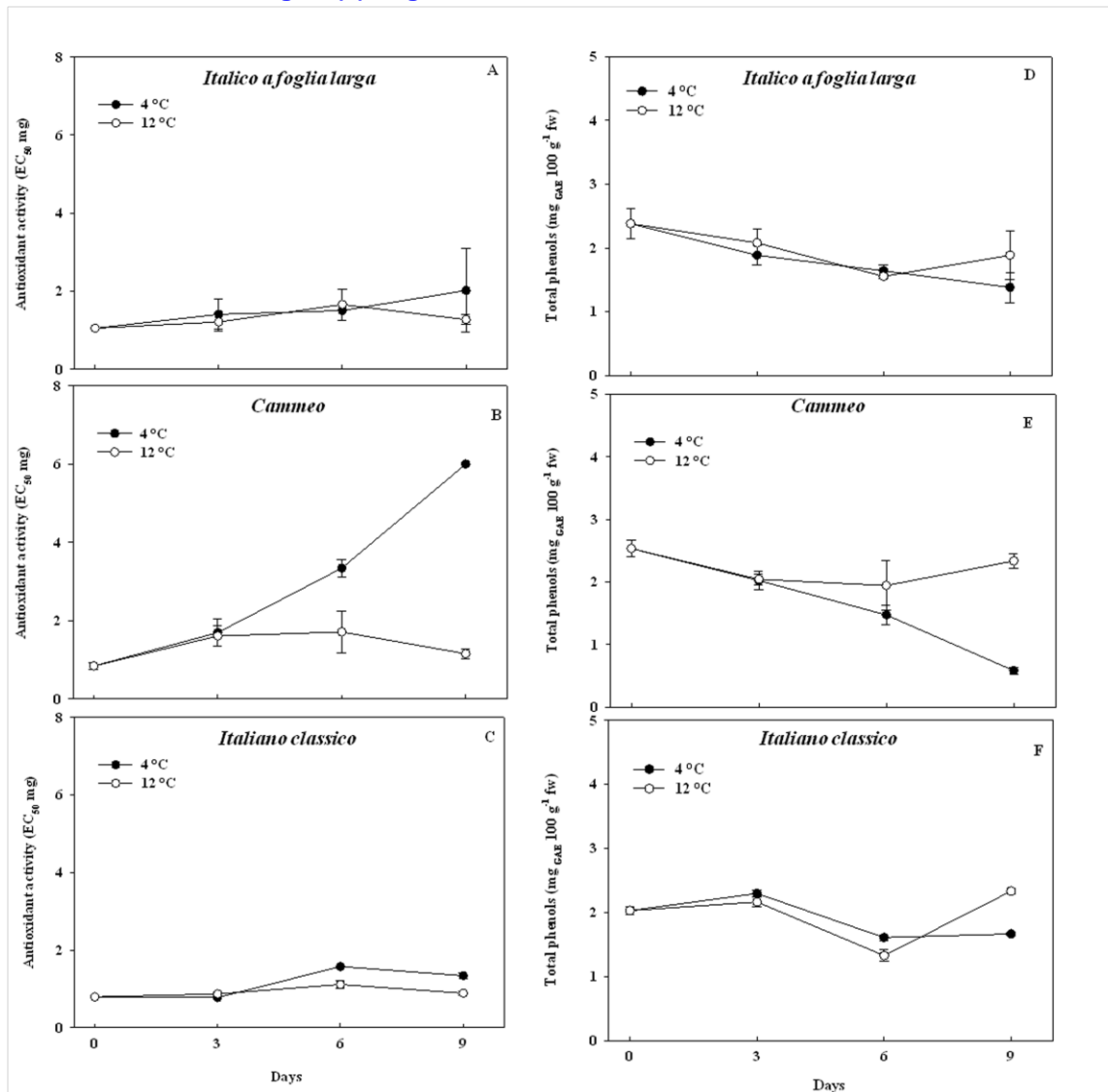


Figure 2